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(39) Title: G PROTEIN COUPLED RECEPTOR PROTEIN, PRODUCTION, AND USE THEREOF

DNA primers effective in screening G protein coupled receptor protein-encoding DNA fragments are provided. The primers which are complementary to nucleotide sequences that are in community with (homologous to) the nucleotide sequences encoding amino acid sequences corresponding to or near the first membrane-spanning domain or the sixth membrane-spanning domain each of known vertebrate G protein coupled receptor proteins were designed and synthesized. Methods of amplifying G protein coupled receptor protein-encoding DNAs using the above DNA primers, and novel targets G protein coupled receptor protein-encoding DNAs are also provided. Screening of cDNA libraries can be efficiently carried out. Human pituitary gland or amygdala-derived and mouse pancreas-derived G protein coupled receptor cDNA clones can be easily screened, partial peptides thereof, DNAs coding for the above G protein coupled receptor proteins, processes for producing the above G protein coupled receptor protein, methods of determining ligands for the above G protein coupled receptor protein, and other G protein coupled receptor protein, methods of determining ligands for the above G protein coupled receptor proteins or screening the binding of compounds or salts thereof obtained by the above screening method and the G protein coupled receptor proteins or screening the binding of compounds or salts thereof inhibited by the above screening method or the screening kit, pharmaceutical compositions containing the above compounds or salts thereof, and antibodies against the above protein coupled receptor proteins or partial peptides thereof are provided.

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DESCRIPTION

G PROTEIN COUPLED RECEPTOR PROTEIN,
PRODUCTION, AND USE THEREOFFIELD OF THE INVENTION

5 The present invention relates to novel DNAs which are useful as DNA primers for a polymerase chain reaction (PCR); methods for amplifying DNAs each coding for a G protein coupled receptor protein via PCR techniques using said DNA; screening methods for DNAs each encoding a G protein coupled receptor protein via PCR techniques using said DNA;

10 G protein coupled receptor protein-encoding DNAs obtained by said screening method; G protein coupled receptor proteins which are encoded by the DNA obtained via said screening method, peptide fragments or segments thereof, and modified peptide derivatives thereof; etc.

15 The present invention also relates to novel G protein coupled receptor proteins; novel G protein coupled receptor protein-encoding DNAs; processes for producing said G protein coupled receptor protein; use of said receptor protein and said protein-encoding DNA; etc.

20 The present invention also relates to novel human amygdaloid nucleus-derived G protein coupled receptor proteins; novel DNAs each coding for said G protein coupled receptor protein; processes for producing said G protein coupled receptor protein; use of said receptor protein and said protein-encoding DNA; etc.

25 The present invention also relates to novel mouse pancreatic β cell line MIN6-derived G protein coupled receptor proteins; novel DNAs each coding for said G protein coupled receptor protein; processes for producing said G protein coupled receptor protein; use of said receptor protein and

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said protein-encoding DNA; etc. Further, the present invention relates to novel human-derived G protein coupled receptor proteins (human prinoceptors); novel DNAs each coding for said G protein coupled receptor protein; processes for producing said G protein coupled receptor protein; use of said receptor protein and said protein-encoding DNA; etc.

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BACKGROUND OF THE INVENTION

A variety of hormones, neurotransmitters and the like control, regulate or adjust the functions of living bodies via specific receptors located in cell membranes. Many of these receptors mediate the transmission of intracellular signals via activation of guanine nucleotide-binding proteins (hereinafter, sometimes referred to as G proteins) with which the receptor is coupled and possess the common (homologous) structure, i.e. seven transmembranes (membrane-spanning regions (domains)). Therefore, such receptors are generically referred to as G protein coupled receptors or seven transmembrane (membrane-spanning) receptors.

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G protein coupled receptor proteins have a very important role as targets for molecules such as hormones, neurotransmitters and physiologically active substances, which molecules control, regulate or adjust the functions of living bodies. Each molecule has its own receptor protein which is specific thereto, whereby the specificities of individual physiologically active substances, including specific target cells and organs, specific pharmacological actions, specific action strength, action time, etc., are decided. Accordingly, it has been believed that, if G protein coupled receptor genes or cDNA can be cloned, those will be helpful not only for the clarification of structure, function, physiological action, etc. of the G protein coupled receptor but also for the development of pharmaceuticals by investigating the substances which act on the receptor. Until now, only several G protein coupled receptor genes or cDNAs have been cloned but it is believed that there are many unknown G protein coupled receptor genes which have not been recognized yet.

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The characteristic feature of the G protein coupled receptor proteins which have been known up to now is that seven clusters of hydrophobic amino acid residues are located in the primary structure and pass through (span) the cell membrane at each region thereof. It has been known that such a structure is common among all of the known G protein coupled receptor proteins and further that the amino acid sequences corresponding to the area where the protein passes through the membrane (membrane-spanning region or transmembrane region) and the amino acid sequences near the membrane-spanning region are often highly conserved among the receptors.

When an unknown protein has such a structure, it is strongly suggested that said protein is within a category of the G protein coupled receptor proteins. In addition, some amino acid residue alignments are common (homologous) and, by taking it as a characteristic feature, it is further strongly suggested that said protein is a G protein coupled receptor protein.

Libert, F, et al. (Science, 244:569-571; 1989) reported a method for cloning novel receptor genes by means of a polymerase chain reaction (hereinafter, sometimes referred to as PCR or a PCR technique) for a synthetic DNA primer which was synthesized based upon the information of common amino acid sequences obtained from a comparison among known G protein coupled receptor proteins. Libert, F. et al. used a pair of synthetic DNA primers corresponding to the portions of the third and the sixth membrane-spanning regions. However, in general, the design of primers used for the PCR regulates the molecular species of DNAs which are to be amplified. In addition, when a similarity (homology) in the amino acid sequence level is used as a basis, the use of different codons affects on the binding (hybridization) of the primer thereby resulting in a decrease in the amplifying efficiency.

Accordingly, although various novel receptor protein DNAs have been obtained using said DNA primers, it is not possible to succeed in amplifying DNAs for all receptor proteins in the prior art.

Further, the amino acid sequence which is common to from the first to the seventh membrane-spanning regions among 74 G protein coupled receptor proteins was reported by William C. Probst, et al. (DNA and Cell Biology, Vol. 11, No. 1, 1992, pp. 1-20). In this report, however, there is no suggestion for a method in which DNA coding for a novel G protein coupled receptor protein is screened by means of PCR using DNA primers which are complementary to the DNA coding for those amino acid sequences.

It would be desirable to develop DNA primers for PCR techniques which allow selective and efficient screenings of DNAs coding for the areas (regions) more nearer the full length of novel G protein coupled receptor proteins by utilizing the common (homologous) sequence(s) of the G protein coupled receptor protein or the DNA coding therefor.

It would also be desirable to develop synthetic DNA primers corresponding to the portions of the third and the sixth membrane-spanning regions, said primer being useful in screening for DNA coding for G protein coupled receptor proteins in more selective and efficient manner as compared with a series of the synthetic DNA primers corresponding to the sequences of the third to the sixth membrane-spanning regions as reported by Libert, F. et al.

G protein coupled receptor proteins are important for investigating substances which control the function of living organisms and proceeding developments thereof as pharmaceuticals. Finding and development of candidate compounds for new pharmaceuticals can be efficiently proceeded by using G protein coupled receptor proteins and by conducting receptor binding experiments and evaluating experiments on agonists/antagonists using intracellular information transmittance systems as indexes. Especially when the presence of a novel G protein coupled receptor protein can be clarified, the presence of a substance having a specific action thereon can be suggested.

If a novel DNA which codes for a novel G protein coupled receptor protein can be efficiently screened and

isolated, it will now be possible to proceed with the isolation of DNA having an entire coding region, the construction of an expression system therefor and the screening of an acting ligand.

A hypothalamo-hypophysial system is one of the passages for controlling, regulating or adjusting the functions of organisms relying upon interactions of hormones and neurotransmitters with G protein coupled receptors. In the hypothalamo-hypophysial system, the secretion of pituitary hormones from the pituitary body (hypophysis) is regulated by hypothalamic hormones (hypophysiotropic releasing factors), and the functions of target cells and organs are controlled by pituitary hormones released into the blood. Functions which are important for the living body are regulated through this system, such as maintenance of homeostasis and control of development and growth of a genital system and an individual organism. Representative examples of the hypothalamic hormones include TRH, LH-RH, CRF, GRF, somatostatin, galanin, etc. Representative examples of the pituitary hormones include TSH, ACTH, FSH, LH, prolactin, growth hormone, oxytocin, vasopressin, etc. In particular, the secretion of pituitary hormones is regulated according to a positive feedback mechanism or a negative feedback mechanism relied on the hypothalamic hormones and peripheral hormones secreted from the target endocrine glands. A variety of receptor proteins present in the pituitary gland play a major role for regulating the hypothalamo-hypophysial system.

It has been widely known that these hormones, factors and receptors are widely distributed in the brain instead of existing only locally in the hypothalamo-hypophysial system. This fact suggests that the substances which are called "hypothalamic hormones" are working as neurotransmitters or neuroregulators in the central nervous system. It is further considered that these substances are similarly distributed even in the peripheral tissues to play the role of important functions. The pancreas plays an important role of carrying out the carbohydrate metabolism by secreting not only

a digestive fluid but also glucagon and insulin. Insulin is secreted from the β cells and its secretion is promoted chiefly by glucose. It has, however, been known that a variety of receptors exist in the β cells, and the secretion of insulin is controlled by various factors such as peptide hormones (galanin, somatostatin, gastric inhibitory polypeptide, glucagon, amylin, etc.), sugars (mannose, etc.), amino acids, and neurotransmitters in addition to glucose.

It has thus been known that in the pituitary gland and in the pancreas are present receptor proteins for many hormones and neurotransmitters, said receptor proteins playing important roles for regulating the functions. As for the galanin and amylin, however, there has not yet been reported any discovery concerning the structure of their receptor protein cDNAs. It is not known whether there exist any unknown receptor proteins or receptor protein subtypes.

For substances regulating the functions of the pituitary gland and pancreas, there exist receptor proteins specific to said substance on the surfaces of various functional cells of the pituitary gland and pancreas. The pituitary gland and the pancreas are associations of a plurality of functional cells, and the actions of the individual substances are defined by the distributions of their target receptor proteins among the functional cells.

Accordingly, a substance, in many cases, exhibits an extensive variety of actions. To comprehend such complex systems, it is necessary to clarify the relations between the acting substances and the specific receptor proteins. It is further necessary to efficiently screen for receptor protein agonists and antagonists capable of regulating the pituitary gland and pancreas, to clarify the structures of genes of receptor proteins from the standpoint of investigating and developing pharmaceuticals, and further to express them in a suitable expression system.

By utilizing the fact that a G protein coupled receptor protein exhibits homology in part of the structure thereof at the amino acid sequence level, an experiment of

looking at DNAs coding for novel receptor proteins relying upon a polymerase chain reaction (hereinafter simply referred to as "PCR") has recently been made.

In the central nervous system, many receptor proteins such as dopamine receptor protein, LH-RH receptor protein, neurotensin receptor protein, opioid receptor protein, CRF receptor protein, CRF receptor protein, somatostatin receptor protein, galanin receptor protein, TRH receptor protein, etc. are G protein coupled receptor proteins, and it has been clarified that ligands to these receptors exert a variety of effects in the central nervous system.

In the immune system, an α - or a β -chemokine receptor protein, an MIP1 α receptor protein, an IL-8 receptor protein, a C5a receptor protein, etc. have been known as such G protein coupled receptor proteins, and are working as receptor proteins responsive to immunoregulating substances to play important roles for regulating the functions of the living body. There is, for example, an IL-6 receptor protein that acts both in the above-mentioned central nervous system and in the immune system. IL-6 is both a β -cell differentiating factor and a biologically active factor related to the proliferation and differentiation of nerve cells.

It has been widely known that these hormones, factors and receptor proteins are usually widely distributed up to the peripheral tissues instead of existing only locally in the central nervous system and in the immune system and are producing important functions, respectively. Agonists and antagonists for these receptor proteins are now being developed as various useful pharmaceuticals.

For substances regulating the functions of the central nervous system and the immune system, there exist receptor proteins specific to said substance on the surfaces of various functional cells of the central nervous system and the immune system. The central nervous system and the immune system are associations of a plurality of functional cells, and the actions of the individual substances are defined by the distributions of their target receptor proteins among the

functional cells. Accordingly, a substance, in many cases, exhibits an extensive variety of actions. Moreover, there is an example wherein many factors play a part in a physiological phenomenon. To comprehend such complex systems, it is necessary to clarify relations between the acting substances and the specific receptor proteins.

As discussed herein above, the G protein coupled receptor protein is present on the cell surface of living body cells and organs and has a very important role as a target for molecules such as hormones, neurotransmitters and physiologically active substances, which molecules control, regulate or adjust the functions of living body cells and organs.

SUMMARY OF THE INVENTION

One object of the present invention is to provide novel DNAs which are useful as DNA primers for a polymerase chain reaction; methods for amplifying a DNA coding for a G protein coupled receptor protein using said DNA; screening methods for the DNA coding for a G protein coupled receptor protein using said DNA; DNAs obtained by said screening method; and G protein coupled receptor proteins encoded by the DNA obtained by said screening method, peptide fragments or segments thereof, modified peptide derivatives thereof or salts thereof.

Another object of the present invention is to provide processes for producing said receptor protein; transformants capable of expressing said receptor protein; cell membrane fractions obtained from said transformant; methods for determining a ligand to the receptor protein; screening methods for a compound or a salt thereof capable of inhibiting the binding of the ligand with the receptor protein; kits for said screening method, pharmaceutical compositions comprising an effective amount of the inhibitory compound; antibodies against said receptor protein; immunoassays using said receptor protein or said antibody and use of said receptor protein and encoding DNA.

Yet another object of the present invention is to provide novel G protein coupled receptor proteins which are expressed in pituitary glands or pancreatic β cells; DNAs comprising a DNA coding for said G protein coupled receptor protein; processes for producing said receptor protein; transformants capable of expressing said receptor protein; cell membrane fractions obtained from said transformant; methods for determining a ligand to the receptor protein; screening methods for a compound or a salt thereof capable of inhibiting the binding of the ligand with the receptor protein; kits for said screening method, pharmaceutical compositions comprising the inhibitory compound; antibodies against said receptor protein; immunoassays using said receptor protein or said antibody and use of said receptor protein and encoding DNA.

Still another object of the present invention is to provide novel human amygdaloid nucleus-derived G protein coupled receptor proteins; DNAs comprising a DNA coding for said G protein coupled receptor protein; processes for producing said receptor protein; transformants capable of expressing said receptor protein; cell membrane fractions obtained from said transformant; methods for determining a ligand to the receptor protein; screening methods for a compound or a salt thereof capable of inhibiting the binding of the ligand with the receptor protein; kits for said screening method, pharmaceutical compositions comprising the inhibitory compound; antibodies against said receptor protein; immunoassays using said receptor protein or said antibody and use of said receptor protein and encoding DNA.

Yet another object of the present invention is to provide novel mouse pancreatic β cell line MIN6-derived G protein coupled receptor proteins; DNAs comprising a DNA coding for said G protein coupled receptor protein; processes for producing said receptor protein; transformants capable of expressing said receptor protein; cell membrane fractions obtained from said transformant; methods for determining a ligand to the receptor protein; screening methods for a

compound or a salt thereof capable of inhibiting the binding of the ligand with the receptor protein; kits for said screening method, pharmaceutical compositions comprising the inhibitory compound; antibodies against said receptor protein; immunoassays using said receptor protein or said antibody and use of said receptor protein and encoding DNA.

The present inventors have succeeded in synthesizing novel DNA primers based upon the similarity (homology) with the base sequences coding for the first membrane-spanning region or the sixth membrane-spanning region each of known G protein coupled receptor proteins. It is to be particularly noted that there has been no report of a DNA primer pair which has been synthesized paying attention to the similarity with the base sequence coding for the first and the sixth membrane-spanning region of the known G protein coupled receptor protein.

Next the present inventors have succeeded in synthesizing other novel DNA primers based upon the similarity (homology) with the base sequences coding for the third or the sixth membrane-spanning region each of known G protein coupled receptor proteins. They have also unexpectedly succeeded in efficiently amplifying DNAs (DNA fragments) coding for G protein coupled receptor proteins by means of PCR using those DNA primers.

They have further succeeded in synthesizing novel DNA primers based upon the similarity (homology) with the base sequences coding for the second or the seventh membrane-spanning region each of known G protein coupled receptor proteins; upon the similarity (homology) with the base sequences coding for first or the third membrane-spanning region each of known G protein coupled receptor proteins; and upon the similarity (homology) with the base sequences coding for the second or the sixth membrane-spanning region each of known G protein coupled receptor proteins. They have furthermore and unexpectedly succeeded in efficiently amplifying DNAs (DNA fragments) coding for G protein coupled receptor proteins by conducting PCR using those DNA primers.

Moreover, the present inventors have succeeded in

efficiently cloning full-length DNA coding for said G protein coupled receptor protein via using amplified DNAs (DNA fragments) coding for said G protein coupled receptor protein. Thus, they have found that novel DNA coding for novel G protein coupled receptor proteins can be isolated, characterized or prepared via conducting amplifications and analyses of various DNA using said DNA primers.

To be more specific, the present inventors have selected amino acid sequences which are each common to the portion corresponding to or near the first and the sixth membrane-spanning region of the known individual G protein coupled receptor proteins and have designed the DNA primer (SEQ ID NO: 1) coding for the amino acid sequence common (homologous) to the first membrane-spanning region and the DNA primer (SEQ ID NO: 2) which is complementary to the nucleotide sequence coding for the amino acid sequence common (homologous) to the area near the sixth membrane-spanning region.

Those DNA primers have a different nucleotide sequence as compared with reported DNA primers (e.g. a set of synthetic DNA primers corresponding to the third and the sixth membrane-spanning regions (SEQ ID NO: 60 and SEQ ID NO: 61) as reported by Libert, F. et al.) and such instant primers are novel and unique.

Especially for an object of conducting an efficient elongation reaction in the PCR, the 3'-terminal region of the instant primers contains the nucleotide sequence which is common (homologous) among many receptor proteins. Even in other areas, the similarity (homology) at the nucleotide sequence level (base sequence level) is utilized for setting the mixed base (nucleotide) parts wherein their nucleotide sequences (base sequences) are matched for as many nucleotides (bases) as possible among many DNA for the receptor proteins. Then the present inventors have amplified cDNA derived from human brain amygdala, human pituitary gland and rat brain, found the amplified products as shown in Figure 17 and, from those products, obtained the G protein coupled receptor protein cDNAs having the sequence as shown in

Figure 18, Figure 19, Figure 20, Figure 21, Figure 22, Figure 23, Figure 27, Figure 29, Figure 34, Figure 37, Figure 40, Figure 43 or Figure 46. Among them, the G protein coupled receptor protein cDNAs having the sequence as shown in Figure 22, Figure 23, Figure 27, Figure 29, Figure 34, Figure 37, Figure 40, Figure 43 or Figure 46 are novel.

Further, the present inventors have selected the amino acid sequences common (homologous) to the third and the sixth membrane-spanning region each of the known G protein coupled receptor proteins and designed the DNA primers coding for the amino acid sequence common (homologous) to the third membrane-spanning region (SEQ ID NO: 3; SEQ ID NO: 5, SEQ ID NO: 6 and SEQ ID NO: 7) and the DNA primers which are complementary to the nucleotide sequence coding for the amino acid sequence common (homologous) to the portion near the sixth membrane-spanning region (SEQ ID NO: 4, SEQ ID NO: 8 and SEQ ID NO: 9). Again, those DNA primers have different base sequences from those of the DNA primers previously reported (e.g., a set of synthetic DNA primers corresponding to the sequence of the third and the sixth membrane-spanning regions (SEQ ID NO: 60 and SEQ ID NO: 61) as reported by Libert, F. et al.) and such instant primers are novel and unique. The present inventors amplified cDNA derived from the smooth muscles of gastric pylorus of rabbits using said DNA primer and obtained G protein coupled receptor protein cDNA having the sequence of Figure 49 or Figure 52. Those cDNAs are novel.

Still further, the present inventors have selected the amino acid sequences common (homologous) to the second and the seventh membrane-spanning region each of the known G protein coupled receptor proteins and designed the DNA primer coding for the amino acid sequence common (homologous) to the second membrane-spanning region (SEQ ID NO: 10) and the DNA primer which is complementary to the base sequence coding for the amino acid sequence common (homologous) to the portions near the seventh membrane-spanning region (SEQ ID NO: 11). Those DNA primers have different base sequences from those of DNA primers previously reported (e.g., a set of synthetic DNA

primers corresponding to the part of the third and the sixth membrane-spanning regions (SEQ ID NO: 60 and SEQ ID NO: 61) as reported by Libert, F. et al) and such instant primers are novel and unique. The present inventors amplified cDNA derived from the smooth muscles of gastric pylorus of rabbits using said DNA primer and obtained G protein coupled receptor protein cDNAs having each the sequence of Figure 55, Figure 56, Figure 72, or Figure 73. Those cDNAs are novel.

Furthermore, the present inventors have selected the amino acid sequences common (homologous) to the first and the third membrane-spanning region each of the known G protein coupled receptor proteins and designed the DNA primer coding for the amino acid sequence common (homologous) to the first membrane-spanning region (SEQ ID NO: 12) and the DNA primer which is complementary to the base sequence coding for the amino acid sequence common (homologous) to the portions near the third membrane-spanning region (SEQ ID NO: 13).

Still further, the present inventors have selected the amino acid sequences common (homologous) to the third and the sixth membrane-spanning region each of the known G protein coupled receptor proteins and designed the DNA primers coding for the amino acid sequence common (homologous) to the third membrane-spanning region (SEQ ID NO: 10 and SEQ ID NO: 18) and the DNA primers which are complementary to the base sequence coding for the amino acid sequence common (homologous) to the parts near the sixth membrane-spanning region (SEQ ID NO: 15 and SEQ ID NO: 19). Further, the present inventors have selected the amino acid sequences common (homologous) to the second and the sixth membrane-spanning region each of the known G protein coupled receptor proteins and designed the DNA primer coding for the amino acid sequence common (homologous) to the second membrane-spanning region (SEQ ID NO: 16) and the DNA primer which is complementary to the base sequence coding for the amino acid sequence common (homologous) to the parts near the sixth membrane-spanning region (SEQ ID NO: 17). Those DNA primers have different base sequences from those of DNA primers previously reported (e.g., a set of synthetic DNA primers

corresponding to the part of the third and the sixth membrane-spanning regions (SEQ ID NO: 60 and SEQ ID NO: 61) as reported by Libert, F. et al) and such instant primers are novel and unique.

Still another object of the present invention is to provide a G protein coupled receptor protein expressed in the pituitary gland and pancreatic β cells, a DNA comprising a DNA coding for said protein, a process for producing said protein, and use of said protein and DNA.

In order to achieve the above-mentioned aims, the present inventors have made extensive investigations.

As a result, the present inventors have succeeded in amplifying cDNA derived from the human pituitary gland and the mouse pancreatic β -cell strain, MIN 6, with a synthetic DNA primer for efficiently isolating G protein coupled receptor protein-encoding DNA, and have forwarded the analysis. Thus, the present inventors have succeeded in isolating novel human and mouse-derived G protein coupled receptor protein-encoding cDNAs, in determining the partial structure thereof, and have considered that these cDNA sequences are preserved very well in the human and in the mouse, and are coding for novel receptor proteins for the same ligand. Based upon the above knowledge, the present inventors have discovered that these DNAs make it possible to obtain a cDNA having a full length open reading frame (ORF) of the receptor protein, hence, to produce the receptor protein. The inventors have further discovered that the above-mentioned receptor protein obtained when the G protein coupled receptor protein-encoding cDNA is expressed by a suitable means permits screening for a ligand to the receptor protein from the living body or from natural or non-natural compounds under guidance of data obtainable in receptor coupling tests or measurements of intracellular second messengers, etc. and further allows screening for a compound that inhibits the binding of the ligand and the receptor protein.

In one embodiment, the present inventors have carried out PCR amplification of novel human pituitary gland-

derived cDNA fragments as shown in Figures 22 and 23, and have subcloned them to obtain a plasmid vector (p19P2). From analysis of the partial sequence, it has been clarified that the cDNA has been encoded a novel receptor protein. The synthetic DNA primers used for amplifying the cDNA are corresponding to seven hydrophobic clusters that exist in the known G protein coupled receptor proteins in common, i.e., corresponding to the first and sixth membrane-spanning regions among the membrane-spanning domains. The nucleotide sequence (SEQ ID NO: 29) has been determined from the primer region at the 5' side (first membrane-spanning domain side) and has been translated into an amino acid sequence (SEQ ID NO: 24) [Figure 22]. As a result, the second and third membrane-spanning domains have been confirmed on the hydrophobicity plotting [Figure 58]. Similarly, the nucleotide sequence (SEQ ID NO: 30) has been determined from the primer region at the 3' side (sixth membrane-spanning domain side) and has been translated into an amino acid sequence (SEQ ID NO: 25) [Figure 23]. As a result, the presence of the sixth and fifth membrane-spanning domains has been confirmed on the hydrophobicity plots [Figure 59]. The size of the amplified cDNA is about 700 bp which is nearly comparable with the number of bases between the first membrane-spanning domain and the sixth membrane-spanning domain of the known G protein coupled receptor protein.

G protein coupled receptor proteins exert common property to some extent at an amino acid sequence level, and are forming one protein family. Therefore, data base retrieval has been carried out based upon the amino acid sequence of the subject novel receptor protein (protein encoded by cDNA included in p19P2). As a result, a high homology has been exhibited as compared with the known G protein coupled receptor protein (rat neuropeptide Y receptor protein encoded by SI2863) that is shown in Figure 60. This fact tells that the novel receptor protein of the present invention belongs to the G protein coupled receptor protein family. Moreover, the data base has been retrieved using, as a template, the amino

acid sequence encoded by the DNA of the invention. It exhibits high homology to the amino acid sequences of the known G protein coupled receptor proteins, mouse-derived ligand unknown RP-23 (B40470), human-derived ligand unknown K-opioid receptor protein (J30098) and human-derived NK-2 receptor protein (J01059). However, none of them are in perfect agreement, from which it is learned that a novel receptor protein had been encoded. The aforementioned abbreviations in parentheses are reference numbers that are assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are, usually, each called "Accession Number".

Next, by using the novel G protein coupled receptor protein-encoding cDNA fragment (p19P2) of the present invention, a cDNA having a full-length open reading frame of the receptor protein of the present invention has been obtained from human pituitary gland cDNA libraries. The nucleotide sequence analysis of a plasmid (pHGR3) carrying the cDNA having a full length open reading frame of the receptor protein shows that the nucleotide sequence of a coding region of this receptor protein is represented by SEQ ID NO: 31, and the amino acid sequence deduced therefrom is represented by SEQ ID NO: 26 [Figure 34]. Based upon the amino acid sequence, hydrophobicity plotting has been carried out. The results are shown in Figure 36. From the hydrophobicity plotting, it has been clarified that the receptor protein of the present invention possessed seven hydrophobic domains. That is, it has been confirmed that the receptor protein encoded by the cDNA obtained according to the present invention is a seven transmembrane (membrane-spanning) G protein coupled receptor protein. An expression of mRNA for receptor genes encoded by the cDNA of the present invention has been checked by northern blotting techniques at a mRNA level, and it has been confirmed that the receptor gene has been expressed in the human pituitary gland [Figure 35].

The present inventors have further succeeded in PCR amplification of a mouse pancreatic β cell strain, MIN6 derived cDNA fragment, and cloning of pG3-2 and pG1-10.

Then, based on the nucleotide sequence of cDNA included in these two plasmid vectors, the nucleotide sequence shown in Figure 27 has been derived. It was learned from the nucleotide sequence that the cDNA encodes a novel receptor protein.

Upon translating the nucleotide sequence into an amino acid sequence, the presence of the third, fourth, fifth and sixth membrane-spanning domains has been confirmed on the hydrophobicity plots (Figure 28). The size of the amplified cDNA is about 400 bp which is nearly comparable with the number of bases between the third membrane-spanning domain and the sixth membrane-spanning domain of the known G protein coupled receptor protein. The amino acid sequence has been compared with amino acid sequences [Figures 22 and 23] encoded by the G protein coupled receptor protein cDNA included in p19P2 cloned from the human pituitary gland. As a result, homology is more than 95% [Figure 61]. From this fact, it was estimated that the protein encoded by the cDNA included in pG3-2 is a mouse type G protein coupled receptor protein relative to the human-derived one encoded by the cDNA included in p19P2.

The present inventors have further amplified a mouse pancreatic β -cell strain, MIN6-derived cDNA fragment by the PCR followed by subcloning into a plasmid vector to obtain a clone (p5S38) having a nucleotide sequence as shown in Figure 62. From the nucleotide sequence (SEQ ID NO: 33), it has been clarified that the cDNA encodes a novel receptor protein. Upon translating the nucleotide sequence into an amino acid sequence (SEQ ID NO: 28), the presence of the third, fourth, fifth and sixth membrane-spanning domains has been confirmed on the hydrophobicity plots (Figure 64). The size of the amplified DNA is about 400 bp that is nearly comparable with the known G protein coupled receptor protein. The amino acid sequence has been compared with amino acid sequences [Figures 22 and 23] encoded by the G protein coupled receptor protein cDNA included in p19P2 cloned from the human pituitary gland, and with amino acid sequences of proteins encoded by pG3-2 and pG1-10 derived from the mouse pancreatic β -cell strain. As a result, homology is more than 95% to them

[Figure 63]. This fact suggests that the protein encoded by the human-derived pituitary gland-derived p19P2, the proteins encoded by the mouse pancreatic β -cell strain-derived pG3-2 and pG1-10, and the protein encoded by the mouse pancreatic β -cell strain-derived p5S38, pertain to a receptor family that recognizes the same ligand.

Another object of the present invention is to provide a novel human amygdaloid nucleus-derived protein coupled receptor protein, a DNA containing a DNA coding for said G protein coupled receptor protein, a process for producing said G protein coupled receptor protein, and use of said protein and DNA.

The present inventors have synthesized DNA primers for efficiently isolating a DNA coding for G protein coupled receptor proteins, amplified an amygdaloid nucleus-derived cDNA with the above primer, and have analyzed it.

As a result, the present inventors have succeeded in isolating, from the human amygdaloid nucleus, a cDNA coding for a novel G protein coupled receptor protein and have determined its partial structure. The nucleotide sequence of the isolated cDNA is preserved very well as compared with that of the mouse glucocorticoid-induced receptor (hereinafter sometimes referred to as "GIR") and is considered to be encoding a receptor protein to the same ligand (Molecular Endocrinology 5:1331-1338, 1991). It is reputed that, in the mouse, the GIR is a receptor which is induced by glucocorticoid and expressed in T-cells and is working as a receptor to immunoregulating factors in the immune system on the T-cells. The present inventors have succeeded in the isolation of this human type GIR from the human amygdaloid nucleus. Accordingly, it is suggested that the isolated GIR is expressed even in the human central nervous system to carry out some function. From these facts, it is considered that the receptor protein is strongly expressed in the human brain and in the immune system and is also functioning therein. These characterized DNAs allow one to obtain a cDNA having a full length open reading frame of the receptor and production of the receptor

proteins. The receptor proteins expressed by a suitable means, furthermore, permit screening for a ligand to the receptor proteins from the living body or from natural and non-natural compounds depending on indications obtainable in receptor protein-binding experiments, measurements of intracellular second messengers, etc. It further allows one to screen for compounds capable of inhibiting the binding between the ligand and the receptor protein.

To be more specific, the present inventors have amplified, as a novel human amygdaloid nucleus-derived cDNA, one species, as shown in Figures 29 and 30, by PCR, cloned it, and clarified from the analysis of a partial sequence thereof that a novel receptor protein is encoded. The synthetic DNA primers used for amplifying the cDNA are corresponding to seven hydrophobic clusters that exist in the G protein coupled receptor proteins in common, i.e., corresponding to the first and sixth membrane-spanning regions among the membrane-spanning domains. The nucleotide sequence has been determined from the primer region at the 5' side (first membrane-spanning domain side) and has been translated into an amino acid sequence. As a result, the second and third membrane-spanning domains have been confirmed on the hydrophobicity plotting [Figure 31]. Similarly, the nucleotide sequence has been determined from the primer region at the 3' side (sixth membrane-spanning domain side) and has been translated into an amino acid sequence. As a result, the presence of the fifth and fourth membrane-spanning domains has been confirmed on the hydrophobicity plots [Figure 32]. The size of the amplified cDNA is about 700 bp which is nearly comparable with the number of bases of the known G protein coupled receptor protein.

The inventors have further retrieved the data base based on, as a template, the nucleotide sequence of the isolated DNA and observed high homology to the DNA that codes for mouse-derived glucocorticoid-induced receptor protein which is a widely known G protein coupled receptor protein [Figure 33]. This result strongly suggests that the DNA of the present invention is encoding a human-type receptor protein of GR.

Yet another object of the present invention is to provide a novel mouse pancreatic β -cell strain, MIN6-derived protein coupled receptor protein, a DNA containing a DNA coding for said G protein coupled receptor protein, a process for producing said G protein coupled receptor protein, and use of said protein and DNA. The present inventors have synthesized DNA primers for efficiently isolating a DNA coding for G protein coupled receptor proteins, amplified a mouse pancreatic β -cell strain, MIN6-derived cDNA with the above primer, and have analyzed it.

As a result, the present inventors have succeeded in isolating a mouse-derived cDNA coding for a novel G protein coupled receptor protein and have determined its partial structure. The isolated cDNA is homologous to known G protein coupled receptors at the nucleotide sequence level and at the amino acid sequence level and is considered to be encoding a novel receptor protein which is expressed in the mouse pancreas and is also functioning therein. These characterized DNAs allow one to obtain a cDNA having a full length open reading frame of the receptor and production of the receptor proteins. Human-derived cDNAs may be cloned by using, as a probe, said mouse-derived cDNA. The receptor proteins expressed by a suitable means, furthermore, permit screening for a ligand to the receptor protein from the living body or from natural and non-natural compounds relying on indications obtainable in receptor protein-binding experiments, measurements of intracellular second messengers, etc. It further allows one to screen for compounds capable of inhibiting the binding of the ligand with the receptor protein.

To be more specific, the present inventors have amplified, as a novel mouse pancreatic β -cell strain, MIN6-derived cDNA, PJH2-17, as shown in Figures 37, by PCR, cloned it, and clarified from the analysis of a partial sequence thereof that a novel receptor protein is encoded. The nucleotide sequence has been translated into an amino acid sequence. As a result, the presence of the third, fourth, fifth and sixth membrane-spanning domains has been confirmed

on the hydrophobicity plots [Figure 38]. The size of the amplified cDNA is about 400 bp which is nearly comparable with that of the known G protein coupled receptor protein.

The inventors have retrieved the data base based on, as a template, the nucleotide sequence of the isolated DNA and observed 30% homology to chicken ATP receptor (P34996), 25% homology to human somatostatin receptor subtype 3 (A46226), 27% homology to human somatostatin receptor subtype 4 (JN0605), and 28% homology to bovine neuropeptide Y receptor (S28787), respectively (Figure 39), which are known G protein coupled receptor proteins. The aforementioned abbreviations in parentheses are reference numbers that are assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are, usually, each called "Accession Number".

An expression of receptor genes encoded by the cDNA fragment included in p3H2-17 of the present invention has been checked by northern blotting techniques at a mRNA level, and it has been confirmed that the receptor gene has been intensely expressed in the mouse thymus and spleen. It has been also confirmed that the receptor gene has been expressed in the mouse brain and pancreas (Figure 65).

Next, by utilizing the information on the nucleotide sequence of the fragment included in p3H2-17, cDNA encoding a full-length open reading frame of the mouse pancreatic β -cell strain, MIN6-derived G protein coupled receptor protein of the present invention has been obtained from mouse

thymic and splenic poly(A)⁺ RNA by 5'RACE (5' rapid amplification of cDNA ends) techniques (Frohman M.A. et al., Proc. Natl. Acad. Sci. USA, 85:8998-9002 (1988); Belyavsky A. et al., Nucleic Acids Res., 17:2919-2932 (1989); Edwards J.B.D.M. et al., Nucleic Acids Res., 19:5227-5232 (1991)) and 3'RACE (3' rapid amplification of cDNA ends) techniques (Frohman M.A. et al., Proc. Natl. Acad. Sci. USA, 85:8998-9002 (1988); Belyavsky A. et al., Nucleic Acids Res., 17:2919-2932 (1989)).

The plasmid (pMAH2-17) carrying cDNA encoding a full-length open reading frame of the receptor protein of the

present invention has been subjected to sequencing analysis.

As a result, the nucleotide sequence of the region coding for the receptor protein is represented by SEQ ID NO: 41 and the amino acid sequence deduced therefrom is represented by SEQ ID NO: 39 (Figure 69). Based on the amino acid sequence, hydrophobicity plotting has been carried out. The results are shown in Figure 70.

It has been clarified from the hydrophobicity plotting that the mouse pancreatic β -cell strain, MIN6-derived receptor protein of the present invention has seven hydrophobic domains. Thus, it has been confirmed that the receptor protein encoded by the cDNA included in pMAH2-17 according to the present invention is a seven transmembrane G protein coupled receptor protein.

Data base retrieval has been carried out based on the full-length amino acid sequence encoded by the cDNA included in pMAH2-17, and it has been observed that the amino acid sequence has 44.0% homology to mouse P_{2U}purinoceptor (P35383) and 38.1% homology to chicken P_{2Y}purinoceptor (P34996), respectively (Figure 71), which are known G protein coupled receptor proteins. The aforementioned abbreviations in parentheses are reference numbers that are assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are, usually, each called "Accession Number".

Since the receptor protein encoded by pMAH2-17 is highly homologous to prinoceptors, it is considered that there are strong possibility of a subtype within prinoceptor families. Therefore, the present inventors have carried out an electrophysiological analysis of the receptor gene in *Xenopus* oocytes and found significant inward currents elicited by *Xenopus* oocytes carrying the subject receptor gene in response to ATP stimulation (Figure 75). As a result, it has been determined that the receptor encoded by pMAH2-17 is one of the subtypes within prinoceptor families. It has been discussed and expected that there are a variety of subtypes among purinoceptors (Pharmac. Ther., Vol. 64, pp. 445-475 (1994)).

All data are supporting that the mouse pancreatic β -cell strain, MIN6-derived receptor protein of the present invention (e.g., SEQ ID NO: 38 and SEQ ID NO: 39, or proteins encoded by pMAH2-17) is a novel purinoceptor subtype which is clearly distinct from chicken P_{2u}1 purinoceptor (FEBS LETTERS, Vol. 324(2), 219-225 (1993)); mouse P_{2y}2 or P_{2u} purinoceptor (Proc. Natl. Acad. Sci. USA, Vol. 90, pp.5113-5117 (1993)); rat P_{2u} or P_{2y}2 purinoceptor (Am. J. Respir. Cell Mol. Biol., Vol. 12, pp. 27-32 (1995)); human P_{2u} or P_{2y}2 purinoceptor (Proc. Natl. Acad. Sci. USA, Vol. 91, pp.3275-3279 (1994)); and rat P_{2x} purinoceptor (Nature, Vol. 371.6, pp.516-519 (1994)).

It is also strongly suggested that agonists and/or antagonists related to the receptor encoded by pMAH2-17 would be useful in therapeutic or prophylactic treatment of diseases or syndromes in connection with purine ligand compounds. It is expected that the agonists of the receptor encoded by pMAH2-17 are useful as an immunomodulator or an antitumor agent, in addition they are useful in therapeutically or prophylactically treating hypertension, diabetes, cystic fibrosis, etc. It is still expected that the antagonists of the receptor encoded by pMAH2-17 are useful as hypotensive agents, analgesics, agents for therapeutically or prophylactically treating incontinence of urine, etc.

Another object of the present invention is to provide a novel human-derived protein coupled receptor protein of prinoceptor type, a DNA containing a DNA coding for said G protein coupled receptor protein, a process for producing said G protein coupled receptor protein, and use of said protein and DNA. The present inventors have synthesized DNA primers for efficiently isolating a DNA coding for prinoceptor type G protein coupled receptor proteins on the basis of the nucleotide sequence of mouse purinoceptor, amplified a human-derived cDNA with the above primer, and have analyzed it.

As a result, the present inventors have succeeded in isolating a human-derived cDNA coding for a novel G protein coupled receptor protein and have determined its full-length structure [figure 77]. The isolated cDNA is homologous to

mouse G protein coupled receptor (purinoceptor) at the nucleotide sequence level and at the amino acid sequence level (87% homology; Figure 79) and is considered to be encoding a novel purinoceptor protein. The receptor proteins expressed by a suitable means, furthermore, permit screening for a ligand to the receptor protein from the living body or from natural and non-natural compounds relying on indications obtainable in receptor protein-binding experiments, etc. It further allows one to screen for compounds capable of inhibiting the binding of the ligand with the receptor protein.

It is also strongly suggested that agonists and/or antagonists related to the human receptor encoded by pMAH2-17 would be useful in therapeutic or prophylactic treatment of diseases or syndromes in connection with purine ligand compounds. It is expected that the agonists of the human receptor are useful as an immunomodulator or an antitumor agent, in addition they are useful in therapeutically or prophylactically treating hypertension, diabetes, cystic fibrosis, etc. It is still expected that the antagonists of the human receptor are useful as hypotensive agents, analgesics, agents for therapeutically or prophylactically treating incontinence of urine, etc.

Accordingly, one aspect of the present invention is

- (1) DNAs comprising a nucleotide sequence represented by a SEQ ID NO selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 19;
- (2) DNAs according to the above (1) comprising a nucleotide sequence represented by a SEQ ID NO selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 9;
- (3) DNAs according to the above (1) comprising a nucleotide sequence represented by SEQ ID NO: 1 or SEQ ID NO: 2;
- (4) DNAs according to the above (1) wherein the DNA is a primer for polymerase chain reaction in order to amplify a DNA coding for a G protein coupled receptor protein;
- (5) a method for amplifying a DNA coding for a G

protein coupled receptor protein by polymerase chain reaction techniques, which comprises:

(i) carrying out a polymerase chain reaction in the presence of a mixture of

- 5 ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and
- 20 ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19; or
- 30 (ii) carrying out a polymerase chain reaction in the presence of a mixture of
- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,

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② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

5

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13;

(6) a method for screening a DNA library for a DNA coding for a G protein coupled receptor protein, which comprises:

10

(i) carrying out a polymerase chain reaction in the presence of a mixture of

① said DNA library,

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② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and

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③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, DNA primers comprising a nucleotide

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- 2 7 -

sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19, to amplify selectively a template DNA coding for G protein coupled receptor protein, contained in the DNA library; or (ii) carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13, to amplify selectively a DNA coding for G protein coupled receptor protein, contained in the DNA library;

(7) a DNA coding for a G protein coupled receptor protein, which is obtained by a method according to the above (5) or (6); and

(8) G protein coupled receptor proteins encoded by a DNA according to the above (7), their peptide segments or fragments and salts thereof.

Another specific aspect of the invention is:

(9) a method for amplifying a DNA coding for G protein coupled receptor protein (e.g. a region corresponding to from the first to sixth membrane-spanning domains of G protein coupled receptor proteins or other domains thereof) by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers

- 2 8 -

comprising a nucleotide sequence represented by SEQ ID NO: 12, and

- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19;

(10) a method for amplifying a DNA coding for G protein coupled receptor protein (e.g. a region corresponding to from the first to seventh membrane-spanning domains of G protein coupled receptor proteins or other domains thereof) by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11;

(11) a method for amplifying a DNA coding for G protein coupled receptor protein (e.g. a region corresponding to from the third to sixth membrane-spanning domains of G protein coupled receptor proteins or other domains thereof) by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein,

② said DNA being capable of acting as a template,
 at least one DNA primer selected from the group
 consisting of DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 3, DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 5, DNA primers comprising a nucleotide sequence
 represented by SEQ ID NO: 6, DNA primers comprising a
 nucleotide sequence represented by SEQ ID NO: 7, DNA
 primers comprising a nucleotide sequence represented by
 SEQ ID NO: 14 and DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 18, and

③ at least one DNA primer selected from the group
 consisting of DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 2, DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 4, DNA primers comprising a nucleotide sequence
 represented by SEQ ID NO: 8, DNA primers comprising a
 nucleotide sequence represented by SEQ ID NO: 9, DNA
 primers comprising a nucleotide sequence represented by
 SEQ ID NO: 15, DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 17 and DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 19;

(12) a method for amplifying a DNA coding for G
 protein coupled receptor protein (e.g. a region corresponding
 to from the third to seventh membrane-spanning domains of G
 protein coupled receptor proteins or other domains thereof) by
 polymerase chain reaction techniques, which comprises carrying
 out a polymerase chain reaction in the presence of a mixture of

① a DNA coding for G protein coupled receptor protein,
 said DNA being capable of acting as a template,

② at least one DNA primer selected from the group
 consisting of DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 3, DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 5, DNA primers comprising a nucleotide sequence
 represented by SEQ ID NO: 6, DNA primers comprising a

nucleotide sequence represented by SEQ ID NO: 7, DNA
 primers comprising a nucleotide sequence represented by
 SEQ ID NO: 14 and DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 18, and

③ at least one DNA primer selected from the group
 consisting of DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 11;

(13) a method for amplifying a DNA coding for G
 protein coupled receptor protein (e.g. a region corresponding
 to from the second to sixth membrane-spanning domains of G
 protein coupled receptor proteins or other domains thereof) by
 polymerase chain reaction techniques, which comprises carrying
 out a polymerase chain reaction in the presence of a mixture of

① a DNA coding for G protein coupled receptor protein,
 said DNA being capable of acting as a template,

② at least one DNA primer selected from the group
 consisting of DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 10 and DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 16, and

③ at least one DNA primer selected from the group
 consisting of DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 2, DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 4, DNA primers comprising a nucleotide sequence
 represented by SEQ ID NO: 8, DNA primers comprising a
 nucleotide sequence represented by SEQ ID NO: 9, DNA
 primers comprising a nucleotide sequence represented by
 SEQ ID NO: 15, DNA primers comprising a nucleotide
 sequence represented by SEQ ID NO: 17 and DNA primers
 comprising a nucleotide sequence represented by SEQ ID
 NO: 19;

(14) a method for amplifying a DNA coding for G
 protein coupled receptor protein (e.g. a region corresponding
 to from the second to seventh membrane-spanning domains of G
 protein coupled receptor proteins or other domains thereof) by
 polymerase chain reaction techniques, which comprises carrying

out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16, and

- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11;

(15) a method for amplifying a DNA coding for G protein coupled receptor protein (e.g. a region corresponding to from the first to third membrane-spanning domains of G protein coupled receptor proteins or other domains thereof) by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13;

(16) a method for amplifying a DNA coding for G protein coupled receptor protein by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, and
- ③ at least one DNA primer selected from the group

consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2;

- (17) a method for amplifying a DNA coding for G protein coupled receptor protein by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4;

(18) a method for amplifying a DNA coding for G protein coupled receptor protein by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8;

(19) a method for amplifying a DNA coding for G protein coupled receptor protein by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide

sequence represented by SEQ ID NO: 11;

(20) a method for amplifying DNA coding for a G protein coupled receptor protein which comprises

(i) carrying out a polymerase chain reaction in the presence of a mixture of

① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,

② at least one DNA primer which is capable of binding with the 3'-side nucleotide sequence of

the - chain (minus chain) of the template DNA coding for G protein coupled receptor protein to

allow the extension of the + chain (plus chain) in the 5' → 3' direction, said DNA primer being

selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID

NO: 1, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a

nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by

SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers

comprising a nucleotide sequence represented by SEQ ID NO: 10, DNA primers comprising a

nucleotide sequence represented by SEQ ID NO: 12, DNA primers comprising a nucleotide sequence

represented by SEQ ID NO: 14, DNA primers comprising a nucleotide sequence represented by

SEQ ID NO: 16 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18,

and

③ at least one DNA primer which is capable of binding with the 3'-side nucleotide sequence of the + chain (plus chain) of the template DNA

coding for G protein coupled receptor protein to allow the extension of the - chain (minus chain)

in the 5' → 3' direction, said DNA primer being selected from the group consisting of DNA primers

comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA

primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide

sequence represented by SEQ ID NO: 11, DNA

primers comprising a nucleotide sequence

represented by SEQ ID NO: 15, DNA primers

comprising a nucleotide sequence represented by

SEQ ID NO: 17 and DNA primers comprising a

nucleotide sequence represented by SEQ ID NO: 19,

or

(ii) carrying out a polymerase chain reaction in the presence of a mixture of

① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,

② at least one DNA primer which is capable of binding with the 3'-side nucleotide sequence of

the - chain (minus chain) of the template DNA coding for G protein coupled receptor protein to

allow the extension of the + chain (plus chain) in the 5' → 3' direction, said DNA primer being

selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID

NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

③ at least one DNA primer which is capable of binding with the 3'-side nucleotide sequence of

the + chain (plus chain) of the template DNA coding for G protein coupled receptor protein to

allow the extension of the - chain (minus chain) in the 5' → 3' direction, said DNA primer being

selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID

NO: 13;

(21) a method for screening DNA libraries for

a DNA coding for G protein coupled receptor protein (e.g. from the first to sixth membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19,

to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the first to sixth membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;

- (22) a method for screening DNA libraries for a DNA coding for G protein coupled receptor protein (e.g. from the first to seventh membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide

sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11,

to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the first to seventh membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;

- (23) a method for screening DNA libraries for a DNA coding for G protein coupled receptor protein (e.g. from the third to sixth membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and

- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 15, DNA

primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19,

5 to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the third to sixth membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;

(24) a method for screening DNA libraries for

10 a DNA coding for G protein coupled receptor protein (e.g. from the third to seventh membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- 15 ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and
- 20 ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11,
- 25 to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the third to seventh membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;
- (25) a method for screening DNA libraries for
- 30 a DNA coding for G protein coupled receptor protein (e.g. from the second to sixth membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises

carrying out a polymerase chain reaction in the presence of a mixture of

- 5 ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16, and
- 10 ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19,
- 15 to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the second to sixth membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;
- (26) a method for screening DNA libraries for
- 20 a DNA coding for G protein coupled receptor protein (e.g. from the second to seventh membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- 25 ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16, and
- 30 ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16, and
- 35

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the second to seventh membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;

(27) a method for screening DNA libraries for a DNA coding for G protein coupled receptor protein (e.g. from the first to third membrane-spanning domains or other domains of G protein coupled receptor protein), which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13, to amplify selectively a template DNA coding for G protein coupled receptor protein (e.g. from the first to third membrane-spanning domains or other domains of G protein coupled receptor protein), contained in the DNA library;

(28) a method for screening DNA libraries for a DNA coding for G protein coupled receptor protein, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2,

to amplify selectively the template DNA coding for G protein coupled receptor protein, contained in the DNA library;

(29) a method for screening DNA libraries to detect a DNA coding for G protein coupled receptor protein, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4,

to amplify selectively a template DNA coding for G protein coupled receptor protein, contained in the DNA library;

(30) a method for screening DNA libraries for a DNA coding for G protein coupled receptor protein, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, and
- ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8,

to amplify selectively a template DNA coding for G protein coupled receptor protein, contained in the DNA library;

(31) a method for screening DNA libraries for a DNA coding for G protein coupled receptor protein, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, and
- ③ at least one DNA primer selected from the group

consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, to amplify selectively a template DNA coding for G protein coupled receptor protein, contained in the DNA library; and (32) a method for screening DNA libraries according to any of the above (6), and (21) to (31) wherein said DNA library is derived from an origin selected from the group consisting of human tissues and human cells. Examples of such human tissues include adrenal, umbilical cord, brain, tongue, liver, lymph gland, lung, thymus, placenta, peritoneum, retina, spleen, heart, smooth muscle, intestine, vessel, bone, kidney, skin, fetus, mammary gland, ovary, testis, pituitary gland, pancreas, submandibular gland, spine, prostate gland, stomach, thyroid gland, trachea (windpipe), skeletal muscle, uterus, adipose tissue, urinary bladder, cornea, olfactory bulb, bone marrow, amnion, etc. Examples of such human cells include nerve cells, epithelial cells, endothelial cells, leukocytes, lymphocytes, gliocytes, fibroblasts, keratinized cells, osteoblasts, osteoclasts, astrocytes, melanocytes, various carcinomas, various sarcomas, various cells derived from the above-mentioned human tissues.

Yet another aspect of the present invention is a degenerate deoxynucleotide which has an oligonucleotide sequence to which a SEQ ID NO selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 19 is assigned.

Another aspect of the present invention is

(33) a G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of amino acid sequences represented by SEQ ID NO: 24 and/or SEQ ID NO: 25 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 24 or SEQ ID NO: 25; or a salt thereof;

(34) a G protein coupled receptor protein according to the above (33) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 26 and substantial equivalents to the amino

acid sequence represented by SEQ ID NO: 26; or a salt thereof; (35) a G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 27 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 27; or a salt thereof;

(36) a G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 28 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 28; or a salt thereof;

(37) a G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of amino acid sequences represented by SEQ ID NO: 34 and/or SEQ ID NO: 35 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 34 or SEQ ID NO: 35; or a salt thereof;

(38) a G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 38 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 38; or a salt thereof;

(39) a G protein coupled receptor protein according to the above (38) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 39 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 39; or a salt thereof;

(40) a G protein coupled receptor protein comprising an amino acid sequence represented by SEQ ID NO: 56 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 56; or a salt thereof;

(41) a peptide segment or fragment of a G protein coupled receptor protein according to any of the above (33) to (40), a modified derivative thereof or a salt thereof;

(42) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (33);

- (43) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (34);
- (44) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (35);
- (45) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (36);
- (46) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (37);
- (47) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (38);
- (48) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (39);
- (49) a DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of the above (40);
- (50) a DNA of the above (42) comprising a nucleotide sequence represented by SEQ ID NO: 29 and/or SEQ ID NO: 30;
- (51) a DNA of the above (43) comprising a nucleotide sequence represented by SEQ ID NO: 31;
- (52) a DNA of the above (44) comprising a nucleotide sequence represented by SEQ ID NO: 32;
- (53) a DNA of the above (45) comprising a nucleotide sequence represented by SEQ ID NO: 33;
- (54) a DNA of the above (46) comprising a nucleotide sequence represented by SEQ ID NO: 36 and/or SEQ ID NO: 37;
- (55) a DNA of the above (47) comprising a nucleotide sequence represented by SEQ ID NO: 40;
- (56) a DNA of the above (48) comprising a nucleotide sequence represented by SEQ ID NO: 41;
- (57) a DNA of the above (49) comprising a nucleotide sequence represented by SEQ ID NO: 57;

- (58) a vector comprising a DNA according to any of the above (42) to (57);
- (59) a transformant (including a transfectant) carrying a vector of the above (58);
- (60) a process for producing a G protein coupled receptor protein or a salt thereof according to any of the above (33) to (40), which comprises culturing a transformant of the above (59) to express said G protein coupled receptor protein on the membrane of the transformant;
- (61) a method for determining a ligand to a G protein coupled receptor protein according to any of the above (33) to (40), which comprises
- contacting
- (i) at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof, with
- (ii) at least one compound to be tested;
- (62) a screening method for a compound capable of inhibiting the binding of a G protein coupled receptor protein according to any of the above (33) to (40) with a ligand, which comprises carrying out a comparison between:
- (i) at least one case where said ligand is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof, according to the above (41), and mixtures thereof, and
- (ii) at least one case where said ligand together with a compound to be tested is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the

above (41), and mixtures thereof;

(63) a kit for the screening of one or more compounds capable of inhibiting the binding of a G protein coupled receptor protein according to any of the above (33) to (40), with a ligand, which comprises at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof; and

(64) an antibody against at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof.

Yet another aspect of the present invention is

(65) a G protein coupled receptor protein according to the above (33) comprising

(i) an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 24, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 24, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 24, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 24 are substituted with one or more other amino acid residues, or/and

(ii) an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 25, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 25, amino acid

sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 25, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 25 are substituted with one or more other amino acid residues, or a salt thereof;

(66) a G protein coupled receptor protein according to the above (34) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 26, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 26, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 26, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 26 are substituted with one or more other amino acid residues, or a salt thereof;

(67) a G protein coupled receptor protein according to the above (35) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 27, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 27, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 10 amino acid residues) in

the amino acid sequence of SEQ ID NO: 27 are substituted with one or more other amino acid residues, or a salt thereof;

- (68) a G protein coupled receptor protein according to the above (36) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 28, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 28, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 28, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 28 are substituted with one or more other amino acid residues, or a salt thereof;

(69) a G protein coupled receptor protein according to the above (37) comprising

- (i) an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 34, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 34, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 34, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 34 are substituted with one or more other amino acid residues, or/and
- (ii) an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 35, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more

- preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 35, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 35, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 35 are substituted with one or more other amino acid residues, or a salt thereof;

- (70) a G protein coupled receptor protein according to the above (38) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 38, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 38, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 38, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 38 are substituted with one or more other amino acid residues, or a salt thereof;

- (71) a G protein coupled receptor protein according to the above (39) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 39, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 39, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 39, and amino acid sequences wherein one

or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 39 are substituted with one or more other amino acid residues, or a salt thereof;

(72) a G protein coupled receptor protein according to the above (40) comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 56, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 56, amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 56, and amino acid sequences wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 56 are substituted with one or more other amino acid residues, or a salt thereof;

(73) a method for determining a ligand according to the above (61) wherein said ligand is selected from the group consisting of angiotensin, bombesin, canavaninoid,

cholecystokinin, glutamine, serotonin, melatonin, neuropeptide Y, opioid, purine, vasopressin, oxytocin, VIP (vasoactive intestinal and related peptides), somatostatin, dopamine, motilin, amylin, bradykinin, CGRP (calcitonin gene related peptides), adrenomedullin, leukotriene, pancreastatin, prostaglandin, thromboxanes, adenosine, adrenaline, α - and β -chemokine (IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HCL4, MCP-3, I-309, MIP1 α , MIP-1 β , RANTES, etc.), endothelin, enterogastrin, histamine, neurotensin, TRH, pancreatic polypeptides and galanin;

(74) a method for the screening of a compound or a salt thereof capable of inhibiting the binding of a ligand with a G protein coupled receptor protein according to any of the above (33) to (40), which comprises measuring amounts of a labeled ligand bound to the said G protein coupled receptor

protein in at least two cases:

(i) where the labeled ligand is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof, and

(ii) where the labeled ligand together with a compound to be tested is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof,

and comparing the measured amounts of the labeled ligand;

(75) a method for the screening of a compound or a salt thereof capable of inhibiting the binding of a ligand with a G protein coupled receptor protein according to any of the above (33) to (40), which comprises measuring amounts of a labeled ligand bound to a cell comprising the said G protein coupled receptor protein in at least two cases:

(i) where the labeled ligand is contacted with the said cell, and

(ii) where the labeled ligand together with a compound to be tested is contacted with the said cell,

and comparing the measured amounts of the labeled ligand;

(76) a method for the screening of a compound or a salt thereof capable of inhibiting the binding of a ligand with a G protein coupled receptor protein according to any of the above (33) to (40), which comprises measuring amounts of a labeled ligand bound to a membrane fraction of a cell comprising the said G protein coupled receptor protein in at least two cases:

(i) where the labeled ligand is contacted with the said membrane fraction, and

(ii) where the labeled ligand together with a compound to be tested is contacted with the membrane fraction,

- 5 1 -

and comparing the measured amounts of the labeled ligand;

(77) a method for the screening of a compound or a salt thereof capable of inhibiting the binding of a ligand with a G protein coupled receptor protein according to any of the above (33) to (40), which comprises measuring amounts of a labeled ligand bound to said G protein coupled receptor protein in at least two cases:

- (i) where the labeled ligand is contacted with a G protein coupled receptor protein according to any of the above (33) to (40) which is expressed on the membrane of a transformant according to the above (59) during incubation of the transformant, and
- (ii) where the labeled ligand together with a compound to be tested is contacted with the G protein coupled receptor protein according to any of the above (33) to (40) which is expressed on the membrane of a transformant according to the above (59) during incubation of the transformant,

and comparing the measured amounts of the labeled ligand;

(78) a method for the screening of a compound or a salt thereof capable of inhibiting the binding of a ligand with a G protein coupled receptor protein according to any of the above (33) to (40), which comprises measuring G protein coupled receptor protein-mediated cell-stimulating activities in at least two cases:

- (i) where a compound capable of activating the G protein coupled receptor protein according to any of the above (33) to (40) is contacted with a cell comprising the said G protein coupled receptor protein, and
- (ii) where the compound capable of activating the G protein together with a compound to be tested is contacted with the cell comprising the said G protein coupled receptor protein,

and comparing the measured cell-stimulating activities;

(79) a method for the screening of a compound or a salt thereof capable of inhibiting the binding of a ligand with a G protein coupled receptor protein according to any of the

- 5 2 -

above (33) to (40), which comprises measuring G protein coupled receptor protein-mediated cell-stimulating activities in at least two cases:

- (i) where a compound capable of activating the G protein coupled receptor protein according to any of the above (33) to (40) is contacted with a G protein coupled receptor protein according to any of the above (33) to (40) which is expressed on the membrane of a transformant according to the above (59) during incubation of the transformant, and
- (ii) where the compound capable of activating the G protein together with a compound to be tested is contacted with the G protein coupled receptor protein according to any of the above (33) to (40) which is expressed on the membrane of a transformant according to the above (59) during incubation of the transformant,

and comparing the measured cell-stimulating activities;

(80) a method according to the above (78) or (79) wherein said compound capable of activating the G protein coupled receptor protein according to any of the above (33) to (40) is selected from the group consisting of angiotensin, bombesin, canavaninoid, cholecystokinin, glutamine, serotonin, melatonin, neuropeptide Y, opioid, purine, vasopressin, oxytocin, VIP (vasoactive intestinal and related peptides), somatostatin, dopamine, motilin, amylin, bradykinin, CGRP (calcitonin gene related peptides), adrenomedullin, leukotriene, pancreastatin, prostaglandin, thromboxane, adenosine, adrenaline, α - and β -chemokine (IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HCL14, MCP-3, I-309, MIP1 α , MIP-1 β , RANTES, etc.), endothelin, enterogastrin, histamine, neurotensin, TRH, pancreatic polypeptides and galanin;

(81) a compound which is determined through a method according to any of the above (62) and (74) to (80) or a salt thereof;

(82) a pharmaceutical composition comprising an

effective amount of a compound according to the above (81) or a salt thereof;

(83) a screening kit according to the above (63), comprising a cell comprising a G protein coupled receptor protein according to any of the above (33) to (40);

(84) a screening kit according to the above (63), comprising a membrane fraction derived from a cell comprising a G protein coupled receptor protein according to any of the above (33) to (40);

(85) a screening kit according to the above (63), comprising a cell of the (59) or (109) mentioned herein below;

(86) a screening kit according to the above (63), comprising a membrane fraction derived from a cell of the (59) or (109);

(87) a compound which is determined by means of a screening kit according to any of the above (63) and (83) to (86) or a salt thereof;

(88) a pharmaceutical composition comprising an effective amount of a compound according to the above (87) or a salt thereof; and

(89) a method for measuring at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof, which comprises contacting an antibody according to the above (64) with the component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide segments or salts thereof according to the above (41), and mixtures thereof.

Still another aspect of the present invention is (90) a ligand to a G protein coupled receptor protein according to any of the above (33) to (40), which is determined through the following step of:

contacting (i) at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above

(33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof,

with (ii) at least one compound to be examined; and

(91) a compound capable of inhibiting the binding of a G protein coupled receptor protein according to any of the above (33) to (40) with a ligand, which is determined through carrying out a comparison between:

(i) at least one case where said ligand is contacted

with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof, and

(ii) at least one case where said ligand together with a compound to be tested is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof.

Another aspect of the present invention is

(92) a recombinant G protein coupled receptor protein and a salt thereof which is obtained by the expression of a DNA according to any of the above (42) to (57), or a modified or fragmented derivative thereof;

(93) a method for amplifying a DNA coding for G protein coupled receptor protein by polymerase chain reaction techniques, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

(1) a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template, and

(2) at least one DNA primer selected from the group consisting of DNA primers comprising either SEQ ID NO: 1 or SEQ ID NO: 2; and

(94) a method for screening DNA libraries for

a DNA coding for G protein coupled receptor protein, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

(1) said DNA library, and

(2) at least one DNA primer selected from the group consisting of DNA primers comprising either SEQ ID NO: 1 or SEQ ID NO: 2,

to amplify selectively the DNA coding for G protein coupled receptor protein, contained in the DNA library.

10 Yet another aspect of the present invention is

(95) a monoclonal antibody against at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof;

(96) a preparation of purified polyclonal antibodies against at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof;

(97) an immunoassay for detecting a G protein coupled receptor protein which comprising

(i) incubating a sample to be tested with an antibody according to the above (64) to allow formation of an antigen-antibody complex; and

(ii) detecting an antigen-antibody complex formed in step (i); and

(98) an immunoassay for detecting antibodies

against a G protein coupled receptor protein which comprising
 (i) incubating a sample to be tested with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof to allow formation of an antigen-antibody complex; and

(ii) detecting an antigen-antibody complex formed in step (a).

Still another aspect of the present invention is

(99) an antisense DNA or RNA which comprises a nucleotide sequence complementary to at least a portion of a DNA according to any of the above (42) to (57), said antisense DNA or RNA being hybridizable to said DNA according to any of the above (42) to (57);

(100) an antisense DNA or RNA according to the above (99) wherein said antisense DNA or RNA comprises the 5' end hairpin loop, 5' end 6-base-pair repeat, 5' end untranslated region, protein translation initiation site or codon, ORF translation initiation site or codon, 3'-untranslated region, 3' end palindrome region, or 3' end hairpin loop of a G protein coupled receptor protein DNA according to any of the above (42) to (57);

(101) an antisense DNA or RNA according to the above (99) in a pharmaceutically acceptable carrier;

(102) an antisense DNA or RNA according to the above (99) comprising from 2 to 50 nucleotides;

(103) a method for modulating the activity of a G protein coupled receptor protein comprising contacting cells expressing the G protein coupled receptor protein with an antisense DNA or RNA according to the above (99);

(104) a method for producing an antibody against a G protein coupled receptor protein according to any of the above (33) to (40), which comprises administering to an individual at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof; and

(105) a method for producing a hybridoma which produces a monoclonal antibody against a G protein coupled receptor protein according to any of the above (33) to (40), which comprises

(i) immunizing an individual with at least one

component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of the above (33) to (40), peptide fragments or segments or salts thereof according to the above (41), and mixtures thereof;

(ii) immortalizing antibody producing cells from the immunized individual;

(iii) selecting an immortal cell which produces antibodies reactive with the G protein coupled receptor protein; and

(iv) growing said immortal cell.

Yet another aspect of the present invention is

(106) a PCR screening kit for a DNA (or nucleotide sequence) coding for G protein coupled receptor protein in a DNA library which comprises

(i) ① at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and

② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, DNA primers comprising a nucleotide

sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19; or

(ii) ① at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ NO: 12, and

② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13;

(107) a vector comprising the DNA according to the above (7);

(108) an expression system comprising an open reading frame (ORF) of DNA derived from a G protein coupled receptor protein DNA according to any of the above (7) and (42) to (57), wherein the ORF is operably linked to a control sequence compatible with a desired host cell;

(109) a transformant (including a transfectant) carrying a vector of the above (107) or an expression system of the above (108);

(110) a process for producing a G protein coupled receptor protein or a salt thereof, which comprises culturing the transformant of the above (109) to express said G protein coupled receptor protein on the membrane of the transformant;

(111) a method for expressing a polypeptide of G protein coupled receptor protein, comprising:

(a) providing a transformant of the above (59) or (109); and

(b) incubating the transformant under conditions which allow expression of the polypeptide of G protein coupled receptor protein;

(112) a method for preparing a transformant according to the above (59) or (109), comprising:

(a) providing a host cell capable of transformation;

(b) providing a vector according to the above (58) or (107) or an expression system according to the above (108); and

(c) incubating (a) with (b) under conditions which allow transformation of the host cell with the vector or the expression system;

(113) a pharmaceutical composition according to the above (82) or (88), comprising an effective amount of a compound according to the above (81) or (87) or a pharmaceutically acceptable salt thereof in admixture with a pharmaceutically acceptable carrier, excipient or diluent;

(114) the pharmaceutical composition according to the above (82) or (88), for inhibiting the binding of a G protein coupled receptor protein according to the present invention with a ligand;

(115) a method for inhibiting the binding of a G protein coupled receptor protein according to the present invention with a ligand in a medium which comprises contacting an effective amount of a compound according to the above (81) or (87) or a salt thereof with said medium;

(116) a method for modulating the activity of a G protein coupled receptor protein comprising contacting cells expressing the G protein coupled receptor protein with an effective amount of a compound according to the above (81) or (87) or a salt thereof;

(117) the ligand according to the above (90) being labeled with a detectable reporter;

(118) the antibody according to the above (64) wherein the antibody is labeled with a detectable reporter;

(119) a pharmaceutical composition for controlling an expression of G protein coupled receptor protein, which comprises an effective amount of the antisense DNA according to the above (99), and

(120) a culture product produced by a transformant according to the above (59) or (109).

Yet another aspect of the present invention is (121) a DNA according to the above (1) wherein the

DNA is an oligonucleotide having from 8 to 60 base residues; (122) a DNA according to the above (1) wherein the DNA is synthetic;

(123) a DNA (or nucleotide sequence) coding for a G protein coupled receptor protein or a fragment thereof, which is obtained through the method according to any of the above (5) to (32);

(124) a DNA (or nucleotide sequence) according to the above (123), wherein said G protein coupled receptor protein is selected from the group consisting of angiotensin receptor, bombesin receptor, canavaninoid receptor, cholecystokinin receptor, glutamine receptor, serotonin receptor, melatonin receptor, neuropeptide Y receptor, opioid receptor, purine receptor, vasopressin receptor, oxytocin receptor, VIP receptor (vasoactive intestinal and related peptide receptor), somatostatin receptor, dopamine receptor, motilin receptor, amylin receptor, bradykinin receptor, CGRP receptor (calcitonin gene related peptide receptor), adrenomedullin receptor, leukotriene receptor, pancreastatin receptor, prostaglandin receptor, thromboxane receptor, adenosine receptor, adrenaline receptor, α - and β -chemokine receptor including IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HCL4, MCP-3, I-309, MIP1 α , MIP-1 β , and RANTES receptors, endothelin receptor, enterogastin receptor, histamine receptor, neurotensin receptor, TRH receptor, pancreatic polypeptide receptor, and galanin receptor; and

(125) a culture product produced by a transformant according to the above (59) or (109).

As used herein the term "substantial equivalent(s)" means that the activity of the protein, e.g., nature of the ligand binding activity, and physical characteristics are substantially the same. Substitutions, deletions or insertions of amino acids often do not produce radical changes in the physical and chemical characteristics of a polypeptide, in which case polypeptides containing the substitution,

deletion, or insertion would be considered to be substantially equivalent to polypeptides lacking the substitution, deletion, or insertion. Substantially equivalent substitutes for an amino acid within the sequence may be selected from other members of the class to which the amino acid belongs.

The non-polar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (HS-1) having a nucleotide sequence represented by SEQ ID NO: 1 with the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 2 depicts the community (homology) of the sequence which is complementary to 3' side synthetic DNA primers (HS-2) having a nucleotide sequence represented by SEQ ID NO: 2 with the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 3 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (3A) having a nucleotide sequence represented by SEQ ID NO: 5 or 5' side synthetic DNA primers (3B) having a nucleotide sequence represented by SEQ ID NO: 6 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 4 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (3C) having a nucleotide sequence represented by SEQ ID NO: 7 or 5' side synthetic DNA primers (3D) having a nucleotide sequence represented by SEQ ID NO: 3 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs

and genes.

Figure 5 depicts the community (homology) of the sequence (6A) which is complementary to 3' side synthetic DNA primers having a nucleotide sequence represented by SEQ ID NO: 8 or the nucleotide sequence (6B) which is complementary to 3' side synthetic DNA primers having a nucleotide sequence represented by SEQ ID NO: 9 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 6 depicts the community (homology) of the sequence (6C) which is complementary to 3' side synthetic DNA primers having a nucleotide sequence represented by SEQ ID NO: 4 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 7 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (T2A) having a nucleotide sequence represented by SEQ ID NO: 10 with the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 8 depicts the community (homology) of the sequence which is complementary to 3' side synthetic DNA primers (T7A) having a nucleotide sequence represented by SEQ ID NO: 11 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 9 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (TM1-A2) having a nucleotide sequence represented by SEQ ID NO: 12 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 10 depicts the community (homology) of the sequence which is complementary to 3' side synthetic DNA primers (TM3-B2) having a nucleotide sequence represented by SEQ ID NO: 13 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 11 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (TM3-C2) having a nucleotide sequence represented by SEQ ID NO: 14 relative to

the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 12 depicts the community (homology) of the sequence which is complementary to 3' side synthetic DNA primers (TM6-E2) having a nucleotide sequence represented by SEQ ID NO: 15 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 13 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (TM2F18) having a nucleotide sequence represented by SEQ ID NO: 16 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 14 depicts the community (homology) of the sequence which is complementary to 3' side synthetic DNA primers (TM6R21) having a nucleotide sequence represented by SEQ ID NO: 17 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 15 depicts the community (homology) of the sequence of 5' side synthetic DNA primers (S3a) having a nucleotide sequence represented by SEQ ID NO: 18 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 16 depicts the community (homology) of the sequence which is complementary to 3' side synthetic DNA primers (S6a) having a nucleotide sequence represented by SEQ ID NO: 19 relative to the nucleotide sequence each of other G protein coupled receptor protein-encoding cDNAs and genes.

Figure 17 is the 1.2% agarose gel electrophoresis profile of cDNA products each obtained from human brain amygdala (1, 2, 7), human pituitary body (3, 4, 8) and rat brain (5, 6, 9) by PCR amplification using the synthetic DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and the synthetic DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, wherein lanes 1 to 6 show the results of when PCR is carried out under severe conditions as disclosed in Examples, lanes 7 to 9 show the results of when PCR is carried out under mild conditions, and M denotes a size

marker which is obtained by cutting λ -phage DNA with restriction enzyme, EcoRI4I.

Figure 18 shows the nucleotide sequence determined by sequencing of clone A58 with a T7 primer wherein the clone A58 is obtained by amplifying human brain amygdala-derived cDNA by PCR under mild conditions and subcloning it to PCRTM II.

Figure 19 shows the nucleotide sequence determined by sequencing of clone A58 with an SP6 primer.

Figure 20 shows the nucleotide sequence determined by sequencing of clone 57-A-2 by using a -21M13 primer wherein the clone 57-A-2 is obtained by amplifying human brain amygdala-derived cDNA by PCR under severe conditions and subcloning it to PCRTM II.

Figure 21 shows the nucleotide sequence determined by sequencing of clone B54 with a T7 primer wherein the clone B54 is obtained by amplifying rat whole brain-derived cDNA by PCR under mild conditions and subcloning it to PCRTM II.

Figure 22 illustrates the nucleotide sequence of the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in the cDNA clone p19P2 isolated by PCR using a human pituitary gland-derived cDNA and the amino acid sequence encoded thereby, wherein the primer used for sequencing is -21M13, and the underlined part corresponds to the synthetic primer.

Figure 23 illustrates the nucleotide sequence of the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in the cDNA clone p19P2 isolated by PCR using a human pituitary gland-derived cDNA and the amino acid sequence encoded thereby, wherein the primer used for sequencing is M13RV-N (Takara, Japan), and the underlined part corresponds to the synthetic primer.

Figure 24 is the partial hydrophobicity plotting profile of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2, prepared based upon the amino acid sequence

shown in Figure 22.

Figure 25 is the partial hydrophobicity plotting profile of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2, prepared based upon the amino acid sequence shown in Figure 23.

Figure 26 shows the partial amino acid sequence (p19P2) of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2, as shown in Figures 22 and 23, relative to the known G protein coupled receptor protein, SI2863, wherein reverse amino acid residues are in agreement, the 1st to 99th amino acid residues of the p19P2 sequence correspond to the 1st to 99th amino acid residues in Figure 22, and the 156th to 230th amino acid residues thereof correspond to the 1st to 68th amino acid residues in Figure 23.

Figure 27 is the nucleotide sequence of the MIN6-derived G protein coupled receptor protein cDNA fragment derived based upon the nucleotide sequences of the MIN6-derived G protein coupled receptor protein cDNA fragments each included in the cDNA clones, pG3-2 and pG1-10, isolated by PCR using a MIN6-derived cDNA and the amino acid sequence encoded thereby, wherein the underlined parts corresponds to the synthetic primers.

Figure 28 is the partial hydrophobicity plotting profile of the MIN6-derived G protein coupled receptor protein, prepared based upon the partial amino acid sequence shown in Figure 27.

Figure 29 is the partial nucleotide sequence of the novel receptor protein cDNA clone, p63A2, obtained from the human amygdaloid nucleus by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined part corresponds to the synthetic primer.

Figure 30 is the partial nucleotide sequence of the novel receptor protein cDNA clone, p63A2, obtained from the human amygdaloid nucleus by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined part

corresponds to the synthetic primer.

Figure 31 is the hydrophobicity plotting profile, prepared based upon the amino acid sequence shown in Figure 29, suggesting the presence of hydrophobic domains as designated by 1 to 3.

Figure 32 is the hydrophobicity plotting profile, prepared based upon the amino acid sequence shown in Figure 30, suggesting the presence of hydrophobic domains as designated by 4 to 6.

Figure 33 is the partial amino acid sequence (p63A2) of the protein encoded by the novel receptor protein cDNA fragment included in p63A2, relative to the partial amino acid sequence of the G protein coupled receptor protein (p30731) expressed and induced by a mouse T cell-derived glucocorticoid, wherein reverse amino acid residues are in agreement.

Figure 34 is the whole nucleotide sequence of the human pituitary gland-derived G protein coupled receptor protein cDNA, included in the cDNA clone, pGHR3, isolated from the human-derived cDNA library by plaque hybridization using an DNA insert in the p19P2 as a probe, and the amino acid sequence encoded thereby.

Figure 35 is the northern blotting profile of the human pituitary gland mRNA of the receptor gene encoded by the human pituitary gland-derived cDNA clone, pGHR3.

Figure 36 is the hydrophobicity plotting profile of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA included in pGHR3, prepared based upon the amino acid sequence shown in Figure 34.

Figure 37 is the partial nucleotide sequence of the novel receptor protein cDNA clone, p3H2-17, obtained from mouse pancreatic β -cell strain, MIN6, by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined part corresponds to the synthetic primer used for the PCR amplification.

Figure 38 is the hydrophobicity plotting profile, prepared based upon the amino acid sequence shown in Figure 37, suggesting the presence of hydrophobic domains as designated

by 3 to 6.

Figure 39 is the partial amino acid sequence encoded by the novel receptor protein cDNA included in p3H2-17, relative to the partial amino acid sequence each of chicken ATP receptor protein (P34996), human somatostatin receptor subtype 3 protein (A46226), human somatostatin receptor subtype 4 protein (JN0605) and bovine neuropeptide Y receptor protein (528787), wherein reverse amino acid residues are in agreement.

Figure 40 is the partial nucleotide sequence of the novel receptor protein cDNA clone, p3H2-34, obtained from mouse pancreatic β -cell strain, MIN6, by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined parts correspond to the synthetic primers used for the PCR amplification.

Figure 41 is the hydrophobicity plotting profile, prepared based upon the amino acid sequence shown in Figure 40, wherein the axis of ordinate represents an index of hydrophobicity, the axis of abscissa represents the number of amino acids and numerals 3 to 6 represent the presence of hydrophobic domains.

Figure 42 is the partial amino acid sequence encoded by the novel receptor protein cDNA included in p3H2-34, relative to the partial amino acid sequence each of human somatostatin receptor subtype 4 protein (JN0605), human somatostatin receptor subtype 2 protein (B41795) and rat-derived ligand unknown receptor protein (A39297), wherein reverse amino acid residues are in agreement.

Figure 43 is the nucleotide sequence of the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, pMD4, obtained from rabbit gastropyrolic part smooth muscles by PCR amplification, and the amino acid sequence encoded thereby, wherein the underlined parts correspond to the synthetic primers used for the PCR amplification.

Figure 44 is the hydrophobicity plotting profile of the protein encoded by the rabbit gastropyrolic part smooth

muscle-derived G protein coupled receptor protein cDNA fragment included in pMD4, prepared based upon the amino acid sequence shown in Figure 35, wherein numerals 1 to 3 suggest the presence of hydrophobic domains.

Figure 45 is the partial amino acid sequence (pMD4) of the protein encoded by the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in pMD4 as shown in Figure 43, relative to the known G protein coupled receptor protein, rat ligand unknown receptor protein (A35639), wherein reverse amino acid residues are in agreement, the 1st to 88th amino acid residues of the pMD4 sequence correspond to the 1st to 88th amino acid residues in Figure 43.

Figure 46 shows the nucleotide sequence of the mouse-derived galanin receptor protein cDNA clone, pMGR20, which has been cloned with, as a probe, the cDNA insert in p3H2-34 and the amino acid sequence encoded thereby.

Figure 47 is the hydrophobicity plotting profile, prepared based upon the amino acid sequence shown in Figure 46, wherein the axis of ordinate represents an index of hydrophobic property, the axis of abscissa represents the number of amino acids, and numerals 1 to 7 represent the presence of hydrophobic domains.

Figure 48 is the amino acid sequence (MOUSEGALRECE) of the mouse-derived galanin receptor protein encoded by pMGR20, relative to the amino acid sequence (HUMAN GALAMI) of the human-derived galanin receptor protein, wherein reverse amino acid residues are in agreement.

Figure 49 is the nucleotide sequence of the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, pMJ10, obtained from rabbit gastropyrolic part smooth muscles by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined parts correspond to the synthetic primers used for the PCR amplification.

Figure 50 is the hydrophobicity plotting profile of

the protein encoded by the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in pMJ10, prepared based upon the amino acid sequence shown in Figure 49, wherein numerals 4 to 6 suggest the presence of hydrophobic domains.

Figure 51 is the partial amino acid sequence (pMJ10) of the protein encoded by the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in pMJ10 shown in Figure 49, relative to human ligand unknown receptor protein (B42009), human N-formylpeptide receptor protein (JC2014), rabbit N-formylpeptide receptor protein (A46520), mouse C5a anaphylatoxin receptor protein (A46525) and bovine neurotensin Y receptor protein (S28787) which are known G protein coupled receptor proteins, wherein reverse amino acid residues are in agreement, and the 1st to 125th amino acid residues of pMJ10 correspond to the 1st to 125th amino acid residues in Figure 49.

Figure 52 is the nucleotide sequence of the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, pMH28, obtained from rabbit gastropyrolic part smooth muscles by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined parts correspond to the synthetic primers used for the PCR amplification.

Figure 53 is the hydrophobicity plotting profile of the protein encoded by the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in pMH28, prepared based upon the amino acid sequence shown in Figure 52, wherein numerals 4 to 6 suggest the presence of hydrophobic domains.

Figure 54 is the partial amino acid sequence (pMH28) of the protein encoded by the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in pMH28 shown in Figure 52, relative to mouse IL-8 receptor protein (P35343), human somatostatin receptor protein 1 (A41795) and human somatostatin receptor protein 4 (A47457)

which are known G protein coupled receptor proteins, wherein reverse amino acid residues are in agreement, and the 1st to 119th amino acid residues of pMH28 correspond to the 1st to 119th amino acid residues in Figure 52.

Figure 55 is the nucleotide sequence of the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, pMN7, obtained from rabbit gastropyrolic part smooth muscles by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined 5'-end nucleotide sequence part corresponds to the synthetic primer used for the PCR amplification.

Figure 56 is the nucleotide sequence of the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, pMN7, obtained from rabbit gastropyrolic part smooth muscles by PCR amplification and the amino acid sequence encoded thereby, wherein the underlined 3'-end nucleotide sequence part corresponds to the synthetic primer used for the PCR amplification.

Figure 57 is the hydrophobicity plotting profile of the protein encoded by the rabbit gastropyrolic part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in pMN7, prepared based upon the amino acid sequence shown in Figures 55 and 56, wherein numerals TM2 to TM6 suggest the presence of hydrophobic domains.

Figure 58 is the partial hydrophobicity plotting profile of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2, prepared based upon the amino acid sequence shown in Figure 22.

Figure 59 is the partial hydrophobicity plotting profile of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2, prepared based upon the amino acid sequence shown in Figure 23.

Figure 60 shows the partial amino acid sequence

- 7 1 -

(p19P2) of the protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2, as shown in Figures 22 and 23, relative to the known G protein coupled receptor protein, S12863, wherein reverse amino acid residues are in agreement, the 1st to 99th amino acid residues of the p19P2 sequence correspond to the 1st to 99th amino acid residues in Figure 22, and the 156th to 230th amino acid residues thereof correspond to the 1st to 68th amino acid residues in Figure 23.

Figure 61 is the partial amino acid sequence (pG3-2/pG1-10) of the MIN6-derived G protein coupled receptor protein, as shown in Figure 27, relative to the partial amino acid sequence (p19P2) of the protein encoded by p19P2, as shown in Figures 22 and 23, wherein reverse amino acid residues are in agreement, the 1st to 99th amino acid residues of the p19P2 sequence correspond to the 1st to 99th amino acid residues in Figure 22, the 156th to 223rd amino acid residues thereof correspond to the 1st to 68th amino acid residues in Figure 23, and the 1st to 223rd amino acid residues of the pG3-2/pG1-10 sequence correspond to the 1st to 223rd amino acid residues in Figure 27.

Figure 62 is the nucleotide sequence of the MIN6-derived G protein coupled receptor protein cDNA fragment included in the cDNA clone, p5S38, isolated by PCR using a MIN6-derived cDNA and the amino acid sequence encoded thereby, wherein the underlined parts corresponds to the synthetic primers.

Figure 63 is the partial amino acid sequence (p5S38) of the MIN6-derived G protein coupled receptor protein, as shown in Figure 62, relative to the partial amino acid sequence (p19P2) of the G protein coupled receptor protein encoded by p19P2, as shown in Figures 22 and 23, as well as the partial amino acid sequence of the G protein coupled receptor protein encoded by the nucleotide sequence derived from the nucleotide sequence of the cDNA fragment included in pG3-2 and pG1-10, as shown in Figure 27, wherein reverse amino acid residues are in agreement, the 1st to 144th amino acid residues of the p5S38

- 7 2 -

sequence correspond to the 1st to 144th amino acid residues in Figure 62, the 1st to 99th amino acid residues of the p19P2 sequence correspond to the 1st to 99th amino acid residues in Figure 22, the 156th to 223rd amino acid residues thereof correspond to the 1st to 68th amino acid residues in Figure 23, and the 1st to 223rd amino acid residues of the pG3-2/pG1-10 sequence correspond to the 1st to 223rd amino acid residues in Figure 27.

Figure 64 is the partial hydrophobicity plotting profile of the protein encoded by the MIN6-derived G protein coupled receptor protein cDNA fragment included in p5S38, prepared based upon the amino acid sequence shown in Figure 62.

Figure 65 shows the northern blot analysis profile of the receptor gene encoded by the cDNA included in the mouse pancreatic β -cell strain MIN6-derived novel receptor protein cDNA clone, p3H2-17, for mouse cell line, MIN6, Neuro-2a cell and mouse brain, thymus, spleen and pancreas poly(A)⁺ RNA, wherein each arrow and number indicates the size marker position (unit of number: kb).

Figure 66 shows the agarose gel electrophoresis analysis profile of the PCR products obtained by 5'RACE PCR of the receptor gene included in p3H2-17 using mouse thymus and spleen poly(A)⁺ RNA.

Lane 1 indicates the size marker 6 (Wako Pure Chemical, Japan).

Lane 2 indicates the internal control which is the thymus-derived PCR product obtained by PCR amplification using the primer having SEQ ID NO: 20 and the primer having SEQ ID NO: 22 with Taq polymerase.

Lane 3 indicates the negative control which is the PCR product obtained by Ex Taq polymerase PCR amplification of thymus cDNA prior to addition of anchors.

Lane 4 indicates the negative control which is the PCR product obtained by Taq polymerase PCR amplification of thymus cDNA prior to addition of anchors.

Lane 5 indicates the PCR product obtained by 5'RACE of thymus poly(A)⁺ RNA with Pfu polymerase.

Lane 6 indicates the PCR product obtained by 5'RACE of thymus poly(A)⁺ RNA with Vent polymerase.

Lane 7 indicates the PCR product obtained by 5'RACE of thymus poly(A)⁺ RNA with Ex Taq polymerase.

Lane 8 indicates the PCR product obtained by 5'RACE of thymus poly(A)⁺ RNA with Taq polymerase.

Lane 9 indicates the size marker 5 (Wako Pure Chemical, Japan).

Lane 10 indicates the internal control which is the spleen-derived PCR product obtained by PCR amplification using the primer having SEQ ID NO: 20 and the primer having SEQ ID NO: 22 with Taq polymerase.

Lane 11 indicates the negative control which is the PCR product obtained by Ex Taq polymerase PCR amplification of spleen cDNA prior to addition of anchors.

Lane 12 indicates the negative control which is the PCR product obtained by Taq polymerase PCR amplification of spleen cDNA prior to addition of anchors.

Lane 13 indicates the PCR product obtained by 5'RACE of poly(A)⁺ RNA with Pfu polymerase.

Lane 14 indicates the PCR product obtained by 5'RACE of spleen poly(A)⁺ RNA with Vent polymerase.

Lane 15 indicates the PCR product obtained by 5'RACE of spleen poly(A)⁺ RNA with Ex Taq polymerase.

Lane 16 indicates the PCR product obtained by 5'RACE of spleen poly(A)⁺ RNA with Taq polymerase.

Lane 17 indicates the size marker 5 (Wako Pure Chemical, Japan).

Each blacked triangle indicates the band recovered.

Figure 67 shows the agarose gel electrophoresis analysis profile of the PCR products obtained by 3'RACE PCR of the receptor gene included in p3H2-17 using mouse thymus and spleen poly(A)⁺ RNA.

Lane 1 indicates the size marker 5 (Wako Pure Chemical, Japan).

Lane 2 indicates the PCR product obtained by 3'RACE of spleen poly(A)⁺ RNA with Taq polymerase.

Lane 3 indicates the PCR product obtained by 3'RACE of spleen poly(A)⁺ RNA with Ex Taq polymerase.

Lane 4 indicates the PCR product obtained by 3'RACE of spleen poly(A)⁺ RNA with Vent polymerase.

Lane 5 indicates the PCR product obtained by 3'RACE of spleen poly(A)⁺ RNA with Pfu polymerase.

Lane 6 indicates the PCR product obtained by 3'RACE of thymus poly(A)⁺ RNA with Taq polymerase.

Lane 7 indicates the PCR product obtained by 3'RACE of thymus poly(A)⁺ RNA with Ex Taq polymerase.

Lane 8 indicates the PCR product obtained by 3'RACE of thymus poly(A)⁺ RNA with Vent polymerase.

Lane 9 indicates the PCR product obtained by 3'RACE of thymus poly(A)⁺ RNA with Pfu polymerase.

Lane 10 indicates the size marker 6 (Wako Pure Chemical, Japan).

Each blacked triangle indicates the band recovered.

Figure 68 depicts the model of the RACE products of the receptor protein cDNA fragment included in p3H2-17 obtained by 5'RACE and 3'RACE. Open squares represent regions which have already been isolated and included in p3H2-17. Small arrows, ①, ②, ③ and ④, indicate the positions and directions of the primers designed in Working Example 19. The big arrow shows a predicted full-length open reading frame of the receptor protein held by p3H2-17. Numbers at both ends, N26, N64, N75, C2, C13 and C15, indicate clone numbers of the RACE products obtained. Among these RACE products, N26, N64 and N75 are inserted into PCRTM II vector and C2, C13 and C15 are inserted into the SmaI site of pUC18. The solid triangle indicates the PCR error position which has been clarified through sequencing.

Figure 69 is the nucleotide sequence of the open reading frame and neighboring regions thereof of mouse G protein coupled receptor protein cDNA included in the cDNA clone pMAH2-17 obtained from mouse spleen and thymus poly(A)⁺ RNA by RACE techniques based on the nucleotide sequence of the cDNA fragment included in p3H2-17 and the amino acid

sequence encoded thereby.

Figure 70 is the hydrophobicity plotting profile of the protein encoded by the receptor protein cDNA included in pMAH2-17, prepared based upon the amino acid sequence shown in Figure 69.

Figure 71 is the amino acid sequence (75+13CODING) of the protein encoded by the mouse-derived G protein coupled receptor protein cDNA fragment included in pMAH2-17, as shown in Figure 69, relative to the known G protein coupled receptor proteins, mouse P_{2U} purinoceptor (P2UR MOUSE) and chicken P_{2Y} purinoceptor (P2YR CHICK), wherein reverse amino acid residues are in agreement.

Figure 72 is the nucleotide sequence (from 1st to 540th nucleotides) of the rabbit gastropyloric part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, PMN128, obtained from rabbit gastropyloric part smooth muscles by PCR amplification, and the amino acid sequence encoded thereby, wherein the underlined 5' part corresponds to the synthetic primer used for the PCR amplification.

Figure 73 is the nucleotide sequence (from 541st to 843rd nucleotides) of the rabbit gastropyloric part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in the novel receptor protein cDNA clone, PMN128, obtained from rabbit gastropyloric part smooth muscles by PCR amplification, and the amino acid sequence encoded thereby, wherein the underlined 3' part corresponds to the synthetic primer used for the PCR amplification.

Figure 74 is the hydrophobicity plotting profile of the protein encoded by the rabbit gastropyloric part smooth muscle-derived G protein coupled receptor protein cDNA fragment included in PMN128, prepared based upon the amino acid sequences shown in Figures 72 and 73, suggesting the presence of hydrophobic domains.

Figure 75 shows inward currents evoked by ATP in *Xenopus* oocytes injected with cDNA of pMAH2-17-encoded receptor.

Figure 76 is the nucleotide sequence of the human-derived G protein coupled receptor protein cDNA fragment included in p3H2-17, relative to the nucleotide sequence of the mouse-derived G protein coupled receptor protein cDNA fragment included in p3H2-17, wherein reverse base residues are in agreement.

Figure 77 is the nucleotide sequence of the open reading frame and neighboring regions thereof of human-derived G protein coupled receptor protein cDNA included in pMAH2-17 and the amino acid sequence encoded thereby.

Figure 78 is the hydrophobicity plotting profile of the protein encoded by the human-derived G protein coupled receptor protein cDNA included in pMAH2-17.

Figure 79 is the amino acid sequence of human type purinoceptor encoded by pMAH2-17, relative to the mouse purinoceptor encoded by p3H2-17, wherein reverse amino acid residues are in agreement.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

According to the present invention, DNA sequences comprising each a nucleotide sequence indicated by a SEQ ID NO selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 19 have been synthesized and characterized. The DNA is a potent primer for polymerase chain reaction in order to amplify DNA sequences encoding part or all of the polypeptide sequence of G protein coupled receptor protein. PCR amplification methods of the DNA coding for part or all of the polypeptide sequence of G protein coupled receptor protein can be advantageously carried out with the said primer DNA.

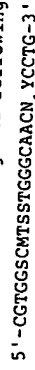
Screening of DNA libraries for the DNA encoding part or all of the polypeptide sequence of G protein coupled receptor protein can be successfully carried out through polymerase chain reaction techniques with the said primer DNA.

As a result, template DNAs coding for part or all of the polypeptide sequence of G protein coupled receptor protein, contained in the DNA library, can be selectively amplified and various DNA sequences encoding part or all of the

polypeptide sequence of G protein coupled receptor protein may be isolated and characterized. Further, G protein coupled receptor proteins, peptide segments or fragments derived from the G protein coupled receptor protein, modified derivatives or analogues thereof, and salts thereof may be recognized, predicted, deduced, produced, expressed, isolated and characterized.

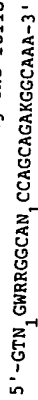
The primer DNA useful in PCR amplification of the DNA sequence encoding part or all of the polypeptide sequence of G protein coupled receptor protein is a degenerate deoxynucleotide which has an oligonucleotide sequence to which a SEQ ID NO selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 19 is assigned.

The nucleotide sequence represented by SEQ ID NO: 1 is a base sequence having the following formula:

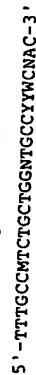


wherein S is G or C, M is A or C, N₁ = A, G, C, or T, and Y is T or C (Figure 1: HS-1).

The nucleotide sequence represented by SEQ ID NO: 2 (HS-2) is a base sequence having the following formula:

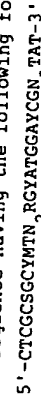


wherein N₁ = A, G, C, or T, W is A or T, R is A or G, and K is G or T, which is complementary to a nucleotide sequence having the following formula:



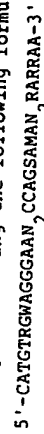
wherein N = A, C, G, or T, M is A or C, Y is T or C, and W is A or T (Figure 2).

The nucleotide sequence represented by SEQ ID NO: 3 is a base sequence having the following formula:



wherein S is G or C, Y is C or T, M is A or C, R is A or G, and N₂ = I (Figure 4: 3D).

The nucleotide sequence represented by SEQ ID NO: 4 is a base sequence having the following formula:



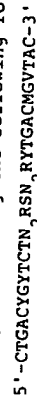
wherein R is A or G, W is T or A, S is G or C, M is A or C, and N₂ = I, which is complementary to a nucleotide sequence

having the following formula:



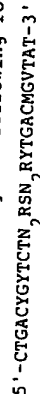
wherein Y is C or T, N₁ = A, G, C, or T, K is G or T, S is G or C, W is A or T (Figure 6: 6C).

The nucleotide sequence represented by SEQ ID NO: 5 is a base sequence having the following formula:



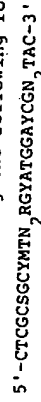
wherein Y is C or T, R is A or G, S is G or C, M is A or C, V is A, C or G, and N₂ is I (Figure 3: 3A).

The nucleotide sequence represented by SEQ ID NO: 6 is a base sequence having the following formula:



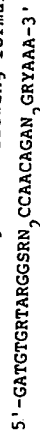
wherein Y is C or T, R is A or G, S is G or C, M is A or C, and V is A, C or G, and N₂ is I (Figure 3: 3B).

The nucleotide sequence represented by SEQ ID NO: 7 is a base sequence having the following formula:



wherein S is G or C, Y is C or T, M is A or C, R is A or G, and N₂ is I (Figure 4: 3C).

The nucleotide sequence represented by SEQ ID NO: 8 is a base sequence having the following formula:

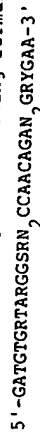


wherein R is A or G, S is G or C, Y is C or T, and N₂ is I, which is complementary to a nucleotide sequence having the following formula:



wherein R is A or G, Y is C or T, S is G or C, and N₁ is A, T, G, or C (Figure 5: 6A).

The nucleotide sequence represented by SEQ ID NO: 9 is a base sequence having the following formula:



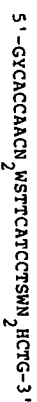
wherein R is A or G, S is G or C, Y is C or T, and N₂ is I, which is complementary to a nucleotide sequence having the following formula:



wherein R is A or G, Y is C or T, S is G or C, and N₁ is A,

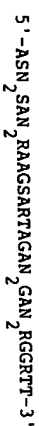
T, G, or C (Figure 5: 6B).

The nucleotide sequence represented by SEQ ID NO: 10 is a base sequence having the following formula:

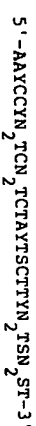


wherein S is G or C, Y is C or T, W is A or T, H is A, C or T, and N₂ is I (Figure 7: T2A).

The nucleotide sequence represented by SEQ ID NO: 11 (Figure 8: T7A) is a base sequence having the following formula:

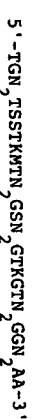


wherein R is A or G, S is G or C, and N₂ is I, which is complementary to a nucleotide sequence having the following formula:



wherein Y is C or T, N₂ is I, and S is G or C (Figure 8).

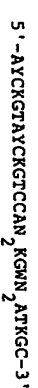
The nucleotide sequence represented by SEQ ID NO: 12 is a base sequence having the following formula:



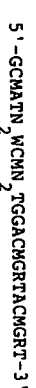
wherein S is G or C, K is G or T, M is A or C, and N₂ is I

(Figure 9: TM1-A2).

The nucleotide sequence represented by SEQ ID NO: 13 (Figure 10: TM3-B2) is a base sequence having the following formula:

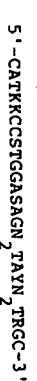


wherein Y is C or T, K is G or T, W is A or T, and N₂ is I, which is complementary to a nucleotide sequence having the following formula:



wherein M is A or C, W is A or T, R is A or G, and N₂ is I (Figure 10).

The nucleotide sequence represented by SEQ ID NO: 14 is a base sequence having the following formula:



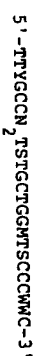
wherein K is G or T, S is G or C, Y is C or T, R is A or G, and N₂ is I (Figure 11: TM3-C2).

The nucleotide sequence represented by SEQ ID NO: 15 (Figure 12: TM6-E2) is a base sequence having the following

formula:

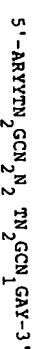


wherein W is A or T, S is G or C, K is G or T, R is A or G, and N₂ is I, which is complementary to a nucleotide sequence having the following formula:



wherein Y is C or T, S is G or C, W is A or C, W is A or T, and N₂ is I (Figure 12).

The nucleotide sequence represented by SEQ ID NO: 16 is a base sequence having the following formula:

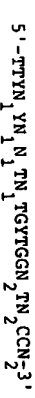


wherein R is A or G, Y is C or T, N₁ is A, T, G, or C, and N₂ is I (Figure 13: TM2F18).

The nucleotide sequence represented by SEQ ID NO: 17 (Figure 14: TM6R21) is a base sequence having the following formula:



wherein R is A or G, N₁ is A, T, G, or C, and N₂ is I which is complementary to a nucleotide sequence having the following formula:



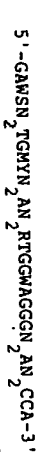
wherein Y is C or T, N₁ is A, T, G, or C, and N₂ is I (Figure 14).

The nucleotide sequence represented by SEQ ID NO: 18 is a base sequence having the following formula:

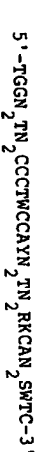


wherein S is G or C, R is A or G, W is A or T, M is A or C, and N₂ is I (Figure 15: S3A).

The nucleotide sequence represented by SEQ ID NO: 19 (Figure 16: S6A) is a base sequence having the following formula:



wherein W is A or T, S is G or C, M is A or C, Y is C or T, R is A or G, and N₂ is I, which is complementary to a nucleotide sequence having the following formula:



wherein W is A or T, Y is C or T, R is A or G, K is G or T, and S is G or C (Figure 16).

In a specific embodiment, symbols in the aforementioned SEQ ID NOs (R, Y, M, K, S, W, H, V and N) indicate the incorporation of plural bases, leading to multiple oligonucleotides in the primer preparation. In other words, SEQ ID NO: 1 to SEQ ID NO: 19 are degenerate nucleotide primers.

The nucleotide sequence represented by SEQ ID NO: 1 (Figure 1: HS-1) is a nucleotide sequence highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the first membrane-spanning (transmembrane) domain each of known G protein coupled receptor proteins such as human-derived TRH receptor protein (HTRHR), human-derived RANTES receptor protein (L10918, HUMRANTES), human Burkitt's lymphoma-derived receptor protein with an unknown ligand (X68149, HSBRL1A), human-derived somatostatin receptor protein (L14856, HUNSOMAT0), rat-derived μ -opioid receptor protein (U02083, RNU02083), rat-derived κ -opioid receptor protein (U00442, U00442), human-derived neuromedin B receptor protein (W73482, HUMNMBR), human-derived muscarinic acetylcholine receptor protein (X15266, HSHM4), rat-derived adrenaline α_1 B receptor protein (L08609, RATAADRE01), human-derived somatostatin 3 receptor protein (M96738, HUMSSTR3X), human-derived C_5 receptor protein (HUMCSAAR), human-derived receptor protein with an unknown ligand (HUMRDC1A), human-derived receptor protein with an unknown ligand (M84605, HUMOP1ODRE), rat-derived adrenaline α_2 B receptor protein (M91466, RATA2BAR) and the like [Figure 1].

The nucleotide sequence represented by SEQ ID NO: 2 (HS-2) is a nucleotide sequence which is complementary to the nucleotide sequence (Figure 2) highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of known G protein coupled receptor proteins such as mouse-derived receptor

protein with an unknown ligand (M80481, MUSGIR), human-derived bombesin receptor protein (L08893, HUMBOMB3S), human-derived adenosine A2 receptor protein (S46950, S46950), mouse-derived receptor protein with an unknown ligand (D21061, MUSGPCR), mouse-derived TRH receptor protein (S43387, S43387), rat-derived neuromedin K receptor protein (J05189, RATNEURA), rat-derived adenosine A1 receptor protein (M69045, RATAJARA), human-derived neurokinin A receptor protein (M57414, HUMNEKA), rat-derived adenosine A3 receptor protein (M94152, RATADENRE), human-derived somatostatin 1 receptor protein (M81829, HUMSTR1A), human-derived neurokinin 3 receptor protein (S86390, S86371S4), rat-derived receptor protein with an unknown ligand (X61496, RNCGPCR), human-derived somatostatin 4 receptor protein (L07061, HUMSSTR4Z), rat-derived GNRH receptor protein (M31670, RATGNRHA) and the like [Figure 2].

The nucleotide sequence represented by SEQ ID NO: 5 (Figure 3: 3A) or the nucleotide sequence represented by SEQ ID NO: 5 ID NO: 6 (Figure 3: 3B) is a nucleotide sequence highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the third membrane-spanning domain each of known G protein coupled receptors such as mouse-derived κ -opioid receptor protein (L11064), mouse-derived δ -opioid receptor protein (L11065), rat-derived μ -opioid receptor protein (D16349), mouse-derived bradykinin-B2 receptor protein (X69676), rat-derived bradykinin-B2 receptor protein (M59967), mouse-derived bombesin receptor protein (M35328), human-derived neuromedin B receptor protein (W73482), human-derived gastrin releasing peptide receptor protein (W73481), human-derived bombesin receptor protein subtype 3 (L08893), mouse-derived substance K receptor protein (X62933), mouse-derived substance P receptor protein (X62934), rat-derived neurokinin 3 receptor protein (J05189), rat-derived endothelin receptor protein (M60786), rat-derived receptor protein with an unknown ligand (L04672), rat-derived receptor protein with an unknown ligand (X61496), rat-derived receptor protein with an unknown ligand (X59249), rat-derived receptor protein with an unknown ligand (L09249),

mouse-derived receptor protein with an unknown ligand (P30731), human-derived receptor protein with an unknown ligand (M31210), human-derived receptor protein with an unknown ligand (U03642) and the like [Figure 3].

5 The nucleotide sequence represented by SEQ ID NO: 7 (Figure 4: 3C) or the nucleotide sequence represented by SEQ ID NO: 3 (Figure 4: 3D) is a nucleotide sequence highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the third membrane-spanning domain each of known G protein coupled receptors such as mouse-derived angiotensin II receptor protein (L32840), rat-derived angiotensin II receptor protein (X64052), rat-derived angiotensin receptor protein subtype (M90065), human-derived angiotensin Ia receptor protein (M91664), rat-derived cholecystokinin a receptor protein (M88096), rat-derived cholecystokinin b receptor protein (M99418), human-derived cholecystokinin b receptor protein (L04473), mouse-derived low affinity interleukin 8 receptor protein (M73969), human-derived high affinity interleukin 8 receptor protein (X65858), mouse-derived C5a anaphylatoxin receptor protein (S46665), human-derived N-formylpeptide receptor protein (M60626) and the like [Figure 4].

20 The nucleotide sequence represented by SEQ ID NO: 10 (Figure 7: T2A) is a nucleotide sequence highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the second membrane-spanning domain each of known G protein coupled receptors such as human galanin receptor (HUMGALAREC), rat α -1B-adrenergic receptor (RATADRI1B), human β -1-adrenergic receptor (HUMADRI1), rabbit IL-8 receptor (RABIL8RSB), human opioid receptor (HUMOPIDORE), bovine substance K receptor (BTSKR), human somatostatin receptor-2 (HUMSRI2A), human somatostatin receptor-3 (HUMSSTR3Y), human gastrin receptor (HUMGARE), human cholecystokinin A receptor (HUMCKAR), human dopamine receptor-D5 (HUMD1B), human serotonin receptor 5HT1E (HUM5HT1E), human dopamine receptor D4 (HUMD4C), mouse serotonin receptor-2 (MMSERO), rat α -1A-adrenergic receptor

(RATADRI1A), rat histamine H2 receptor (S57565) and the like [Figure 7].

5 The nucleotide sequence represented by SEQ ID NO: 8 (complementary to 6A of Figure 5) or the nucleotide sequence represented by SEQ ID NO: 9 (complementary to 6B of Figure 5) is a nucleotide sequence which is complementary to the nucleotide sequence (Figure 5) highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of known G protein coupled receptors such as mouse-derived κ -opioid receptor protein (L11065), rat-derived μ -opioid receptor protein (D16349), mouse-derived bradykinin B2 receptor protein (X69676), rat-derived bradykinin B2 receptor protein (M5967), mouse-derived bombesin receptor protein (M35328), human-derived neuromedin B receptor protein (M73482), human-derived gastrin releasing peptide receptor protein (M73481), human-derived bombesin receptor protein subtype 3 (L08893), mouse-derived substance K receptor protein (X62933), mouse-derived substance P receptor protein (X62934), rat-derived neurokinin 3 receptor protein (J05189), rat-derived endothelin receptor protein (M60786), rat-derived receptor protein with an unknown ligand (L04672), rat-derived receptor protein with an unknown ligand (X61496), rat-derived receptor protein with an unknown ligand (X59249), rat-derived receptor protein with an unknown ligand (L09249), mouse-derived receptor protein with an unknown ligand (P30731), human-derived receptor protein with an unknown ligand (M31210) human-derived receptor protein with an unknown ligand (U03642) and the like [Figure 5].

30 The nucleotide sequence represented by SEQ ID NO: 4 (complementary to 6C of Figure 6) is a nucleotide sequence which is complementary to the nucleotide sequence (Figure 6) highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of known G protein coupled receptors such as mouse-derived angiotensin II receptor protein (L32840),

rat-derived angiotensin Ib receptor protein (X64052),
 rat-derived angiotensin receptor protein subtype (M90065),
 human-derived angiotensin Ia receptor protein (M91464),
 rat-derived cholecystokinin a receptor protein (M88096),
 rat-derived cholecystokinin b receptor protein (M99418),
 human-derived cholecystokinin 8 receptor protein (L04473),
 mouse-derived low affinity interleukin 8 receptor protein
 (M73969), human-derived high affinity interleukin 8 receptor
 protein (X65858), mouse-derived C5a anaphylatoxin receptor
 protein (S46665), human-derived N-formylpeptide receptor
 protein (M60626) and the like [Figure 6].

The nucleotide sequence represented by SEQ ID NO: 11
 (Figure 8: T7A) is a nucleotide sequence which is

complementary to the nucleotide sequence (Figure 8) highly
 homologous to the DNA sequence coding for the amino acid
 sequence corresponding to or near the seventh

membrane-spanning domain each of known G protein coupled

receptors such as human galanin receptor (HUMGALAREC),

rat A1 adenosine receptor (RATIDREC), porcine angiotensin

receptor (PIGA2R), rat serotonin receptor (RAT5HTRC),

human dopamine receptor (S58541), human gastrin releasing

peptide receptor (HUMGRPR), mouse GRP/bombesin receptor

(MUSGRPBOB), rat vascular type 1 angiotensin receptor

(RRVTIATR), human muscarinic acetylcholine receptor (HSHM4),

human β -1 adrenergic receptor (HUMDRB1), human gastrin

receptor (HUMGARE), rat cholecystokinin receptor (RATCCKAR),

rat receptor with an unknown ligand (S59748), human

somatostatin receptor (HUMSST28A), rat receptor with an unknown

ligand (RNGPROCR), mouse somatostatin receptor-1 (MUSSTR1A),

human α -1-adrenergic receptor (HUMALADR), mouse

delta-opioid receptor (S66181), human somatostatin receptor-3

(HUMSSTR3Y) and the like [Figure 8].

The nucleotide sequence represented by SEQ ID NO: 12

(Figure 9: TM1-A2) is a nucleotide sequence highly homologous

to the DNA sequence coding for the amino acid sequence

within the first membrane-spanning (transmembrane) domain

each of known G protein coupled receptors such as

mouse-derived bradykinin B₂ receptor (MUSBB2R),
 bovine-derived substance K receptor (BTSKR), bovine-derived
 endothelin ET_B receptor (BOVEETBR), human-derived
 neuropeptide Y receptor (MMSUBKREC), human-derived
 prostaglandin E₂ receptor (HUMPE2R), human-derived
 prostacyclin receptor (HUMPIR), human-derived κ -opioid
 receptor (HSU11053), rat-derived melanocortin 3 receptor
 (RRMC3RA), human-derived melanocortin receptor (HUMMR),
 mouse-derived bombesin/GRP receptor (MUSGRPBOB),
 rat-derived cholecystokinin B receptor (RATCCKAR),
 rat-derived cholecystokinin A receptor (RATCCKAR) and the
 like [Figure 9].

The nucleotide sequence represented by SEQ ID NO: 13
 (Figure 10: TM3-B2) is a nucleotide sequence which

is complementary to the nucleotide sequence (Figure 10) highly

homologous to the DNA sequence coding for the amino acid

sequence corresponding to or near the end of the third

membrane-spanning domain of known G protein coupled receptors

such as human-derived cholecystokinin receptor (HUMCCKR),

human-derived cholecystokinin B receptor (HUMCCKBR),

mouse-derived melanocortin 5 receptor (MMGMC5R),

human-derived vasopressin receptor (HUMV2R), rat-derived

neuromedin K receptor (RATNEURA), dog-derived gastrin receptor

(DOGGSTRN), rat-derived serotonin receptor (RAT5HT5A),

mouse-derived α_2 -adrenaline receptor (MUSALP2ADA),

human-derived adenosine A₁ receptor (HUMADORA1X),

human-derived opioid (presumed) receptor (HUMOPIDORE),

mouse-derived bombesin/GRP receptor (MUSGRPBOB),

rat-derived cholecystokinin A receptor (RATCCKAR),

human-derived TRH receptor (HSTRHREC) and the like [Figure 10].

The nucleotide sequence represented by SEQ ID NO: 14

(Figure 11: TM3-C2) is a nucleotide sequence highly homologous

to the DNA sequence coding for the amino acid sequence

corresponding to or near the end of the third membrane-spanning

domain of known G protein coupled receptors such as

human-derived neurokinin 3 receptor (HUMNK3R), human-derived

oxytocin receptor (HSMRNOXY), guinea pig-derived

cholecystokinin A receptor (S68242), dog-derived cholecystokinin A receptor with an unknown ligand (CFGRCR4), mouse-derived substance P receptor (MMSUBPREC), human-derived receptor with an unknown ligand (HUMOPIDORE), human-derived galanin receptor (HUMGALAREC), human-derived serotonin receptor (HSS31G), human-derived β_3 -adrenaline receptor (HUMARB3A), human-derived prostacyclin receptor (HUMHPR), rat-derived cholecystokinin A receptor (RATCCRAR) and the like [Figure 11].

The nucleotide sequence represented by SEQ ID NO: 15 (Figure 12: TM6-E2) is a nucleotide sequence which is complementary to the nucleotide sequence (Figure 12) highly homologous to the DNA sequence coding for the amino acid sequence within the sixth membrane-spanning domain of known G protein coupled receptors such as human-derived neurokinin A receptor (HUMNEKAR), human-derived substance P receptor (HUMSUBPRA), rat-derived substance K receptor (RATSKR), mouse-derived bombesin/GRP receptor (MUSGRPBOM), human-derived opioid (presumed) receptor (HUMOPIDORE), human-derived adenosine A_2 receptor (HUMA2XXX), human-derived β_2 -adrenaline receptor (HUMADBR), canine-derived receptor RDC5 with an unknown ligand (CFGRCR8), human-derived endothelin receptor (HUMETSR), mouse-derived neuropeptide Y1 receptor (MMNPY1CDS), human-derived oxytocin receptor (HSMRNOXY), rat-derived cholecystokinin A receptor (RATCCRAR) and the like [Figure 12].

The nucleotide sequence represented by SEQ ID NO: 16 (Figure 13: TM2F18) is a nucleotide sequence highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the second membrane-spanning domain of known G protein coupled receptors such as human-derived TSH receptor (HUMTSHX), human-derived neurokinin A receptor (HUMNEKAR), human-derived FMLP receptor (HUMFMLP), human-derived IL8 receptor B (HUMINTLE08), human-derived α -1 adrenergic receptor (HUMALADR), human-derived IL8 receptor A (HUMIL8RA), human-derived dopamine D2 receptor (HSD2), human-derived angiotensin type I receptor (HUMANTIR),

human-derived somatostatin receptor (HUSOMAT), human-derived TRH receptor (HSTRHREC), human-derived delta-opioid receptor (HSU07882) and the like [Figure 13].

The nucleotide sequence represented by SEQ ID NO: 17 (Figure 14: TM6R21) is a nucleotide sequence which is complementary to the nucleotide sequence (Figure 14) highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of known G protein coupled receptors such as human-derived β -adrenergic receptor (HSBAR), human-derived neurokinin A receptor (HUMNEKAR), human-derived endothelin-1 receptor (HUMETN1R), human-derived histamine H_2 receptor (HUMHSH2R), human-derived α -1 adrenergic receptor (HUMALADR), human-derived IL8 receptor A (HUMIL8RA), human-derived neuromedin B receptor (HUMNMBR), human-derived neurokinin 1 receptor (HUMNK1RX), human-derived substance P receptor (HUMSUBPRA), human-derived 5-HT1D serotonin receptor (HUM5HT1DA), human-derived formylpeptide receptor (HUMFPR2A), human-derived dopamine D2 receptor (HSD2), human-derived neuropeptide Y receptor (HUMNEUYREC), human-derived adenosine A_2 receptor (HUMA2XXX), human-derived bradykinin receptor BK-2 (HUMBK2A), human-derived FMLP-related receptor II (HUMFMRPX), human-derived somatostatin receptor subtype 3 (HUMSTR3X), human-derived cholecystokinin receptor (HUMCCR), human-derived neurotensin receptor (HSNEURA) and the like [Figure 14].

The nucleotide sequence represented by SEQ ID NO: 18 (Figure 15: S3A) is a nucleotide sequence highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the third membrane-spanning domain of known G protein coupled receptors such as human-derived galanin receptor (HUMGALAREC), human-derived CCK-B receptor (S70057), human-derived ET_A receptor (S67127), human-derived ET_B receptor (S44866), human-derived CSA receptor (HUMCSAAR), human-derived angiotensin II receptor (HUMANTIR), human-derived bradykinin receptor (HUMBK2R), human-derived neurotensin receptor (HSNEURA), human-derived GRP receptor (HUMGRPR), human-derived somatostatin 5 receptor (HUMSRS),

human-derived IL-8 receptor (HUMIL8RA), human-derived neurokinin 2 (neurokinin A) receptor (HUMNEKAR) and the like [Figure 15].

The nucleotide sequence represented by SEQ ID NO: 19 (Figure 16: S6A) is a nucleotide sequence which is complementary to the nucleotide sequence (Figure 16) highly homologous to the DNA sequence coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of known G protein coupled receptors such as human-derived galanin receptor (HUMGLAREC), human-derived CCK-B receptor (S70057), human-derived ET_A receptor (S67127), human-derived ET_B receptor (S44866), human-derived C5A receptor (HUMC5AAR), human-derived angiotensin II receptor (HUMANITIR), human-derived bradykinin receptor (HUMBK2R), human-derived neurotensin receptor (HSNEURA), human-derived GRP receptor (HUMGRPR), human-derived somatostatin 5 receptor (HUMFSRS), human-derived IL-8 receptor (HUMIL8RA), human-derived neurokinin 2 (neurokinin A) receptor (HUMNEKAR) and the like [Figure 16].

The above-mentioned abbreviations in the parentheses are the identifiers (or reference numbers) which are shown when GenBank/EMBL Data Bank is searched using a DNASIS Gene/Protein Sequence Data Base (CD019; Hitachi Software Engineering, Japan) and are usually called "Accession Numbers" or "Entry Names".

HTRHR is, however, the sequence as described in Japanese Patent Application No. Hei 5-286986 (or No. 286986/1993) (EPA 638645).

The DNA (or nucleotides) of the present invention may be manufactured by DNA synthetic methods which are known per se or by methods similar thereto. The DNA (or nucleotides) of the present invention may be an oligonucleotide sequence having 8 to 60 base residues, preferably 12 to 50 base residues, more preferably 15 to 40 residues and most preferably 18 to 30 residues.

Among the DNAs of the present invention, the DNA having the nucleotide sequence represented by SEQ ID NO: 1 or

SEQ ID NO: 12 is a nucleotide sequence which is commonly present in the nucleotide sequence of the DNA encoding the amino acid sequence corresponding to or near the first membrane-spanning domain of the above-mentioned known G protein coupled receptor protein. Therefore, it can be complementarily bonded (i.e. is hybridizable) with RNA or DNA (including genome DNA, cDNA) coding for the amino acid sequence corresponding to or near the first membrane-spanning domain of known or unknown G protein coupled receptor proteins and, furthermore, it can be complementarily bonded (i.e. is hybridizable) with nucleotide sequences encoding other membrane-spanning domains as well.

The DNA having a nucleotide sequence represented by SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 14 or SEQ ID NO: 18 is a nucleotide sequence which is commonly present in the nucleotide sequence of the DNA coding for the amino acid sequence corresponding to or near the third membrane-spanning domain of the above-mentioned known G protein coupled receptor protein. Therefore, it can be complementarily bonded with RNA or DNA (including genome DNA, cDNA) coding for the part corresponding to or near the third membrane-spanning domain of known or unknown G protein coupled receptor proteins and, furthermore, it can be complementarily bonded with nucleotide sequences encoding other membrane-spanning domains as well.

The DNA having a nucleotide sequence represented by SEQ ID NO: 10 or SEQ ID NO: 16 is a nucleotide sequence which is commonly present in the nucleotide sequence of the DNA coding for the amino acid sequence corresponding to or near the second membrane-spanning domain of the above-mentioned known G protein coupled receptor protein. Therefore, it can be complementarily bonded with RNA or DNA (including genome DNA, cDNA) coding for the part corresponding to or near the second membrane-spanning domain of known or unknown G protein coupled receptor proteins and, furthermore, it can be complementarily bonded with nucleotide sequences encoding other membrane-spanning domains as well.

The DNA having a nucleotide sequence represented by SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 15, SEQ ID NO: 17 or SEQ ID NO: 19 is a nucleotide sequence which is commonly present in the nucleotide sequence of the DNA coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of the above-mentioned known G protein coupled receptor protein. Therefore, it can be complementarily bonded with RNA or DNA (including genome DNA, cDNA) coding for the part corresponding to or near the sixth membrane-spanning domain of known or unknown G protein coupled receptor proteins and, furthermore, it can be complementarily bonded with nucleotide sequences encoding other membrane-spanning domains as well.

The DNA having a nucleotide sequence represented by SEQ ID NO: 11 is a nucleotide sequence which is commonly present in the nucleotide sequence of the DNA coding for the amino acid sequence corresponding to or near the seventh membrane-spanning domain of the above-mentioned known G protein coupled receptor protein. Therefore, it can be complementarily bonded with RNA or DNA (including genome DNA, cDNA) coding for the part corresponding to or near the seventh membrane-spanning domain of known or unknown G protein coupled receptor proteins and, furthermore, it can be complementarily bonded with nucleotide sequences encoding other transmembrane domains as well.

The DNA having a nucleotide sequence represented by SEQ ID NO: 13 is a nucleotide sequence which is commonly present in the nucleotide sequence of the DNA coding for the amino acid sequence corresponding to or near the third membrane-spanning domain of the above-mentioned known G protein coupled receptor protein. Therefore, it can be complementarily bonded with RNA or DNA (including genome DNA, cDNA) coding for the part corresponding to or near the third membrane-spanning domain of known or unknown G protein coupled receptor proteins and, furthermore, it can be complementarily bonded with nucleotide sequences encoding other membrane-spanning domains as well.

Accordingly, the DNAs (or nucleotides) of the present invention can be used as DNA primers for a polymerase chain reaction (hereinafter, sometimes referred to as PCR).

For example:

- (i) a polymerase chain reaction is carried out by mixing
 - (1) a small amount of DNA (or DNA fragment(s)) which codes for G protein coupled receptor protein, said DNA (or DNA fragment(s)) acting as a template,
 - (2) at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1, DNA primers having a nucleotide sequence represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7, DNA primers having a nucleotide sequence represented by SEQ ID NO: 10, DNA primers having a nucleotide sequence represented by SEQ ID NO: 12, DNA primers having a nucleotide sequence represented by SEQ ID NO: 14, DNA primers having a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18 and
 - (3) at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 11, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19; or
- (ii) a polymerase chain reaction is carried out by mixing

(1) a small amount of DNA (or DNA fragment(s)) coding for G protein coupled receptor protein, said DNA (or DNA fragment(s)) acting as a template,

(2) at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12 and (3) at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 13

so that it is possible to amplify the target DNA (or DNA fragment(s)) coding for said receptor protein.

When the PCR is carried out using at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 11, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19, said DNA primer(s) is(are) bonded (hybridized) with the nucleotide sequence at the 3'-side of the + chain (plus chain) of template RNA or DNA (or fragment(s) thereof) coding for the sixth membrane-spanning domain or other membrane-spanning domains of G protein coupled receptor protein whereupon an elongation of the - chain (minus chain) proceeds in the 5' - 3' direction.

When the PCR is carried out using at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 11, said DNA primer is bonded with the nucleotide sequence at the 3'-side of the + chain (plus chain) of template RNA or DNA (or fragment(s) thereof) coding for the seventh membrane-spanning domain or other membrane-spanning domains of the G protein

coupled receptor protein whereupon an elongation of the - chain (minus chain) proceeds in the 5' - 3' direction.

When the PCR is carried out using at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12, said DNA primer is bonded with the nucleotide sequence at the 3'-side of the - chain (minus chain) of template RNA (or fragment(s) thereof) coding for the first membrane-spanning domain or other membrane-spanning domains of G protein coupled receptor protein whereupon an elongation of the + chain (plus chain) proceeds in the 5' - 3' direction.

When the PCR is carried out using at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 16, said DNA primer is bonded with the nucleotide sequence at the 3'-side of the - chain (minus chain) of template RNA (or fragment(s) thereof) coding for the second membrane-spanning domain or other membrane-spanning domains of G protein coupled receptor protein whereupon an elongation of the + chain (plus chain) proceeds in the 5' - 3' direction.

When the PCR is carried out using at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7, DNA primers having a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18, said DNA primer is bonded with the nucleotide sequence at the 3'-side of the - chain (minus chain) of template RNA or DNA (or fragment(s) thereof) coding for the third membrane-spanning domain or other membrane-spanning domains of G protein coupled receptor protein whereupon an elongation of the + chain (plus chain) proceeds in the 5' - 3' direction.

Accordingly, when the DNA primers having

nucleotide sequences represented by any of SEQ ID NO: 1 to SEQ ID NO: 19 of the present invention are used in combination each other, DNA (or DNA fragment(s)) coding for G protein coupled receptor protein can be successfully amplified.

One embodiment of the present invention provides:

(A) a method of amplifying DNA coding for the G protein coupled receptor protein (e.g., from the first to sixth membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19;

(B) a method of amplifying DNA coding for the G protein coupled receptor protein (e.g., from the first to seventh membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a

nucleotide sequence represented by SEQ ID NO: 12 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 11;

(C) a method of amplifying a DNA coding for the G protein coupled receptor protein (e.g., from the second to sixth membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 16 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19;

(D) a method of amplifying a DNA coding for the G protein coupled receptor protein (e.g., from the second to seventh membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 16 and

③ at least one DNA primer selected from the group

consisting of DNA primers having a nucleotide sequence

represented by SEQ ID NO: 11;

(E) a method of amplifying a DNA coding for the G protein coupled receptor protein (e.g., from the third to sixth membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence

represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a

nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7,

DNA primers having a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence

represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a

nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9,

DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence

represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19;

(F) a method of amplifying a DNA coding for the G protein coupled receptor protein (e.g., from the third to seventh membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence

represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7,

DNA primers having a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 11; and

(G) a method of amplifying a DNA coding for the G protein coupled receptor protein (e.g., from the first to third membrane-spanning (transmembrane) domains or other segments of the G protein coupled receptor protein), characterized in that a polymerase chain reaction is carried out by mixing

① a DNA coding for the G protein coupled receptor protein, said DNA acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence

represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 13.

An example of more preferred combination of the DNA primers in the amplification according to the above-mentioned (A) includes a combination of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 1 with a DNA primer having a nucleotide sequence represented by SEQ ID NO: 2 and the like.

An example of more preferred combination of the DNA primers in the amplification according to the above-mentioned (D) includes a combination of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 10 with a DNA primer having a nucleotide sequence represented by SEQ ID NO: 11 and the like.

An example of more preferred combination of the DNA

- 9 9 -

primers in the amplification according to the above-mentioned (E) includes:

(i) a combination of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 5 or a DNA primer having a nucleotide sequence represented by SEQ ID NO: 6 with a DNA primer having a nucleotide sequence represented by SEQ ID NO: 8 or a DNA primer having a nucleotide sequence represented by SEQ ID NO: 9;
 (ii) a combination of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 3 or a DNA primer having a nucleotide sequence represented by SEQ ID NO: 7 with a DNA primer having a nucleotide sequence represented by SEQ ID NO: 4 and the like.

An example of more preferred combination of the DNA primers in the amplification according to the above-mentioned (G) includes a combination of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 12 with a DNA primer having a nucleotide sequence represented by SEQ ID NO: 13 and the like.

The amplification may be carried out in accordance with known PCR techniques. For example, it may be carried out by the method described in Saiki, R. K. et al., Science, 239:487-491 (1988). Temperature, time, buffer, number of reaction cycles, enzyme such as DNA polymerase, addition of 2'-deoxy-7-deazaguanosine triphosphate or inosine, etc. in the PCR amplification may be suitably selected depending upon the type of target DNA and other factors.

When RNA is used as a template, PCR amplification may be carried out, for example, by the method described in Saiki, R. K. et al., Science, 239:487-491 (1988).

Moreover, the DNA having a nucleotide sequence represented by SEQ ID NO: 1 or SEQ ID NO: 12 of the present invention can be selectively and complementarily bonded (hybridized) with the nucleotide sequence at the 3'-side of the - chain of the DNA coding for the amino acid sequence corresponding to or near the first membrane-spanning domain of the G protein coupled receptor protein; the DNA having a nucleotide sequence represented by SEQ ID NO: 10 or SEQ ID

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NO: 16 of the present invention can be selectively and complementarily bonded (hybridized) with the nucleotide sequence at the 3'-side of the - chain of the DNA coding for the amino acid sequence corresponding to or near the second membrane-spanning domain of the G protein coupled receptor protein; the DNA having a nucleotide sequence represented by SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 14 or SEQ ID NO: 18 of the present invention can be selectively and complementarily bonded (hybridized) with the nucleotide sequence at the 3'-side of the - chain of the DNA coding for the amino acid sequence corresponding to or near the third membrane-spanning domain of the G protein coupled receptor protein; the DNA having a nucleotide sequence represented by SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 15, SEQ ID NO: 17 or SEQ ID NO: 19 of the present invention can be selectively and complementarily bonded (hybridized) with the nucleotide sequence at the 3'-side of the + chain of the DNA coding for the amino acid sequence corresponding to or near the sixth membrane-spanning domain of the G protein coupled receptor protein; the DNA having a nucleotide sequence represented by SEQ ID NO: 11 of the present invention can be selectively and complementarily bonded (hybridized) with the nucleotide sequence at the 3'-side of the + chain of the DNA coding for the amino acid sequence corresponding to or near the third membrane-spanning domain of the G protein coupled receptor protein; and the DNA having a nucleotide sequence represented by SEQ ID NO: 13 of the present invention can be selectively and complementarily bonded (hybridized) with the nucleotide sequence at the 3'-side of the + chain of the DNA coding for the amino acid sequence corresponding to or near the third membrane-spanning domain of the G protein coupled receptor protein and, accordingly, said DNA is also advantageously useful as a probe for screening DNA libraries for DNA (or DNA fragment(s)) encoding part or all of the polypeptide sequence of G protein coupled receptor proteins.

These screening methods for DNA (or DNA fragment(s))

encoding part or all of the polypeptide sequence of G protein coupled receptor proteins from the DNA library by using as a reagent, because it can be used as a probe the DNA of the present invention may be carried out according to DNA cloning methods known per se by those of skill in the art or methods similar thereto. Especially when the DNA of the present invention is used as a DNA primer for the PCR, both amplification and screening of the DNA (or DNA fragment) coding for the G protein coupled receptor protein can be conducted in a single step.

Thus, when the DNAs of the present invention are suitably combined and used as the DNA primer for the PCR, said DNA primer(s) is(are) bonded (hybridized) with RNA or DNA (or fragment(s) thereof) encoding the amino acid sequence of the first membrane-spanning (transmembrane) domain, the second membrane-spanning domain, the third membrane-spanning domain, the sixth membrane-spanning domain, the seventh membrane-spanning domain or other membrane-spanning domains of G protein coupled receptor proteins to amplify, for example,

- ① RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the first membrane-spanning to the sixth membrane-spanning domains of G protein coupled receptor proteins,
- ② RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the first membrane-spanning to the seventh membrane-spanning domains of G protein coupled receptor proteins,
- ③ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the third membrane-spanning to the sixth membrane-spanning domains of G protein coupled receptor proteins,
- ④ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the third membrane-spanning to the seventh membrane-spanning domains of G protein coupled receptor proteins,
- ⑤ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the second membrane-spanning to the

sixth membrane-spanning domains of G protein coupled receptor proteins or RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of other domains thereof,

- ⑥ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the second membrane-spanning to the seventh membrane-spanning domains of G protein coupled receptor proteins,

- ⑦ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of from the first membrane-spanning to the third membrane-spanning domains of G protein coupled receptor proteins or

- ⑧ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence of other domains of G protein coupled receptor proteins.

Through using the DNA primer according to the present invention, therefore, selective amplifications of:

- ① RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the first membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor proteins;

- ② RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the first membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor proteins;

- ③ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the third membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor proteins;

- ④ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the third membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor proteins;

- ⑤ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the second membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor proteins or RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering other areas thereof,

③ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the second membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor proteins;

④ RNA or DNA (or fragment(s) thereof) coding for the amino acid sequence covering from the first membrane-spanning domain to the third membrane-spanning domain of G protein coupled receptor proteins; and the like,

from DNA libraries can be successfully achieved.

Among the DNA primers of the present invention,

the combination of

① a DNA primer having a nucleotide sequence represented by SEQ ID NO: 1 or SEQ ID NO: 2; with

② at least one DNA primer selected from the group consisting of a DNA primer having a nucleotide sequence represented by

SEQ ID NO: 2, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 4, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 8, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 9, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 15, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 17 and a DNA primer having a nucleotide sequence represented by SEQ ID NO: 19;

is, unlike conventional primers, capable of selectively amplifying a broad area covering from the first membrane-spanning domain to the sixth membrane-spanning domain or other domains of G protein coupled receptor proteins.

Among the DNA primers of the present invention, the

combination of

① a DNA primer having a nucleotide sequence represented by SEQ ID NO: 1 or SEQ ID NO: 12; with

② a DNA primer having a nucleotide sequence represented by SEQ ID NO: 11;

is, unlike conventional primers, capable of selectively amplifying a broad area covering from the first membrane-spanning domain to the seventh membrane-spanning domain or other domains of G protein coupled receptor proteins.

Among the DNA primers of the present invention, the

combination of

① a DNA primer having a nucleotide sequence represented by SEQ ID NO: 10 or SEQ ID NO: 16; with

② at least one DNA primer selected from the group consisting of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 2, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 4, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 8, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 9, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 15, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 17 and a DNA primer having a nucleotide sequence represented by SEQ ID NO: 19;

is, unlike conventional primers, capable of selectively amplifying a broad area covering from the second membrane-spanning domain to the sixth membrane-spanning domain or other domains of G protein coupled receptor proteins.

Among the DNA primers of the present invention, the

combination of

① a DNA primer having a nucleotide sequence represented by SEQ ID NO: 10 or SEQ ID NO: 16; with

② a DNA primer having a nucleotide sequence represented by SEQ ID NO: 11;

is, unlike conventional primers, capable of selectively amplifying a broad area covering from the second membrane-spanning domain to the seventh membrane-spanning domain or other domains of G protein coupled receptor proteins.

Among the DNA primers of the present invention, the

combination of

① at least one DNA primer selected from the group consisting of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 3, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 5, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 6, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 7, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 14 and a DNA

primer having a nucleotide sequence represented by SEQ ID NO: 18; with

② a DNA primer having a nucleotide sequence represented by SEQ ID NO: 11;

is, unlike conventional primers, capable of selectively amplifying a broad area covering from the third membrane-spanning domain to the seventh membrane-spanning domain or other domains of G protein coupled receptor proteins.

Therefore, the protein hydrophobicity plotting of G protein coupled receptor proteins and the homology at the amino acid level or the nucleic acid level between G protein coupled receptor proteins and other similar receptor proteins (said hydrophobicity plotting and homology both serve as standards for determining whether or not RNA or DNA (or fragment(s) thereof) obtained according to the present invention is (are) encoding part or all of the amino acid sequence of G protein coupled receptor protein] can now be more clearly calculated.

Among the DNA primers of the present invention, the combination of

① at least one DNA primer selected from the group consisting of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 3, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 5, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 6, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 7, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 14 and a DNA primer having a nucleotide sequence represented by SEQ ID NO: 18; with

② at least one DNA primer selected from the group consisting of a DNA primer having a nucleotide sequence represented by SEQ ID NO: 2, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 4, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 8, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 9, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 15, a DNA primer having a nucleotide sequence represented by SEQ ID NO: 17 and a DNA primer having a nucleotide sequence represented by SEQ ID

NO: 19;

is capable of amplifying the areas covering from the third membrane-spanning domain to the sixth membrane-spanning domain thereof at once like the conventional DNA primers and, moreover, it is capable of more selectively and efficiently amplifying DNA coding for G protein coupled receptor proteins though it has not been obtained through the conventional DNA primers.

Moreover, among the DNA primers of the present invention, the combination of

① at least one DNA primer selected from DNA primers having a nucleotide sequence of SEQ ID NO: 1 and DNA primers having a nucleotide sequence of SEQ ID NO: 12; with

② a DNA primer having a nucleotide sequence represented by SEQ ID NO: 13;

is capable of amplifying the areas covering from the first membrane-spanning domain to the third membrane-spanning domain thereof at once.

Then (a) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the first membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor protein, (b) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the first membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor protein

(c) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the third membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor protein, (d) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the third membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor protein, (e) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the second membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor protein,

(f) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the second membrane-spanning domain

to the seventh membrane-spanning domain of G protein coupled receptor protein, (g) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of from the first membrane-spanning domain to the third membrane-spanning domain of G protein coupled receptor protein or (h) the amplified DNA (or fragment(s) thereof) coding for the amino acid sequence of other domains of G protein coupled receptor protein may be used as a probe(s) to screen for full-length DNA which completely encodes G protein coupled receptor proteins from DNA libraries according to methods known per se by those of skill in the art or methods similar thereto.

The DNA libraries used in the present invention include any of genome DNA libraries, cDNA libraries and RNA libraries. The term "DNA library" or "DNA libraries" as used herein refers to a DNA library or DNA libraries including all of those libraries.

The present invention further provides screening methods for target DNA (or fragment(s) thereof) coding for G protein coupled receptor protein from the DNA library containing DNA (or fragment(s) thereof) coding for receptor proteins, which comprise employing the DNA of the present invention as a DNA primer for the PCR.

One preferred embodiment of the present invention is a method for cloning full-length DNA which completely encodes an amino acid sequence of G protein coupled receptor protein from DNA libraries which comprises the steps of

- (i) using the DNA of the present invention as a DNA primer for PCR;
- (ii) carrying out PCR in the presence of a mixture of said DNA primer with the DNA library to amplify and select (i.e. screen for) a DNA fragment coding for the amino acid sequence of from the first membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor protein, a DNA fragment coding for the amino acid sequence of from the first membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor protein, a DNA fragment coding for the amino acid sequence of from the third membrane-

spanning domain to the sixth membrane-spanning domain of G protein coupled receptor protein, a DNA fragment coding for the amino acid sequence of from the third membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor protein, a DNA fragment coding for the amino acid sequence of from the second membrane-spanning domain to the sixth membrane-spanning domain of G protein coupled receptor protein, a DNA fragment coding for the amino acid sequence of from the second membrane-spanning domain to the seventh membrane-spanning domain of G protein coupled receptor protein, a DNA fragment coding for the amino acid sequence of from the first membrane-spanning domain to the third membrane-spanning domain of G protein coupled receptor protein or a DNA fragment coding for other domains of G protein coupled receptor protein; and

- (iii) cloning said full-length DNA from the DNA library according to cloning methods known per se by those of skill in the art or methods similar thereto by using, as a probe, the DNA fragment obtained in the above step (ii).

Preferably, an embodiment of the present invention is a screening method of DNA coding for G protein coupled receptor proteins from DNA libraries, which comprises carrying out a polymerase chain reaction in the presence of

- ① the DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1, DNA primers having a nucleotide sequence represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7, DNA primers having a nucleotide sequence represented by SEQ ID NO: 10, DNA primers having a nucleotide sequence represented by SEQ ID NO: 14, DNA primers having a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18 and

③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 11, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19 to selectively amplify template DNA coding for G protein coupled receptor protein contained in the DNA library.

More preferably, embodiments of the present invention include:

- (1) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the first transmembrane domain to the sixth transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of
 - ① the DNA library,
 - ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12 and
 - ③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19

to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the first transmembrane domain to the sixth transmembrane domain of G protein coupled receptor protein or other domains thereof) contained in the DNA library;

(2) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the first transmembrane domain to the seventh transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① the DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12 and
- ③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 11

to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the first transmembrane domain to the seventh transmembrane domain of G protein coupled receptor protein or other domains thereof) contained in the DNA library;

(3) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the second transmembrane domain to the sixth transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① the DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by

SEQ ID NO: 10 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 16 and

- ③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19
- to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the second transmembrane domain to the sixth transmembrane domain of G protein coupled receptor protein or other domains thereof) contained in the DNA library;

- (4) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the second transmembrane domain to the seventh transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① the DNA library,
 - ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 10 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 16 and
 - ③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 11
- to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the second transmembrane domain to the seventh transmembrane domain of G protein coupled

receptor protein or other domains thereof) contained in the DNA library;

- (5) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the third transmembrane domain to the sixth transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① the DNA library,
 - ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7, DNA primers having a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18 and
 - ③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 2, DNA primers having a nucleotide sequence represented by SEQ ID NO: 4, DNA primers having a nucleotide sequence represented by SEQ ID NO: 8, DNA primers having a nucleotide sequence represented by SEQ ID NO: 9, DNA primers having a nucleotide sequence represented by SEQ ID NO: 15, DNA primers having a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 19
- to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the third transmembrane domain to the sixth transmembrane domain of G protein coupled receptor protein or other domains thereof) contained in the DNA library;
- (6) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like

(e.g. the regions spanning from the third transmembrane domain to the seventh transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of

- ① the DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 3, DNA primers having a nucleotide sequence represented by SEQ ID NO: 5, DNA primers having a nucleotide sequence represented by SEQ ID NO: 6, DNA primers having a nucleotide sequence represented by SEQ ID NO: 7, DNA primers having a nucleotide sequence represented by SEQ ID NO: 14 and having a nucleotide sequence represented by SEQ ID NO: 14 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 18 and
- ③ at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 11

to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the third transmembrane domain to the seventh transmembrane domain of G protein coupled receptor protein or other domains thereof) contained in the DNA library; and

- (7) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the first transmembrane domain to the third transmembrane domain of G protein coupled receptor protein or other domains thereof) from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① the DNA library,
- ② at least one DNA primer selected from the group consisting of DNA primers having a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers having a nucleotide sequence represented by SEQ ID NO: 12 and
- ③ at least one DNA primer selected from the group consisting

of DNA primers having a nucleotide sequence represented by SEQ ID NO: 13

to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein and the like (e.g. the regions spanning from the first transmembrane domain to the third transmembrane domain of G protein coupled receptor protein or other domains thereof) contained in the DNA library.

Particularly preferably, embodiments of the present invention include:

- (8) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① the DNA library,
- ② a DNA primer having a nucleotide sequence represented by SEQ ID NO: 1 and
- ③ a DNA primer having a nucleotide sequence represented by SEQ ID NO: 2

to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein contained in the DNA library;

- (9) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein from a DNA library, which comprises carrying out a polymerase chain reaction in the presence of a mixture of
- ① the DNA library,
- ② a DNA primer having a nucleotide sequence represented by SEQ ID NO: 3 and
- ③ a DNA primer having a nucleotide sequence represented by SEQ ID NO: 4

to selectively amplify the DNA coding for the amino acid sequence of G protein coupled receptor protein contained in the DNA library;

- (10) a screening method of DNA coding for the amino acid sequence of G protein coupled receptor protein from a DNA library, which comprises carrying out a polymerase chain

reaction in the presence of a mixture of

① the DNA library,

② a DNA primer having a nucleotide sequence represented by

SEQ ID NO: 6 and

③ a DNA primer having a nucleotide sequence represented by

SEQ ID NO: 8

to selectively amplify the DNA coding for the amino acid
sequence of G protein coupled receptor protein contained in
the DNA library; and

(11) a screening method of DNA coding for the amino acid
sequence of G protein coupled receptor protein from a DNA
library, which comprises carrying out a polymerase chain
reaction in the presence of a mixture of

① the DNA library,

② a DNA primer having a nucleotide sequence represented by

SEQ ID NO: 10 and

③ a DNA primer having a nucleotide sequence represented by

SEQ ID NO: 11

to selectively amplify the DNA coding for the amino acid
sequence of G protein coupled receptor protein contained in
the DNA library.

The cloned DNAs can be analyzed, usually by
restriction enzyme analysis and/or sequencing.

Target RNA or DNA (or fragment(s) thereof) coding
for G protein coupled receptor protein in the amplification
and the screening by the PCR techniques wherein the DNA of
the present invention is employed may include RNA, DNA or
fragments thereof coding for known (or prior art) G protein
coupled receptor proteins and RNA, DNA or fragments thereof
coding for unknown (novel) G protein coupled receptor
proteins.

These target RNA or DNA (or fragment(s) thereof) may
include novel nucleotide sequences and even known nucleotide
sequences.

Examples of such nucleotide sequences are RNA or DNA
(or fragment(s)) coding for a G protein coupled receptor
protein, said RNA or DNA (or fragment(s)) being derived from

all cells and tissues (e.g. pituitary gland, brain, pancreas,
lung, adrenal gland, etc.) of vertebrate animals (e.g. mice,
rats, cats, dogs, swines, cattle, horses, monkeys, human
beings, etc.), insects or other invertebrate animals (e.g.
dirosophila, silkworms, Barathra brassicae, etc.), plants
(e.g. rice plant, wheat, tomato, etc.) and cultured cell
lines derived therefrom, etc.

Specific examples of the nucleotide sequences are
RNA or DNA (or fragment(s)) coding for G protein coupled
receptor proteins such as receptor proteins to angiotensin,
bombesin, canavinoic, cholecystokinin, glutamine, serotonin,
melatonin, neuropeptide Y, opioid, purine, vasopressin,
oxytocin, VIP (vasoactive intestinal and related peptide),
somatostatin, dopamine, motilin, amylin, bradykinin, CGRP
(calcitonin gene related peptide), adrenomedullin, leukotriene,
pancreastatin, prostaglandin, thromboxane, adenosine,
adrenaline, α - and β -chemokine (IL-8, GRO α , GRO β , GRO γ ,
NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HCl4, MCP-3, I-309,
MIP1 α , MIP-1 β , RANTES, etc.), endothelin, enterogastatin,
histamine, neurotensin, TRH, pancreatic polypeptide, galanin,
family members thereof, etc.

In the PCR amplification using the DNA of the
present invention, the DNA (or DNA fragment) acting as a
template may include any DNA so far as it is derived from the
above-mentioned tissues and cells. More specifically, the
template DNA (or DNA fragment) includes any of genome DNA,
genome DNA libraries, cDNA derived from the tissues and cells
and cDNA libraries derived from the tissues and cells. cDNA
libraries derived from human tissues and cells are
particularly suitable. Vectors to be used in the DNA library
may include any of bacteriophages, plasmids, cosmids,
phagimids, etc. It is also possible to directly amplify the
template DNA (or DNA fragment) by reverse transcriptase
polymerase chain reaction (RT-PCR) techniques using mRNA
fractions prepared from the tissues and cells. The DNA which
is to be a template may be either DNA completely coding for
G protein coupled receptor proteins or DNA fragments (or

segments) thereof.

Preferably, the RNA or DNA (or fragment(s) thereof) obtained via the instant screening method for G protein coupled receptor protein coding DNA wherein said method uses the DNA according to the present invention is a G protein coupled receptor protein-encoding RNA or DNA (or fragment(s) thereof) contained in the used DNA library. More specifically, it is an RNA or DNA (or RNA fragment(s) or DNA fragment(s) (hereinafter, may be often abbreviated as just "DNA") coding for G protein coupled receptor proteins such as angiotensin receptor, bombesin receptor, canavaninoid receptor, cholecystokinin receptor, glutamine receptor, serotonin receptor, melatonin receptor, neuropeptide Y receptor, opioid receptor, purine receptor, vasopressin receptor, oxytocin receptor, VIP receptor (vasoactive intestinal and related peptide receptor), somatostatin receptor, dopamine receptor, motilin receptor, amylin receptor, bradykinin receptor, CGRP receptor (calcitonin gene related peptide receptor), adrenomedullin receptor, leukotriene receptor, pancreastatin receptor, prostaglandin receptor, thromboxane receptor, adenosine receptor, adrenaline receptor, α - and β -chemokine receptor (receptors to IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HC14, MCP-3, I-309, MIP1 α , MIP-1 β , RANTES, etc.), endothelin receptor, enterogastrin receptor, histamine receptor, neurotensin receptor, TRH receptor, pancreatic polypeptide receptor, galanin receptor, their family member receptors, etc.

When the DNA obtained by the screening method of the present invention is the DNA fragment which partially codes for a G protein coupled receptor protein, it is possible to isolate DNA completely encoding said G protein coupled receptor protein from a suitable DNA library according to cloning techniques known per se by using said DNA fragment as a probe.

Means for cloning the DNA completely encoding G protein coupled receptor proteins may include a PCR amplification employing a synthetic DNA primer having the partial nucleotide sequence of the DNA fragment partially

coding for the G protein coupled receptor protein and a selection of the target DNA via a hybridization with DNA or synthetic DNA having part or all of the region of said DNA fragments. The hybridization may be conducted, for example, by the methods described in Molecular Cloning, 2nd ed.; J. Sambrook et al., Cold Spring Harbor Lab. Press, 1989. When the commercially available library is used, it may be conducted according to the manners described in the protocol attached thereto.

The DNA completely encoding G protein coupled receptor protein (full-length G protein coupled receptor protein DNA) may be used, depending upon its object, either as it is or after digesting with a restriction enzyme or after ligating with a linker if desired. Said DNA may have ATG at the 5'-terminal as the translation initiation codon and TAA, TGA or TAG at the 3' terminal as the translation termination codon. These translation initiation codons and translation termination codons may be added using a suitable synthetic DNA adaptor. In addition, it is possible to determine said receptor protein-expressing tissues/cells by northern blottings using said DNA as a probe. It is also possible to express target receptor proteins by introducing DNA having the entire coding region of the receptor protein into animal cells after binding with a suitable promoter.

The G protein coupled receptor protein according to the present invention is a G protein coupled receptor protein encoded by the G protein coupled receptor protein-encoding DNA obtained by the screening method of the present invention. More specifically, the G protein coupled receptor protein according to the present invention includes G protein coupled receptor proteins such as angiotensin receptor protein, bombesin receptor protein, canavaninoid receptor protein, cholecystokinin receptor protein, glutamine receptor protein, serotonin receptor protein, melatonin receptor protein, neuropeptide Y receptor protein, opioid receptor protein, purine receptor protein, vasopressin receptor protein, oxytocin receptor protein, VIP receptor protein (vasoactive

intestinal and related peptide receptor protein), somatostatin receptor protein, dopamine receptor protein, motilin receptor protein, amylin receptor protein, bradykinin receptor protein, CGRP receptor protein (calcitonin gene related peptide receptor protein), adrenomedullin receptor protein, leukotriene receptor protein, pancreastatin receptor protein, prostaglandin receptor protein, thromboxane receptor protein, adenosine receptor protein, adrenaline receptor protein, α - and β -chemokine receptor protein (receptor protein responsive to IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PFA, IP10, GCP-2, MCP-1, HCL4, MCP-3, I-309, MIP1 α , MIP-1 β , RANTES, etc.), endothelin receptor protein, enterogastrin receptor protein, histamine receptor protein, neurotensin receptor protein, TRH receptor protein, pancreatic polypeptide receptor protein, galanin receptor protein, family members thereof, etc.

According to the present invention, novel G protein coupled receptors proteins, peptide segments or fragments derived from the G protein coupled receptor protein, modified derivatives or analogues thereof, and salts thereof may be recognized, cloned, produced, isolated or characterized.

These G protein coupled receptor proteins are those derived from all cells and tissues (e.g. pituitary gland, pancreas, brain, kidney, liver, gonad, thyroid gland, cholecyst, bone marrow, adrenal, skin, muscle, lung, digestive duct, blood vessel, heart, etc.) of warm-blooded animals (e.g. guinea pig, rat, mouse, swine, sheep, cattle, monkey, human beings, rabbit, cat, dog, horse, etc.), and any of proteins as long as they comprise an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 24, an amino acid sequence represented by SEQ ID NO: 25, an amino acid sequence represented by SEQ ID NO: 26, an amino acid sequence represented by SEQ ID NO: 27, an amino acid sequence represented by SEQ ID NO: 28, an amino acid sequence represented by SEQ ID NO: 34, an amino acid sequence represented by SEQ ID NO: 35, an amino acid sequence represented by SEQ ID NO: 38, an amino acid sequence represented by SEQ ID NO: 39, an amino acid sequence

represented by SEQ ID NO: 56, and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 38, SEQ ID NO: 39, and/or SEQ ID NO: 56.

In one embodiment of the present invention, G protein coupled receptor proteins are those derived from all cells and tissues (e.g. pituitary gland, pancreas, brain, kidney, liver, gonad, thyroid gland, cholecyst, bone marrow, adrenal, skin, muscle, lung, digestive duct, blood vessel, heart, etc.) of warm-blooded animals (e.g. guinea pig, rat, mouse, swine, sheep, cattle, monkey, human beings, cat, dog, horse, etc.), and any of proteins as long as they comprise an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 24, an amino acid sequence represented by SEQ ID NO: 25, an amino acid sequence represented by SEQ ID NO: 26, an amino acid sequence represented by SEQ ID NO: 27, an amino acid sequence represented by SEQ ID NO: 28, and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, or SEQ ID NO: 28. These G protein coupled receptor proteins may include proteins having an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 24, an amino acid sequence represented by SEQ ID NO: 25, an amino acid sequence represented by SEQ ID NO: 26, an amino acid sequence represented by SEQ ID NO: 27 and an amino acid sequence represented by SEQ ID NO: 28, proteins wherein the amino acid sequence thereof is about 90% to 99.9% homologous to an amino acid sequence represented by SEQ ID NO: 24, an amino acid sequence represented by SEQ ID NO: 25, an amino acid sequence represented by SEQ ID NO: 26, an amino acid sequence represented by SEQ ID NO: 27 or an amino acid sequence represented by SEQ ID NO: 28 and the activity thereof is substantially equivalent to the protein having an amino acid sequence represented by SEQ ID NO: 24, an amino acid sequence represented by SEQ ID NO: 25, an amino acid sequence

represented by SEQ ID NO: 26, an amino acid sequence represented by SEQ ID NO: 27 or an amino acid sequence represented by SEQ ID NO: 28 and the like. The substantially equivalent activity may include ligand binding activity, signal information transmitting, etc. The term "substantially equivalent" or "substantial equivalent" means that the nature of the ligand binding activity and the like is equivalent. Therefore, it is allowable that even differences among grades such as ligand binding affinity grades and ligand binding activity grades and quantitative factors such as molecular weights of receptor proteins are present.

In another embodiment of the present invention, G protein coupled receptor proteins include human pituitary gland-derived G protein coupled receptor proteins comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 24, and/or an amino acid sequence represented by SEQ ID NO: 25, mouse pancreas-derived G protein coupled receptor proteins comprising an amino acid sequence represented by SEQ ID NO: 27, mouse pancreas-derived G protein coupled receptor proteins comprising an amino acid sequence represented by SEQ ID NO: 28, etc.

Examples of the human pituitary gland-derived G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 24, and an amino acid sequence represented by SEQ ID NO: 25, are human pituitary gland-derived G protein coupled receptor proteins comprising an amino acid sequence represented by SEQ ID NO: 24, etc. These G protein coupled receptor proteins may include proteins wherein one or more amino acid

residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27 or SEQ ID NO: 28, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27 or SEQ ID NO: 28, proteins

wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27 or SEQ ID NO: 28, are substituted with one or more amino acid residues, etc.

In yet another embodiment of the present invention, G protein coupled receptor proteins include those derived from all cells and tissues (e.g. amygdaloid nucleus, pituitary gland, pancreas, brain, kidney, liver, gonad, thyroid gland, cholecyst, bone marrow, lung, digestive duct, blood vessel, heart, thymus, spleen, leukocyte, etc.) of warm-blooded animals (e.g. guinea pig, rat, mouse, pig, sheep, cattle, monkey, human beings, etc.), and any of proteins as long as they comprise an amino acid sequence selected from the group

consisting of an amino acid sequence represented by SEQ ID NO: 34 and/or an amino acid sequence represented by SEQ ID NO: 35. These G protein coupled receptor proteins may include proteins having an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 34 or/and an amino acid sequence represented by SEQ ID NO: 35, proteins wherein the amino acid sequence thereof is about 90% to 99.9% homologous to an amino acid sequence represented by SEQ ID NO: 34 or/and an amino acid sequence

represented by SEQ ID NO: 35 and the activity thereof is substantially equivalent to the protein having an amino acid sequence represented by SEQ ID NO: 34 and/or an amino acid sequence represented by SEQ ID NO: 35, and the like. The

substantially equivalent activity may include ligand binding activity, signal information transmitting, etc. The term "substantially equivalent" or "substantial equivalent" means that the nature of the ligand binding activity and the like is equivalent. Therefore, it is allowable that even differences among grades such as ligand binding affinity grades and ligand binding activity grades and quantitative factors such as molecular weights of receptor proteins are present.

Examples of the G protein coupled receptor protein are human amygdaloid nucleus-derived G protein coupled receptor proteins

having an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 34 and/or an amino acid sequence represented by SEQ ID NO: 35, etc. These G protein coupled receptor proteins may include proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 34 or SEQ ID NO: 35, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 34 or SEQ ID NO: 35, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 34 or SEQ ID NO: 35, are substituted with one or more amino acid residues, etc.

In still another embodiment of the present invention, these G protein coupled receptor proteins are those derived from all cells and tissues (e.g. amygdaloid nucleus, pituitary gland, pancreas, brain, kidney, liver, gonad, thyroid body, cholecyst, bone marrow, lung, digestive duct, blood vessel, heart, thymus, leukocyte, etc.) of warm-blooded animals (e.g. guinea pig, rat, mouse, swine, sheep, cattle, monkey, human beings, etc.), and any of proteins as long as they comprise an amino acid sequence represented by SEQ ID NO: 38, or substantial equivalents to the amino acid sequence represented by SEQ ID NO: 38, preferably an amino acid sequence represented by SEQ ID NO: 39, or substantial equivalents to the amino acid sequence represented by SEQ ID NO: 39.

These G protein coupled receptor proteins may include proteins having an amino acid sequence represented by SEQ ID NO: 38, proteins wherein the amino acid sequence thereof is about 90% to 99.9% homologous to an amino acid sequence represented by SEQ ID NO: 38 and the activity thereof is substantially equivalent to the protein having an amino acid sequence represented by SEQ ID NO: 38 and the like.

These G protein coupled receptor proteins are preferably

proteins having an amino acid sequence represented by SEQ ID NO: 39, proteins wherein the amino acid sequence thereof is about 90% to 99.9% homologous to an amino acid sequence represented by SEQ ID NO: 39 and the activity thereof is substantially equivalent to the protein having an amino acid sequence represented by SEQ ID NO: 39, etc. The substantially equivalent activity may include ligand binding activity, signal information transmitting, etc. The term "substantially equivalent" or "substantial equivalent" means that the nature of the ligand binding activity and the like is equivalent. Therefore, it is allowable that even differences among grades such as ligand binding affinity grades and ligand binding activity grades and quantitative factors such as molecular sizes or weights of receptor proteins are present.

It is suggested by data that the mouse pancreatic β -cell strain, MIN6-derived receptor protein of the present invention (e.g., SEQ ID NO: 38 and SEQ ID NO: 39, or proteins encoded by pMH2-17) is a novel purinoceptor subtype which is clearly distinct from prior art purinoceptors.

In another more specific embodiment of the present invention, G protein coupled receptor proteins include mouse pancreatic β -cell line, MIN6, derived G protein coupled receptor proteins comprising an amino acid sequence represented by SEQ ID NO: 38, mouse pancreatic β -cell line, MIN6, derived G protein coupled receptor proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 38, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 38, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues) are substituted with other amino acid residues in the amino acid sequence of SEQ ID NO: 38, etc. Further preferably these G protein coupled receptor proteins include mouse pancreatic β -cell line, MIN6, derived G

protein coupled receptor proteins comprising an amino acid sequence represented by SEQ ID NO: 39, mouse pancreatic β -cell line, MIN6, derived G protein coupled receptor proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 39, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 39, proteins wherein one or more amino acid residues, more preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) in the amino acid sequence of SEQ ID NO: 39 are substituted with other amino acid residues, etc.

In still another embodiment of the present invention, these G protein coupled receptor proteins are those derived from all cells and tissues (e.g. placenta, gonad, amygdaloid nucleus, pituitary body, pancreas, brain, kidney, liver, thyroid gland, cholecyst, bone marrow, lung, digestive duct, blood vessel, heart, thymus, leukocyte, etc.) of human beings, and any of proteins as long as they comprise an amino acid sequence represented by SEQ ID NO: 56, or substantial

equivalents to the amino acid sequence represented by SEQ ID NO: 56. These G protein coupled receptor proteins may include proteins having an amino acid sequence represented by SEQ ID NO: 56, proteins wherein the amino acid sequence thereof is about 90% to 99.9% homologous to an amino acid sequence represented by SEQ ID NO: 56 and the activity thereof is substantially equivalent to the protein having an amino acid sequence represented by SEQ ID NO: 56 and the like.

The substantially equivalent activity may include ligand binding activity, signal information transmitting, etc. The term "substantially equivalent" or "substantial equivalent" means that the nature of the ligand binding activity and the like is equivalent. Therefore, it is allowable that even differences among grades such as ligand binding affinity grades and ligand binding activity grades and quantitative factors

such as molecular sizes or weights of receptor proteins are present.

In another more specific embodiment of the present invention, G protein coupled receptor proteins include G protein coupled receptor proteins comprising an amino acid sequence represented by SEQ ID NO: 56, G protein coupled receptor proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are deleted from the amino acid sequence of SEQ ID NO: 56, proteins wherein one or more amino acid residues (preferably from 2 to 30 amino acid residues, more preferably from 2 to 10 amino acid residues) are added to the amino acid sequence of SEQ ID NO: 56, proteins wherein one or more amino acid residues, etc.

A portion of the amino acid sequence may be modified (e.g. addition, deletion, substitution with other amino acids, etc.) in the G protein coupled receptor proteins of the present invention.

Furthermore, the G protein coupled receptor proteins of the present invention includes those wherein N-terminal Met is protected with a protecting group (e.g., C₁₋₆ acyl group such as formyl, acetyl, etc.), those wherein the N-terminal side of Glu is cleaved in vivo to make said Glu pyroglutaminated, those wherein the intramolecular side chain of amino acids is protected with a suitable protecting group (e.g., C₁₋₆ acyl group such as formyl, acetyl, etc.), conjugated proteins such as so-called "glycoproteins" wherein saccharide chains are bonded, etc.

The salt of said G protein coupled receptor protein of the present invention includes preferably physiologically acceptable acid addition salts. Examples of such salts are salts thereof with inorganic acids (e.g. hydrochloric acid, phosphoric acid, hydrobromic acid, sulfuric acid, etc.), salts

thereof with organic acids (e.g. acetic acid, formic acid, propionic acid, fumaric acid, maleic acid, succinic acid, tartaric acid, citric acid, malic acid, oxalic acid, benzoic acid, methanesulfonic acid, benzenesulfonic acid, etc.).

The G protein coupled receptor protein or its salt of the present invention may be manufactured from the tissues or cells of warm-blooded animals by purifying methods which are known per se by those skilled in the art or methods similar thereto or may be manufactured by culturing the transformant (or transfectant) (as described herein below) containing G protein coupled receptor protein encoding DNA. The protein or its salt of the present invention may be manufactured by the peptide synthesis as described herein below.

The G protein coupled receptor protein fragment (the partial peptide of said G protein coupled receptor protein) may include, for example, the site which is exposed outside cell membranes, among the G protein coupled receptor protein molecule. Examples of the fragment are peptides containing a region which is analyzed as an extracellular area (hydrophilic region or site) in a hydrophobic plotting analysis on the G protein coupled receptor protein represented by any of Figures 24, 25, 28, 31, 32, 36, 38, 41, 44, 47, 50, 53, 57, 58, 59, 64, 70, 74, and 78.

A peptide which partly contains a hydrophobic region or site may be used as well. Further, a peptide which separately contains each domain may be used too although the partial peptide (peptide fragment) which contains plural domains at the same time will be used as well.

The salt of said G protein coupled receptor protein fragment (partial peptide thereof) includes preferably physiologically acceptable acid addition salts. Examples of such salts are salts thereof with inorganic acids (e.g. hydrochloric acid, phosphoric acid, hydrobromic acid, sulfuric acid, etc.), salts thereof with organic acids (e.g. acetic acid, formic acid, propionic acid, fumaric acid, maleic acid, succinic acid, tartaric acid, citric acid, malic acid, oxalic acid, benzoic acid, methanesulfonic acid, benzenesulfonic

acid, etc.), etc.

The G protein coupled receptor protein fragment (the partial peptide of the G protein coupled receptor protein) may be manufactured by synthesizing methods for peptides which are known per se by those skilled in the art or methods similar thereto or by cleaving (digesting) G protein coupled receptor proteins by a suitable peptidase. Methods of synthesizing peptide may be any of a solid phase synthesis and a liquid phase synthesis. Thus, a partial peptide (peptide fragment) or amino acids which can construct the protein of the present invention is condensed with the residual part thereof and, when the product has a protective group, said protective group is detached whereupon a desired peptide can be manufactured. Examples of the known methods for condensation and for detachment of protective groups include the following

- ① to ⑤:
- ① M. Bodanszky and M. A. Ondetti: *Peptide Synthesis*, Interscience Publishers, New York (1966).
- ② Schroeder and Luebke: *The Peptide*, Academic Press, New York, 1965.
- ③ Nobuo Izumiya et al.: *Fundamentals and Experiments of the Peptide Synthesis*, Maruzen KK, Japan (1975).
- ④ Haruaki Yajima and Shunpei Sakakibara: "Seikagaku Jikken Koza 1" (Experiments of Biochemistry, Part 1), "Tanpakusitu No Kagaku IV" (Chemistry of Protein, IV), p.205 (1977), Japan.
- ⑤ Haruaki Yajima (ed): *Development of Pharmaceuticals* (Second Series), Vol. 14, *Peptide Synthesis*, Hirokawa Shoten, Japan.

After the reaction, conventional purifying techniques such as salting-out, extraction with solvents, distillation, column chromatography, liquid chromatography, electrophoresis, recrystallization, etc. are optionally combined so that the protein of the present invention can be purified and isolated. When the protein obtained as such is a free compound, it may be

converted to a suitable salt by known methods while, when it is obtained as a salt, the salt may be converted to a free compound or other salt compounds by known methods.

Furthermore, the product may be manufactured by culturing the transformant (transfectant) containing the DNA coding for said partial peptide.

The G protein coupled receptor protein-encoding DNA obtained by the above-mentioned screening method using the DNA of the present invention and the G protein coupled receptor protein encoded by said DNA or the peptide fragment (partial peptide thereof) encoded by said DNA may, for example, be used for the determination of a ligand to said G protein coupled receptor protein or for the screening of a compound which inhibits the binding of said protein coupled receptor protein with a ligand.

In that case, an expression system for the G protein coupled receptor protein-encoding DNA is at first constructed. Hosts for said DNA may be any of animal cells, insect cells, yeasts, *Bacillus subtilis*, *Escherichia coli*, etc. Promoters used therefor may be anyone so far as it is suitable as a promoter for the host used for gene expression. Incidentally, the utilization of enhancers for expression is effective as well.

Then the expressing cells per se which constructed to express the G protein coupled receptor protein or the cell membrane fractions prepared therefrom by methods known per se by those skilled in the art or methods similar thereto may be subjected to a variety of receptor binding experiments.

Ligands used therefor may include any of compounds labeled by a commercially available radioisotope, etc., culture supernatants and tissue extracts which are directly labeled by a chloramine T method or by a lactoperoxidase method.

Separation of bonded or free ligands may be carried out by a direct washing when cells adhered to substrates are used, while, in the case of floating cells or cell membrane fractions thereof, it may be carried out by means of centrifugal separation or filtration. Nonspecific binding with container,

etc. may be estimated by addition of unlabeled ligands which are about 100 times as much concentrated relatively to the poured labeled ligand.

The ligand which is obtained by such a receptor binding experiment may be subjected to a discrimination of agonist versus antagonist.

To be more specific, a natural substance or compound which is presumed to be a ligand with the G protein coupled receptor protein-expressing cell is cultured and, after that, the culture supernatant liquid is collected or the cell is extracted. A change in the components contained therein is measured by, for example, a commercially available measuring kit (e.g. kits for cAMP, diacylglycerol, cGMP, protein kinase A, etc.). Alternatively, it is possible to measure physiological responses such as liberation of Fura-2, [³H]arachidonic acid and [³H]inositol phosphate metabolites by methods known per se by those skilled in the art or methods similar thereto. The compound or natural substance which is obtained by such a screening is an agonist for said G protein coupled receptor protein and is presumed to act on the tissues and cells in which said receptor is distributed. Accordingly, it is possible to check the pharmaceutical response (pharmaceutical effect) more efficiently by referring to the distribution disclosed (clarified) by a northern blotting or the like. Moreover, a development of compounds having a novel pharmaceutical response (pharmaceutical effect) in, for example, central nervous tissues, circulatory system, kidney, pancreas, etc. is expected. An efficient development of pharmaceuticals can be proceeded by amplifying G protein coupled receptor protein-encoding DNA selectively from tissues.

The G protein coupled receptor protein-encoding DNA of the present invention may be any coding DNA as long as it contains a nucleotide sequence coding for a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 24 and/or which has an activity substantially

equivalent to the amino acid sequence having SEQ ID NO: 24, a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 25 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 25, a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 26 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 26, a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 27 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 27, or a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 28 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 28.

Still the G protein coupled receptor protein-encoding DNA of the present invention may be any coding DNA as long as it contains a nucleotide sequence coding for a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 34 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 34, or a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 35 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 35.

Yet the G protein coupled receptor protein-encoding DNA of the present invention may be any coding DNA as long as it contains a nucleotide sequence coding for a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 38 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 38, or

preferably a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 39 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 39.

Yet the G protein coupled receptor protein-encoding DNA of the present invention may be any coding DNA as long as it contains a nucleotide sequence coding for a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 56 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 56, or preferably a G protein coupled receptor protein which contains an amino acid sequence substantially equivalent to the amino acid sequence having SEQ ID NO: 56 and/or which has an activity substantially equivalent to the amino acid sequence having SEQ ID NO: 56.

The DNA of the present invention may be any one of a human genome DNA, a human genome DNA library, a human tissue and cell-derived cDNA, a human tissue and cell-derived cDNA library and a synthetic DNA. The vector used for the library may include bacteriophage, plasmid, cosmid, phagemid, etc. The DNA can be further amplified directly by the reverse transcriptase polymerase chain reaction (hereinafter briefly referred to as "RT-PCR") using mRNA fractions prepared from tissues and cells.

In an embodiment, the DNA coding for the human pituitary gland-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 24 includes DNA having a nucleotide sequence represented by SEQ ID NO: 29, etc. The DNA coding for the human pituitary gland-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 29, etc. The DNA coding sequence represented by SEQ ID NO: 30, etc. The DNA coding for the human pituitary gland-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 26 includes DNA having a nucleotide sequence represented by

SEQ ID NO: 31, etc. The DNA coding for the mouse pancreas-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 27 includes DNA having a nucleotide sequence represented by SEQ ID NO: 32, etc. The DNA coding for the mouse pancreas-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 28 includes DNA having a nucleotide sequence represented by SEQ ID NO: 33, etc.

In another embodiment, the DNA coding for the human amygdaloid nucleus-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 34 includes DNA having a nucleotide sequence represented by SEQ ID NO: 36, etc. The DNA coding for the human amygdaloid nucleus-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 35 includes DNA having a nucleotide sequence represented by SEQ ID NO: 37, etc. The DNA coding for the human amygdaloid nucleus-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 34 or the amino acid sequence of SEQ ID NO: 35 includes DNA having a nucleotide sequence represented by SEQ ID NO: 36, etc. Still in another embodiment, the DNA coding for the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 38 includes DNA having a nucleotide sequence represented by SEQ ID NO: 40, etc. The DNA coding for the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 39 includes DNA having a nucleotide sequence represented by SEQ ID NO: 41, etc. Yet in another embodiment, the DNA coding for the human-derived G protein coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 56 includes DNA having a nucleotide sequence represented by SEQ ID NO: 57, etc.

The DNA completely coding for the G protein coupled receptor protein of the present invention can be cloned by (1) carrying out the PCR amplification using a synthetic DNA primer having a partial nucleotide sequence (nucleotide

fragment) of the G protein coupled receptor protein; or (2) effecting the selection of a DNA constructed in a suitable vector, based on the hybridization with a labeled DNA fragment having part or all of the region encoding a human G protein coupled receptor protein or a labeled synthetic DNA having part or all of the coding region thereof.

The hybridization is carried out according to methods as disclosed in, for example, Molecular Cloning, 2nd Ed., J. Sambrook et al., Cold Spring Harbor Lab. Press, 1989. When a DNA library commercially available in the market is used, the hybridization is carried out according to protocols manuals attached thereto.

The cloned G protein coupled receptor protein-encoding DNA of the present invention can be used as it is, or can be used, as desired, after modifications including digestion with a restriction enzyme or addition of a linker or adapter, etc. depending upon objects. The DNA may have an initiation codon, ATG, on the 5' terminal side and a termination codon, TAA, TGA or TAG, on the 3' terminal side. These initiation and termination codons can be ligated by using a suitable synthetic DNA adapter.

An expression vector for G protein coupled receptor proteins can be produced by, for example, (a) cutting out a target DNA fragment from the G protein coupled receptor protein-encoding DNA of the present invention and (b) ligating the target DNA fragment with the downstream site of a promoter in a suitable expression vector.

The vector may include plasmids derived from *Escherichia coli* (e.g., pBR322, pBR325, pUC12, pUC13, etc.), plasmids derived from *Bacillus subtilis* (e.g., pUB110, pTP5, pC194, etc.), plasmids derived from yeasts (e.g., pSH19, pSH15, etc.), bacteriophages such as λ -phage, and animal virus such as retrovirus, vaccinia virus and baculovirus.

According to the present invention, any promoter can be used as long as it is compatible with a host which is used for expressing a gene. When the host for the transformation is *E. coli*, the promoters are preferably trp promoters, lac

promoters, recA promoters, λ PL promoters, lpp promoters, etc. When the host for the transformation is the Bacillus, the promoters are preferably SP01 promoters, SP02 promoters, penP promoters, etc. When the host is an yeast, the promoters are preferably PH05 promoters, PGK promoters, GAP promoters, ADH promoters, etc. When the host is an animal cell, the promoters include SV40-derived promoters, retrovirus promoters, metallothionein promoters, heat shock promoters, cytomegalovirus promoters, SRA promoters, etc. An enhancer can be effectively utilized for the expression.

As required, furthermore, a host-compatible signal sequence is added to the N-terminal side of the G protein coupled receptor protein. When the host is E. coli, the utilizable signal sequences may include alkaline phosphatase signal sequences, ompA signal sequences, etc. When the host is the Bacillus, they may include α -amylase signal sequences, subtilisin signal sequences, etc. When the host is an yeast, they may include mating factor α signal sequences, invertase signal sequences, etc. When the host is an animal cell, they may include insulin signal sequences, α -interferon signal sequences, antibody molecule signal sequences, etc.

A transformant or transfectant is produced by using the vector thus constructed, which carries the G protein coupled receptor protein-encoding DNA of the present invention.

The host may be, for example, Escherichia microorganisms, Bacillus microorganisms, yeasts, insect cells, animal cells, etc. Examples of the Escherichia and Bacillus microorganisms include Escherichia coli K12-DH1 [Proc. Natl. Acad. Sci. USA, Vol. 60, 160 (1968)], JM103 [Nucleic Acids Research, Vol. 9, 309 (1981)], JA221 [Journal of Molecular Biology, Vol. 120, 517 (1978)], HB101 [Journal of Molecular Biology, Vol. 41, 459 (1969)], C600 [Genetics, Vol. 39, 440 (1954)], etc.

Examples of the Bacillus microorganism are, for example, Bacillus subtilis Mill4 [Gene, Vol. 24, 255 (1983)], 207-21 [Journal of Biochemistry, Vol. 95, 87 (1984)], etc. The yeast may be, for example, Saccharomyces cerevisiae AH22, AH22R, NA87-11A, DKD-5D, 20B-12, etc. The insect may include

a silkworm (Bombyx mori larva), [Maeda et al, Nature, Vol. 315, 592 (1985)] etc. The host animal cell may be, for example, monkey-derived cell line, COS-7, Vero, Chinese hamster ovary cell line (CHO cell), DHFR gene-deficient Chinese hamster cell line (dhfr⁻CHO cell), mouse L cell, murine myeloma cell, human FL cell, etc.

Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. Transformation of Escherichia microorganisms can be carried out in accordance with methods as disclosed in, for example, Proc. Natl. Acad. Sci. USA, Vol. 69, 2110 (1972), Gene, Vol. 17, 107 (1982), etc. Transformation of Bacillus microorganisms can be carried out in accordance with methods as disclosed in, for example, Molecular & General Genetics, Vol. 168, 111 (1979), etc. Transformation of the yeast can be carried out in accordance with methods as disclosed in, for example, Proc. Natl. Acad. Sci. USA, Vol. 75, 1929 (1978), etc. The insect cells can be transformed in accordance with methods as disclosed in, for example, Bio/Technology, 6, 47-55, 1988. The animal cells can be transformed by methods as disclosed in, for example, Virology, Vol. 52, 456, 1973, etc. The transformants or transfectants which are transformed with expression vectors containing a G protein coupled receptor protein-encoding DNA are produced according to the aforementioned techniques.

Cultivation of the transformant (transfectant) in which the host is Escherichia or Bacillus microorganism can be carried out suitably in a liquid culture medium. The culture medium may contain carbon sources, nitrogen sources, minerals, etc. necessary for growing the transformant. The carbon source may include glucose, dextrin, soluble starch, sucrose, etc. The nitrogen source may include organic or inorganic substances such as ammonium salts, nitrates, corn steep liquor, peptone, casein, meat extracts, bean-cakes, potato extracts, etc. Examples of the minerals may include calcium chloride, sodium dihydrogen phosphate, magnesium chloride, etc. It is further allowable to add yeasts, vitamins, growth-

promoting factors, etc. It is desired that the culture medium is pH from about 5 to about 8.

The Escherichia microorganism culture medium is preferably an M9 medium containing, for example, glucose and casamino acid (Miller, Journal of Experiments in Molecular Genetics), 431-433, Cold Spring Harbor Laboratory, New York, 1972. Depending on necessity, the medium may be supplemented with drugs such as 3 β -indolyl acrylic acid in order to improve efficiency of the promoter. In the case of the Escherichia host, the cultivation is carried out usually at about 15 to 43 °C for about 3 to 24 hours. As required, aeration and stirring may be applied. In the case of the Bacillus host, the cultivation is carried out usually at about 30 to 40 °C for about 6 to 24 hours. As required, aeration and stirring may be also applied. In the case of the transformant in which the host is a yeast, the culture medium used may include, for example, a Burkholder minimum medium [Bostian, K.L. et al., Proc. Natl. Acad. Sci. USA, Vol. 77, 4505 (1980)], an SD medium containing 0.5% casamino acid [Bitter, G.A. et al., Proc. Natl. Acad. Sci. USA, Vol. 81, 5330 (1984)], etc.

It is preferable that pH of the culture medium is adjusted to be from about 5 to about 8. The cultivation is carried out usually at about 20 to 35 °C for about 24 to 72 hours. As required, aeration and stirring may be applied.

In the case of the transformant in which the host is an insect, the culture medium used may include those obtained by suitably adding additives such as passivated (or immobilized) 10 \times bovine serum and the like to the Grace's insect medium (Grace, T.C.C., Nature, 195, 788 (1962)). It is preferable that pH of the culture medium is adjusted to be about 6.2 to 6.4. The cultivation is usually carried out at about 27 °C for about 3 to 5 days. As desired, aeration and stirring may be applied. In the case of the transformant in which the host is an animal cell, the culture medium used may include MEM medium [Science, Vol. 122, 501 (1952)], DMEM medium [Virology, Vol. 8, 396 (1959)], RPMI 1640 medium [Journal of the American Medical Association, Vol. 199, 519 (1967)], 199 medium [Proceedings of

the Society of the Biological Medicine, Vol. 73, 1 (1950)], etc. which are containing, for example, about 5 to 20% of fetal calf serum. It is preferable that the pH is from about 6 to about 8. The cultivation is usually carried out at about 30 to 40 °C for about 15 to 60 hours. As required, aeration and stirring may be applied.

Separation and purification of the G protein coupled receptor protein from the above-mentioned cultures can be carried out according to methods described herein below.

To extract G protein coupled receptor proteins from the cultured microorganisms or cells, the microorganisms or cells are collected by known methods after the cultivation, suspended in a suitable buffer solution, disrupted by ultrasonic waves, lysozyme and/or freezing and thawing, etc. and, then, a crude extract of the G protein coupled receptor protein is obtained by centrifugation or filtration. Other conventional extracting or isolating methods can be applied. The buffer solution may contain a protein-denaturing agent such as urea or guanidine hydrochloride or a surfactant such as Triton X-100 (registered trademark, hereinafter often referred to as "TM").

In case where G protein coupled receptor proteins are secreted into culture media, supernatant liquids are separated from the microorganisms or cells after the cultivation is finished and the resulting supernatant liquid collected by widely known methods. The culture supernatant liquid and extract containing G protein coupled receptor proteins can be purified by suitable combinations of widely known methods for separation, isolation and purification. The widely known methods of separation, isolation and purification may include methods which utilizes solubility, such as salting out or sedimentation with solvents methods which utilizes chiefly a difference in the molecular size or weight, such as dialysis, ultrafiltration, gel filtration and SDS-polyacrylamide gel electrophoresis, methods utilizing a difference in the electric charge, such as ion-exchange chromatography, methods utilizing specific

affinity such as affinity chromatography, methods utilizing a difference in the hydrophobic property, such as inverse-phase high-performance liquid chromatography, and methods utilizing a difference in the isoelectric point such as isoelectric electrophoresis, etc.

In case where the G protein coupled receptor protein thus obtained is in a free form, the free protein can be converted into a salt thereof by known methods or method analogous thereto. In case where the G protein coupled receptor protein thus obtained is in a salt form vice versa, the protein salt can be converted into a free form or into any other salt thereof by known methods or method analogous thereto.

The G protein coupled receptor protein produced by the transformant can be arbitrarily modified or a polypeptide can be partly removed therefrom, by the action of a suitable protein-modifying enzyme before or after the purification. The protein-modifying enzyme may include trypsin, chymotrypsin, arginyl endopeptidase, protein kinase, glycosidase, etc. The activity of the G protein coupled receptor protein thus formed can be measured by experimenting the coupling (or binding) with a ligand or by enzyme immunoassays (enzyme linked immunoassays) using specific antibodies.

The G protein coupled receptor protein-encoding DNA and the G protein coupled receptor protein of the present invention can be used for:

- ① methods of determining ligands for the G protein coupled receptor protein of the present invention,
- ② obtaining an antibody and an antiserum,
- ③ constructing a system for expressing a recombinant receptor protein,
- ④ developing a receptor-binding assay system using the above developing system and screening pharmaceutical candidate compounds,
- ⑤ designing drugs based upon the comparison with ligands and receptors which have a similar or analogous structure,

⑥ preparing a probe in the analysis of genes and preparing a PCR primer, and

⑦ gene manipulating therapy.

In particular, it is allowable to screen a G protein coupled receptor agonist or antagonist specific to a warm-blooded animal such as human being by a receptor-binding assay system which uses a system for expressing a recombinant G protein coupled receptor protein of the present invention. The agonist or antagonist thus screened or characterized permits various applications including prevention and/or therapy of a variety of diseases.

Concretely described below are uses of G protein coupled receptor proteins, partial peptide thereof (peptide fragment thereof), G protein coupled receptor protein-encoding DNAs and antibodies against the G protein coupled receptor protein according to the present invention.

As hereunder, more detailed description will be made on the usefulness of the G protein coupled receptor protein-encoding DNA obtained by the screening method for G protein coupled receptor protein-encoding DNAs according to the present invention, the G protein coupled receptor proteins encoded by said DNA, peptide fragments or segments thereof (including partial peptides thereof) or salts thereof (hereinafter, those including their salts, will be referred to as the "G protein coupled receptor protein or a peptide fragment thereof"), cells or cell membrane fractions thereof each containing the recombinant type G protein coupled receptor protein, etc. Their various applications are also disclosed herein below.

(1) Method for Determining Ligands to the G Protein Coupled

Receptor Protein

The G protein coupled receptor protein (or the peptide segment thereof) is useful as a reagent for investigating or determining a ligand to said G protein coupled receptor protein.

According to the present invention, methods for determining a ligand to the G protein coupled receptor protein

which comprises contacting the G protein coupled receptor protein or the peptide segment or fragment thereof with the compound to be tested are provided.

The compound to be tested may include not only known ligands such as angiotensins, bombesins, canavanoids, cholecystokinins, glutamine, serotonin, melatonin, neuropeptides γ , opioids, purine, vasopressins, oxytocins, VIP (vasoactive intestinal and related peptides), somatostatins, dopamine, motilins, amylin, bradykinins, CGRP (calcitonin gene related peptides), adrenomedullins, leukotrienes, pancreastatins, prostaglandins, thromboxanes, adenosine, adrenaline, α - and β -chemokines (IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HCL4, MCP-3, I-309, MIP1 α , MIP-1 β , RANTES, etc.), endothelins, enterogastrins, histamine, neurotensins, TRH, pancreatic polypeptides, galanin, modified derivatives thereof, analogues thereof, family members thereof and the like but also tissue extracts, cell culture supernatants, etc. of warm-blooded animals (such as mice, rats, swines, cattle, sheep, monkeys and human being), etc. For example, said tissue extract, said cell culture supernatant, etc. is added to the G protein coupled receptor protein for measurement of the cell stimulating activity, etc. and fractionated by relying on the measurements whereupon a single ligand can be finally obtained.

In one specific embodiment of the present invention, said method for determining the ligand includes a method for determining a compound or a salt thereof capable of stimulating a target cell which comprises binding said compound with the G protein coupled receptor protein either in the presence of the G protein coupled receptor protein or the peptide segment thereof or in a receptor binding assay system in which the expression system for the recombinant type receptor protein is constructed and used; and measuring the receptor-mediated cell stimulating activity, etc.

Examples of said cell stimulating activities include promoting activity or inhibiting activity on biological responses,

e.g. liberation of arachidonic acid, liberation of acetylcholine, liberation of endocellular Ca^{2+} , production of endocellular cAMP, production of endocellular cGMP, production of inositol phosphate, changes in the cell membrane potential, phosphorylation of endocellular protein, activation of c-fos, lowering in pH, activation of G protein, cell promulgation, etc. Examples of said compound or salt capable of stimulating the cell via binding with the G protein coupled receptor protein include peptides, proteins, nonpeptidic compounds, synthetic compounds, fermented products, etc.

In said method for determining the ligand, the characteristic feature is that when the G protein coupled receptor protein or the peptide segment thereof is contacted with the test compound, for example, the binding amount, the cell stimulating activity, etc. of the test compound to the G protein coupled receptor protein or the peptide segment thereof is measured.

In more specific embodiments of the present invention, said methods for determining the ligand includes:

① a method of determining a ligand to a G protein coupled receptor protein, which comprises contacting a labeled test compound with a G protein coupled receptor protein or a peptide segment thereof, and measuring the amount of the labeled test compound binding with said protein or salt thereof; or with said peptide fragment or salt thereof;

② a method of determining a ligand to a G protein coupled receptor protein, which comprises contacting a labeled test compound with cells containing the G protein coupled receptor protein or the membrane fraction of said cell, and measuring the amount of the labeled test compound binding with said cells or said cell fraction;

③ a method of determining a ligand to a G protein coupled receptor protein, which comprises contacting a labeled test compound with the G protein coupled receptor protein expressed on cell membranes by culturing transformants containing the DNA coding for the G protein coupled receptor protein, and measuring the amount of the labeled test compound binding with

said G protein coupled receptor protein;

⑥ a method of determining a ligand to a G protein coupled receptor protein, which comprises contacting a test compound with cells containing the G protein coupled receptor protein, and measuring the cell stimulating activity (e.g. promoting or inhibiting activity on biological responses such as liberation of arachidonic acid, liberation of acetylcholine, liberation of endocellular Ca^{2+} , production of endocellular cAMP, production of endocellular cGMP, production of inositol phosphate, changes in the cell membrane potential, phosphorylation of endocellular protein, activation of c-fos, lowering in pH, activation of G protein, cell promulgation, etc.) via the G protein coupled receptor protein; and

⑦ a method of determining a ligand to the G protein coupled receptor protein, which comprises contacting a test compound with the G protein coupled receptor protein expressed on the cell membrane by culturing transformants containing the DNA coding for the G protein coupled receptor protein, and measuring the cell stimulating activity (activity for promoting or inhibiting physiological responses such as liberation of arachidonic acid, liberation of acetylcholine, liberation of endocellular Ca^{2+} , production of endocellular cAMP, production of endocellular cGMP, production of inositol phosphate, changes in the cell membrane potential, phosphorylation of endocellular protein, activation of c-fos, lowering in pH, activation of G protein, cell promulgation, etc.) via the G protein coupled receptor protein.

Described below are specific explanations on the determining method of ligands according to the present invention which are provided only for illustrative purposes.

First, the G protein coupled receptor protein used for the method for determining the ligand may include any material so far as it contains a G protein coupled receptor protein or a peptide fragment or segment thereof (including a partial peptide thereof) or a salt thereof although it is preferable to express a large amount of G protein coupled receptor proteins in animal cells.

In the manufacture of the G protein coupled receptor protein, the above-mentioned method can be used and it may be carried out by expressing said protein encoding DNA in mammalian cells or in insect cells. With respect to the DNA fragment coding for the aimed region, complementary DNA may be used although it is not limited thereto. For example, gene fragments or synthetic DNA may be used as well.

In order to introduce the G protein coupled receptor protein-encoding DNA fragment into host animal cells and to express it efficiently, it is preferred that said DNA fragment is incorporated into the downstream site of polyhedron promoters derived from nuclear polyhedrosis virus belonging to baculovirus, promoters derived from SV40, promoters derived from retrovirus, metallothionein promoters, human heat shock promoters, cytomegalovirus promoters, SRA promoters, etc. Examinations of the quantity and the quality of the expressed receptor can be carried out by methods per se known to those of skill in the art or methods similar thereto. For example, they may be conducted by methods described in publications such as Namai, P. et al: The Journal of Biochemical Society, vol.267, pages 19555-19559 (1992).

Accordingly, with respect to the determination of the ligand, the material containing a G protein coupled receptor protein or peptide segment thereof may include products containing G protein coupled receptor proteins which are purified by methods per se known to those of skill in the art or methods similar thereto, peptide fragments of said G protein coupled receptor protein, cells containing said G protein coupled receptor protein, membrane fractions of the cell containing said protein, etc.

When the G protein coupled receptor protein-containing cell is used in the determining method of the ligand, said cell may be immobilized with binding agents including glutaraldehyde, formalin, etc. The immobilization may be carried out by methods per se known to those of skill in the art or methods similar thereto.

The G protein coupled receptor protein-

containing cells are host cells expressing the G protein coupled receptor protein. Examples of said host cells are microorganisms such as *Escherichia coli*, *Bacillus subtilis*, yeasts, insect cells, animal cells, etc.

The cell membrane fraction is a cell membrane-rich fraction which is prepared by methods per se known to those of skill in the art or methods similar thereto after disruption of cells. Examples of cell disruption may include a method for squeezing cells using a Potter-Elvehjem homogenizer, a disruption by a Waring blender or a Polytron (manufactured by Kinematica), a disruption by ultrasonic waves, a disruption via blowing out cells from small nozzles together with applying a pressure using a French press or the like, etc. In the fractionation of the cell membrane, a fractionation method by means of centrifugal force such as a fractional centrifugal separation and a density gradient centrifugal separation is mainly used. For example, disrupted cellular liquid is centrifuged at a low speed (500 rpm to 3,000 rpm) for a short period (usually, from about one to ten minutes), the supernatant liquid is further centrifuged at a high speed (1,500 rpm to 3,000 rpm) usually for 30 minutes to two hours and the resulting precipitate is used as a membrane fraction. Said membrane fraction contains a lot of the expressed G protein coupled receptor protein and a lot of membrane components such as phospholipids and membrane proteins derived from the cells.

The amount of the G protein coupled receptor protein in the membrane fraction cell containing said G protein coupled receptor protein is preferably 10^3 - 10^8 molecules per cell or, suitably, 10^5 to 10^7 molecules per cell. Incidentally, the more the expressed amount, the higher the ligand binding activity (specific activity) per membrane fraction whereby the construction of a highly sensitive screening system becomes possible and, moreover, it may enable us to measure the large amount of samples within the same lot.

In conducting the above-mentioned methods ① to ② wherein ligands capable of binding with the G protein coupled

receptor protein are determined, a suitable G protein coupled receptor fraction and a labeled test compound are necessary. The G protein coupled receptor fraction is preferably a

naturally occurring (natural type) G protein coupled receptor, a recombinant type G protein coupled receptor having the activity equivalent to that of the natural type. Here, the term "activity equivalent to" means the equivalent ligand binding activity, etc.

Suitable examples of the labeled test compound are angiotensin, bombesin, canavanine, cholecystokinin, glutamine, serotonin, melatonin, neuropeptide Y, opioid, purine, vasopressin, oxytocin, VIP (vasoactive intestinal and related peptides), somatostatin, dopamine, motilin, amylin, bradykinin, CGRP (calcitonin gene related peptides), adrenomedullin, leukotriene, pancreastatin, prostaglandin, thromboxane, adenosine, adrenaline, α - and β - chemokine (IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HC14, MCP-3, I-309, MIP-1 β , RANTES, etc.), endothelin, enterogastrin, histamine, neurotensin, TRH, pancreatic polypeptides, galanin, an analogue derivative thereof, etc. which are labeled with [3 H], [125 I], [14 C], [35 S], etc.

Specifically, the determination of ligands capable of binding with G protein coupled receptor proteins is carried out as follows:

First, cells or cell membrane fractions containing the G protein coupled receptor protein are suspended in a buffer suitable for the determining method to prepare the receptor sample in conducting the method of determining the ligand binding with the G protein coupled receptor protein. The buffer may include any buffer such as Tris-HCl buffer or phosphate buffer with pH 4-10 (preferably, pH 6-8), etc., as long as it does not inhibit the binding of the ligand with the receptor. In addition, surface-active agents such as CHAPS, Tween 80TM (Kao-Atlas, Japan), digitonin, deoxycholate, etc. and various proteins such as bovine serum albumin (BSA), gelatin, milk derivatives, etc. may be added to the buffer with an object of decreasing the non-specific binding.

- 1 4 7 -

Further, a protease inhibitor such as PMSF, leupeptin, E-64 (manufactured by Peptide Laboratory), pepstatin, etc. may be added with an object of inhibiting the decomposition of the receptor and the ligand by protease. A test compound labeled with a predetermined (or certain) amount (5,000 cpm to 500,000 cpm) of [³H], [¹²⁵I], [¹⁴C], [³⁵S], etc. is made copresent in 0.01 ml to 10 ml of said receptor solution.

In order to know the non-specific binding amount (NSB), a reaction tube to which a great excessive amount of the unlabeled test compound is added is prepared as well.

The reaction is carried out at 0-50°C (preferably at 4-37°C) for 20 minutes to 24 hours (preferably 30 minutes to three hours). After the reaction, it is filtered through a glass fiber filter or the like, washed with a suitable amount of the same buffer and the radioactivity remaining in the glass fiber filter is measured by means of a liquid scintillation counter or a gamma-counter. The test compound in which the count (B - NSB) obtained by subtracting the non-specific binding amount (NSB) from the total binding amount (B) is more than 0 cpm can be selected as a ligand to the G protein coupled receptor protein of the present invention.

In conducting the above-mentioned methods ④ to ⑤ wherein ligands capable of binding with the G protein coupled receptor protein are determined, the cell stimulating activity (e.g. the liberation of arachidonic acid, the liberation of acetylcholine, endocellular Ca²⁺ liberation, endocellular CAMP production, the production of inositol phosphate, changes in the cell membrane potential, the phosphorylation of endocellular protein, the activation of c-fos, lowering of pH, the activation of G protein, cell promulgation, etc.) mediated by the G protein coupled receptor protein may be measured by known methods or by the use of commercially available measuring kits. To be more specific, G protein coupled receptor protein-containing cells are at first cultured in a multi-well plate or the like.

In conducting the determination of ligand, it is substituted with a fresh medium or a suitable buffer which

- 1 4 8 -

does not show toxicity to the cells in advance of the experiment, and incubated for certain period after adding a test compound, etc. thereto. Then, the cells are extracted or the supernatant liquid is recovered and the resulting product is determined by each of the methods. When it is difficult to identify the production of the substance (e.g. arachidonic acid) which is to be an index for the cell stimulating activity due to the decomposing enzyme contained in the cell, an assay may be carried out by adding an inhibitor against said decomposing enzyme. With respect to the activity such as an inhibitory action against CAMP production, it may be detected as an inhibitory action against the production of the cells whose fundamental production is increased by forskolin or the like.

The kit used for the method of determining the ligand binding with the G protein coupled receptor protein includes a G protein coupled receptor protein or a peptide fragment thereof, cells containing the G protein coupled receptor protein, a membrane fraction from the cells containing the G protein coupled receptor protein, etc.

Examples of the kit for determining the ligand are as follows:

1. Reagent for Determining the Ligand.

① Buffer for Measurement and Buffer for Washing.

The buffering product wherein 0.05% of bovine serum

albumin (manufactured by Sigma) is added to Hanks' Balanced

Salt Solution (manufactured by Gibco).

This product may be sterilized by filtration through a membrane filter with a 0.45 μ m pore size, and stored at 4°C or may be formulated upon use.

② G Protein Coupled Receptor Protein Sample.

CHO cells in which G protein coupled receptor proteins are expressed are subcultured at the rate of 5×10^5 cells/well in a 12-well plate and cultured at 37°C in a humidified 5% CO₂/95% air atmosphere for two days to prepare the sample.

③ Labeled Test Compound.

The compound which is labeled with commercially

available [^3H], [^{125}I], [^{14}C], [^{35}S], etc. or labeled with a suitable method.

The product in a state of an aqueous solution is stored at 4°C or at -20°C and, upon use, diluted to $1\ \mu\text{M}$ with a buffer for the measurement. In the case of the test compound which is hardly soluble in water, it is dissolved in dimethylformamide, DMSO, methanol, etc.

④ Unlabeled Test Compound.

The same compound for the labeled one is prepared in a concentration of 100 to 1,000-fold concentrated state.

2. Method of Measurement.

① G protein coupled receptor protein-expressing CHO cells cultured in a 12-well tissue culture plate are washed twice with 1 ml of buffer for the measurement and then $490\ \mu\text{l}$ of buffer for the measurement is added to each well.

② Five μl of the labeled test compound is added and the mixture is made to react at room temperature for one hour. For measuring the nonspecific binding amount, $5\ \mu\text{l}$ of the unlabeled test compound is added.

③ The reaction solution is removed from each well, which is washed with 1 ml of a buffer for the measurement three times. The labeled test compound which is binding with the cells is dissolved in 0.2N NaOH -1% SDS and mixed with 4 ml of a liquid scintillator A (manufactured by Wako Pure Chemical, Japan).

④ Radioactivity is measured using a liquid scintillation counter (manufactured by Beckmann).

The ligand which can bind with the G protein coupled receptor protein include substances occurring or existing, for example, in brain, pituitary gland, pancreas, etc. Examples of the ligand are angiotensin, bombesin, canavaninoid, cholecystokinin, glutamine, serotonin, melatonin, neuropeptide Y, opioid, purine, vasopressin, oxytocin, VIP (vasoactive intestinal and related peptide), somatostatin, dopamine, motilin, amylin, bradykinin, CGRP (calcitonin gene related

peptide), adrenomedullin, leukotriene, pancreastatin, prostaglandin, thromboxane, thromboxatin, adenosine, adenaline, α - and β -chemokine (IL-8, GRO α , GRO β , GRO γ , NAP-2, ENA-78, PF4, IP10, GCP-2, MCP-1, HCL14, MCP-3, I-309, MIP1 α , MIP-1 β , RANTES, etc.), endothelin, enterogastrin, histamine, neurotensin, TRH, pancreatic polypeptide, galanin, modified derivatives thereof, analogues thereof, etc.

Since the receptor protein encoded by pMAH2-17 is highly homologous to prinoceptors, it is considered that the are strong possibility of a subtype within prinoceptor families. All data including electrophysiological measurements are supporting that the mouse pancreatic β -cell strain, MIN6-derived receptor protein of the present invention (e.g., SEQ ID NO: 38 and SEQ ID NO: 39, or proteins encoded by pMAH2-17) is a novel purinoceptor subtype. In other words, it is suggested that the ligand capable of binding with the mouse pancreatic β -cell strain, MIN6-derived receptor protein of the present invention (e.g., SEQ ID NO: 38 and SEQ ID NO: 39, or proteins encoded by pMAH2-17) is a purine compound such as ATP. Further, the receptor protein (e.g., SEQ ID NO: 56, or proteins encoded by pMAH2-17) is considered to be a novel human type purinoceptor. It is presumed that it is advantageously useful in efficiently screening for agonists or antagonists to receptor proteins which control or regulate functions in the central nervous system or immune system, related to purine compounds, and in developing pharmaceuticals.

(2) Preventive and Therapeutic Agent for of G Protein Conjugated Receptor Protein Deficiency Diseases

If a ligand to the G protein coupled receptor protein is disclosed via the aforementioned method (1), the G protein coupled receptor protein-encoding DNA can be used a preventive and/or therapeutic agent for treating said G protein coupled receptor protein deficiency diseases depending upon the action that said ligand exerts.

For example, when there is a patient for whom the physiological action of the ligand cannot be expected because

- 1 5 1 -

of a decrease in the G protein coupled receptor protein in vivo, the amount of the G protein coupled receptor protein in the brain cells of said patient can be increased whereby the action of the ligand can be fully achieved by:

- (a) administering the G protein coupled receptor protein-encoding DNA to the patient to express it; or
 - (b) inserting the G protein coupled receptor protein-encoding DNA into brain cells or the like to express it, followed by transplanting said brain cells or the like to said patient.
- Accordingly, the G protein coupled receptor protein-encoding DNA can be used as a safe and less toxic preventive and therapeutic agent for the G protein coupled receptor protein deficiency diseases. In an embodiment, it is suggested that the ligands capable of binding with the mouse pancreatic β -cell strain, MIN6-derived receptor protein of the present invention (e.g., SEQ ID NO: 38 and SEQ ID NO: 39, or proteins encoded by pMAH2-17) and further with the human-derived receptor protein of the present invention (e.g., SEQ ID NO: 56, or proteins encoded by pHAH2-17) are purine compounds such as ATP. Therefore, the disease to be treated may include diseases or syndromes in connection with purine ligand compounds. Examples of such diseases may include cancer, immunodeficiency, autoimmune disease, rheumatoid arthritis, rejection on internal organ transplant, hypertension, diabetes, cystic fibrosis, hypotension, incontinence of urine, pain, etc.

(3) Preventive and Therapeutic Pharmaceutical Composition for Human-Derived G Protein Conjugated Receptor Protein Deficiency Diseases

If the human-derived G protein coupled receptor protein-encoding DNA is screened and a ligand for said human-derived G protein coupled receptor protein can be clarified using the above-mentioned method (1), the human-derived G protein coupled receptor protein-encoding DNA can be used as an agent for the prevention or therapy of the deficiency diseases of said human-derived G protein coupled receptor protein depending upon the action that said ligand exhibits.

- 1 5 2 -

For example, when there is a patient for whom the physiological action of the ligand cannot be expected because of a decrease in the G protein coupled receptor protein in vivo, the amount of the G protein coupled receptor protein in the brain cells of said patient can be increased whereby the action of the ligand can be fully achieved by:

- (a) administering the G protein coupled receptor protein-encoding DNA to the patient to express it; or
 - (b) inserting the G protein coupled receptor protein-encoding DNA into brain cells or the like to express it, followed by transplanting said brain cells or the like to said patient.
- Accordingly, the G protein coupled receptor protein-encoding DNA can be used as a safe and less toxic preventive and therapeutic agent for the G protein coupled receptor protein deficiency diseases.

When the G protein coupled receptor protein-encoding DNA is used as the above-mentioned agent, said DNA may be used alone or after inserting it into a suitable vector such as retrovirus vector, adenovirus vector, adenovirus-associated virus vector, etc. followed by subjecting the product vector to a conventional means. Thus, it may be administered orally parenterally, by inhalation spray, rectally, or topically as pharmaceutical compositions or formulations. Oral formulations include tablets (sugar-coated if necessary), capsules, elixirs, microcapsules, etc. Parenteral formulations include injections such as an aseptic solution or a suspension in water or in other pharmaceutically acceptable liquid. For example, the DNA of the present invention is admixed in a unit dose form which is required for preparing generally approved pharmaceutical preparations together with a physiologically acceptable carriers, flavoring agents, adjuvants, excipients, diluents, fillers, vehicles, antiseptics, stabilizers, binders, etc. whereupon the preparation can be manufactured. The amount of the effective component in those preparations is to be in such an extent that the suitable dose within an indicated range is achieved.

Examples of the additives which can be admixed in the

tablets, capsules, etc. are binders such as gelatin, corn starch, tragacanth and gum arabicum; fillers such as crystalline cellulose; swelling agents such as corn starch, gelatin and alginic acid; lubricating agents such as magnesium stearate; sweetening agents such as sucrose, lactose and saccharine; and flavoring agents such as pepper mint, akamono oil and cherry. When the unit dose form of the preparation is a capsule, a liquid carrier such as fat/oil may be further added in addition of the above-mentioned types of materials.

The aseptic composition for injection may be formulated by conventional practices for the preparations such as that the active substance in a vehicle such as water for injection is dissolved or suspended in naturally occurring plant oil such as sesame oil and palm oil.

Examples of an aqueous liquid for injection are a physiological saline solution and isotonic solutions containing glucose and other auxiliary agents (e.g. D-sorbitol, D-mannitol, sodium chloride, etc.) wherein a suitable auxiliary solubilizers such as alcohol (e.g. ethanol, etc.), polyalcohol (e.g. propylene glycol polyethylene glycol, etc.), nonionic surface-active agent (e.g. Polysorbate 80TM, HCO-50, etc.), etc. may be jointly used. Examples of an oily liquid include sesame oil, soybean oil, etc. wherein benzyl benzoate, benzyl alcohol, etc. may be jointly used as auxiliary solubilizers. In addition, buffers (e.g. phosphate buffer, sodium acetate buffer, etc.), analgesic agents (e.g. benzalkonium chloride, procaine hydrochloride, etc.), stabilizers (e.g. human serum albumin, polyethylene glycol, etc.), stabilizers (e.g. benzyl alcohol phenol, etc.), antioxidants, etc. may be admixed therewith too. The prepared injection solution is filled in suitable ampoules. The preparation prepared as such is safe and less toxic and, therefore, it can be administered to warm-blooded animals (e.g., rat, rabbit, sheep, swine, cattle, cat, dog, monkey, human beings, etc.).

Specific dose levels of said DNA may vary depending upon a variety of factors including the activity of drugs employed, the age, body weight, general health, sex, diet,

time of administration, route of administration, drug combination, and the severity of the symptom. In the case of oral administration, it is usually about 0.1-100 mg, preferably about 1.0-50 mg or, more preferably, about 1.0-20 mg per day for adults (as 60 kg). When it is administered parenterally, its dose at a time may vary depending upon the object (patient) to be administered, organs to be administered, symptoms, administering methods, etc. but, in the case of injections, it is usually convenient to give by an intravenous route in an amount of about 0.01-30 mg, preferably about 0.1-20 mg or, more preferably, about 0.1-10mg per day to adults (as 60 kg). In the case of other animals, the dose calculated for 60 kg may be administered as well.

(4) Quantitative Determination of Ligand to the G Protein Conjugated Receptor Protein of the Present Invention.

The G protein coupled receptor protein or a peptide fragment thereof has a binding property to ligand and, therefore, it is capable of determining quantitatively an amount of ligands *in vivo* with good sensitivity.

This quantitative determination may be carried out by, for example, combining with a competitive method. Thus, samples to be determined is contacted with G protein coupled receptor proteins or peptide fragments thereof so that the ligand concentration in said sample can be determined. In one embodiment of the quantitative determination, the protocols described in the following ① and ② or the methods similar thereto may be used:

① Hiroshi Irie (ed): "Radioimmunoassay" (Kodansha, Japan, 1974); and

② Hiroshi Irie (ed): "Radioimmunoassay, Second Series" (Kodansha, Japan, 1979).

(5) Screening of Compound Inhibiting the Binding of Ligand with the G Protein Conjugated Receptor Protein of the Present Invention.

G Protein coupled receptor proteins or peptide

fragments thereof are used. Alternatively, expression systems for recombinant type G Protein coupled receptor proteins or peptide fragments thereof are constructed and receptor binding assay systems using said expression system are used. In these assay systems, it is possible to screen compounds (e.g. peptides, proteins, nonpeptidic compounds, synthetic compounds, fermented products, cell extracts, plant extracts, animal tissue extracts, etc.) or salts thereof which inhibits the binding of a ligand with the G protein coupled receptor protein. Such a compound includes a compound exhibiting a G protein coupled receptor-mediated cell stimulating activity (e.g. activity of promoting or activity of inhibiting physiological reactions including liberation of arachidonic acid, liberation of acetylcholine, endocellular Ca^{2+} liberation, endocellular cAMP production, endocellular cGMP production, production of inositol phosphate, changes in cell membrane potential, phosphorylation of endocellular proteins, activation of c-fos, lowering of pH, activation of G protein, cell promulgation, etc.) (so-called "G protein coupled receptor-agonist"), a compound free of such a cell stimulating activity (so-called "G protein coupled receptor-antagonist"), etc.

Thus, the present invention provides a method of screening a compound which inhibits the binding of a ligand with a G protein coupled receptor protein or a salt thereof, characterized in comparing the following two cases:

(i) the case wherein the ligand is contacted with the G protein coupled receptor protein or salt thereof, or a peptide fragment thereof or a salt thereof; and
(ii) the case wherein the ligand is contacted with a mixture of the G protein coupled receptor protein or salt thereof or the peptide fragment or salt thereof and said test compound.

In said screening method, one characteristic feature of the present invention resides in that the amount of the ligand bonded with said G protein coupled receptor protein or the peptide fragment thereof, the cell stimulating activity of the ligand, etc. are measured in the case where (i) the

ligand is contacted with G protein coupled receptor proteins or peptide fragments thereof and in the case where (ii) the ligand and the test compound are contacted with the G protein coupled receptor protein or the peptide fragment thereof, respectively and then compared therebetween.

In one more specific embodiment of the present invention, the following is provided:

① a method of screening a compound or a salt thereof which inhibits the binding of a ligand with a G protein coupled receptor protein, characterized in that, when a labeled ligand is contacted with a G protein coupled receptor protein or a peptide fragment thereof and when a labeled ligand and a test compound are contacted with a G protein coupled receptor protein or a peptide fragment thereof, the amounts of the labeled ligand bonded with said protein or peptide fragment thereof or salt thereof are measured and compared;

② a method of screening a compound or a salt thereof which inhibits the binding of a ligand with a G protein coupled receptor protein, characterized in that, when a labeled ligand is contacted with cells containing G protein coupled receptor proteins or a membrane fraction of said cells and when a labeled ligand and a test compound are contacted with cells containing G protein coupled receptor proteins or a membrane fraction of said cells, the amounts of the labeled ligand binding with said protein or peptide fragment thereof or salt thereof are measured and compared;

③ a method of screening a compound or a salt thereof which inhibits the binding of a ligand with a G protein coupled receptor protein, characterized in that, when a labeled ligand is contacted with G protein coupled receptor proteins expressed on the cell membrane by culturing a transformant containing a G protein coupled receptor protein encoding DNA and when a labeled ligand and a test compound are contacted with G protein coupled receptor proteins expressed on the cell membrane by culturing a transformant containing a G protein coupled receptor protein encoding DNA, the amounts of the labeled ligand binding with said G protein coupled receptor

protein are measured and compared;

④ a method of screening a compound or a salt thereof which inhibits the binding of a ligand with a G protein coupled receptor protein, characterized in that, when a G protein coupled receptor protein-activating compound (e.g. a ligand to the G protein coupled receptor protein) is contacted with cells containing G protein coupled receptor proteins and when the G protein coupled receptor protein-activating compound and a test compound are contacted with cells containing G protein coupled receptor proteins, the resulting G protein coupled receptor protein-mediated cell stimulating activities (e.g. activities of promoting or activities of inhibiting physiological responses including liberation of arachidonic acid, liberation of acetylcholine, endocellular Ca^{2+} liberation, endocellular cAMP production, endocellular cGMP production, production of inositol phosphate, changes in cell membrane potential, phosphorylation of endocellular proteins, activation of c-fos, lowering of pH, activation of G protein, cell promulgation, etc.) are measured and compared; and

⑤ a method of screening a compound or a salt thereof which inhibits the binding of a ligand with a G protein coupled receptor protein, characterized in that, when a G protein coupled receptor protein-activating compound (e.g. a ligand to the G protein coupled receptor protein) is contacted with G protein coupled receptor proteins expressed on cell membranes by culturing transformants containing G protein coupled receptor protein-encoding DNA and when a G protein coupled receptor protein-activating compound and a test compound are contacted with the G protein coupled receptor protein expressed on the cell membrane by culturing the transformant containing the G protein coupled receptor protein-encoding DNA, the resulting G protein coupled receptor protein-mediated cell stimulating activities (activities of promoting or activities of inhibiting physiological responses such as liberation of arachidonic acid, liberation of acetylcholine, endocellular Ca^{2+} liberation, endocellular cAMP production, endocellular cGMP production, production of inositol phosphate, changes in cell

membrane potential, phosphorylation of endocellular proteins, activation of c-fos, lowering of pH, activation of G protein, and cell promulgation) are measured and compared.

Before the G protein coupled receptor protein of the present invention was obtained, the G protein coupled receptor agonist or antagonist had to be screened by, first, obtaining a candidate compound by using G protein coupled receptor protein-containing cells, tissues or cell membrane fractions derived from rat or the like (primary screening) and, then, making sure whether the candidate compound really inhibits the binding between human G protein coupled receptor proteins and ligands (secondary screening). Other receptor proteins inevitably exist when the cells, the tissues or the cell membrane fractions are used as they are, whereby they intrinsically make it difficult to screen agonists or antagonists to the desired receptor proteins. By using the human-derived G protein coupled receptor protein, however, there is no need of effecting the primary screening, whereby it is allowable to efficiently screen a compound that inhibits the binding between a ligand and a G protein coupled receptor. Besides, it is allowable to evaluate whether the compound that is screened is a G protein coupled receptor agonist or a G protein coupled receptor antagonist.

Specific explanations of the screening method will be given as hereunder.

First, with respect to the G protein coupled receptor protein used for the screening method of the present invention, any product may be used so far as it contains G protein coupled receptor proteins or peptide fragment thereof although the use of a membrane fraction of mammalian organs is suitable. However, human organs is extremely hardly available and, accordingly, G protein coupled receptor proteins which are expressed in a large amount using a recombinant are suitable for the screening.

In the manufacture of the G protein coupled receptor protein, the above-mentioned method can be used and it may be carried out by expressing the DNA coding for said protein in

mammalian cells or in insect cells. With respect to the DNA fragment coding for the target region, complementary DNA may be used although it is not limited thereto. Thus, for example, gene fragments or synthetic DNA may be used as well.

In order to introduce the G protein coupled receptor protein-encoding DNA fragment into host animal cells and to express it efficiently, it is preferred that said DNA fragment is incorporated into the downstream of polyhedron promoter of nuclear polyhedrosis virus belonging to baculovirus, promoter derived from SV40, promoter of retrovirus, metallothionein promoter, human heat shock promoter, cytomegalovirus promoter, SRA promoter, etc. Examinations of the quantity and the quality of expressed receptors can be carried out by known methods per se or modified methods substantially analogous thereto. For example, they may be conducted by the method described in publications such as Nambi, P. et al.: The Journal of Biochemical Society, vol.267, pages 19555-19559 (1992).

Accordingly, in the screening method, the substance containing a G protein coupled receptor protein or a peptide fragment thereof may be a G protein coupled receptor protein which is purified by known methods per se or a G protein coupled receptor protein fragment which is purified by known methods per se, or a cell containing said protein or a cell membrane fraction of the cell containing said protein, etc.

When the G protein coupled receptor protein-containing cells are used in the screening method, said cells may be immobilized with glutaraldehyde, formalin, etc. The immobilization may be carried out by known methods per se or modified methods substantially analogous thereto.

The G protein coupled receptor protein-containing cells are host cells expressing the G protein coupled receptor protein. Examples of said host cells may include *Escherichia coli*, *Bacillus subtilis*, yeasts, insect cells, animal cells such as CHO cell and COS cell, etc.

Cell membrane fractions are fractions which contain a lot of cell membranes prepared by known methods per se or

modified methods substantially analogous thereto after disrupting or crushing the cells. Examples of disruptions of the cell may include methods by squeezing the cells with a Potter-Elvehjem homogenizer, disrupting or crushing by a Waring blender or a Polytron (manufactured by Kinematica), disrupting or crushing by means of ultrasonic wave, disrupting by blowing out the cells from small nozzles together with applying a pressure with a French press or the like, etc.

Fractionation of the cell membrane is carried out mainly by fractionation techniques by means of centrifugal force such as a fractional centrifugal separation and a density gradient centrifugal separation. For example, disrupted liquid of cells is centrifuged at a low speed (500 rpm to 3,000 rpm) for a short period (usually, from about one to ten minutes), the supernatant liquid is further centrifuged at a high speed (1,500 rpm to 3,000 rpm) usually for 30 minutes to two hours and the resulting precipitate is used as a membrane fraction. Said membrane fraction contains a lot of expressed G protein coupled receptor proteins and membrane components such as phospholipids and membrane proteins derived from the cells.

The amount of the G protein coupled receptor protein in the G protein coupled receptor protein-containing cell and in the cell membrane fraction obtained from the cell is preferably 10^3 - 10^8 molecules per cell or, suitably, 10^5 to 10^7 molecules per cell. Incidentally, the more the expressed amount, the higher the ligand binding activity (specific activity) per membrane fraction whereby the construction of a highly sensitive screening system is possible and, moreover, it is possible to measure the large amount of samples in the same lot.

In conducting the above-mentioned methods ① to ③ for screening the compound capable of inhibiting the binding of the ligand with the G protein coupled receptor protein, a suitable G protein coupled receptor fraction and a labeled ligand are necessary. With respect to the G protein coupled receptor fraction, it is preferred to use naturally occurring G protein coupled receptors (natural type G protein coupled

receptors) or recombinant type G protein coupled receptor fractions with the activity equivalent to that of the natural type G protein coupled. Here the term "activity equivalent to" means the same ligand binding activity, or the substantially equivalent ligand binding activity.

With respect to the labeled ligand, it is possible to use labeled ligands, labeled ligand analogized compounds, etc. For example, ligands labeled with [^3H], [^{125}I], [^{14}C], [^{35}S], etc. and other labeled substances may be utilized.

Specifically, G protein coupled receptor protein-containing cells or cell membrane fractions are first suspended in a buffer which is suitable for the determining method to prepare the receptor sample in conducting the screening for a compound which inhibits the binding of the ligand with the G protein coupled receptor protein.

With respect to the buffer, any buffer such as Tris-HCl buffer or phosphate buffer of pH 4-10 (preferably, pH 6-8) which does not inhibit the binding of the ligand with the receptor may be used.

In addition, a surface-active agent such as CHAPS, Tween 80TM (Kao-Atlas, Japan), digitonin, deoxycholate, etc. and/or various proteins such as bovine serum albumin (BSA), gelatine, etc. may be added to the buffer with an object of decreasing the nonspecific binding. Further, a protease inhibitor such as PMSF, leupeptin, E-64 (manufactured by Peptide Laboratory, Japan), pepstatin, etc. may be added with an object of inhibiting the decomposition of the receptor and the ligand by protease. A labeled ligand in a certain amount (5,000 cpm to 500,000 cpm) is added to 0.01 ml to 10 ml of said receptor solution and, at the same time, 10^{-4} M to 10^{-10} M of a test compound is made copresent. In order to determine the nonspecific binding amount (NSB), a reaction tube to which a great excessive amount of unlabeled test compounds is added is prepared as well.

The reaction is carried out at 0-50°C (preferably at 4-37°C) for 20 minutes to 24 hours (preferably 30 minutes to three hours). After the reaction, it is filtered through a

glass fiber filter, a filter paper, or the like, washed with a suitable amount of the same buffer and the radioactivity retained in the glass fiber filter, etc. is measured by means of a liquid scintillation counter or a gamma-counter.

Supposing that the count (B_0 - NSB) obtained by subtracting the nonspecific binding amount (NSB) from the total binding amount (B_0) wherein an antagonizing substance is not present is set at 100%, the test compound in which the specific binding amount (B - NSB) obtained by subtracting the nonspecific binding amount (NSB) from the total binding amount (B) is, for example, less than 50% may be selected as a candidate ligand to the G protein coupled receptor protein of the present invention.

In conducting the above-mentioned methods ④ to ⑥ for screening the compound which inhibits the binding of the ligand with the G protein coupled receptor protein, the G protein coupled receptor protein-mediated cell stimulating activity (e.g. activities of promoting or activities of inhibiting physiological responses such as liberation of arachidonic acid, liberation of acetylcholine, endocellular Ca^{2+} liberation, endocellular cAMP production, production of inositol phosphate, changes in the cell membrane potential, phosphorylation of endocellular proteins, activation of c-fos, lowering of pH, activation of G protein and cell promulgation, etc.) may be measured by known methods or by the use of commercially available measuring kits. To be more specific, G protein coupled receptor protein-containing cells are at first cultured in a multiwell plate or the like.

In conducting the screening, it is substituted with a suitable buffer which does not show toxicity to fresh media or cells in advance, incubated for a certain period after adding a test compound, etc. thereto. The resultant cells are extracted or the supernatant liquid is recovered and the resulting product is determined, preferably quantitatively, by each of the methods. When it is difficult to identify the production of the index substance (e.g. arachidonic acid, etc.) which is to be an index for the cell stimulating activity due to the presence of decomposing enzymes contained in the cell,

- 1 6 3 -

an assay may be carried out by adding an inhibitor against said decomposing enzyme. With respect to the activities such as an inhibitory action against cAMP production, it may be detected as an inhibitory action against the cAMP production in the cells whose fundamental production has been increased by forskolin or the like.

In conducting a screening by measuring the cell stimulating activity, cells in which a suitable G protein coupled receptor protein is expressed are necessary.

Preferred G protein coupled receptor protein-expressing cells are naturally occurring G protein coupled receptor protein (natural type G protein coupled receptor protein)-containing cell lines or strains (e.g. mouse pancreatic β cell line, MIN6, etc.), the above-mentioned recombinant type G protein coupled receptor protein-expressing cell lines or strains, etc.

Examples of the test compound includes peptides, proteins, non-peptidic compounds, synthesized compounds, fermented products, cell extracts, plant extracts, animal tissue extracts, serum, blood, body fluid, etc. Those compounds may be novel or known.

A kit for screening the compound which inhibits the binding of the ligand with the G protein coupled receptor protein or a salt thereof of the present invention comprises a G protein coupled receptor protein or a peptide fragment thereof, or G protein coupled receptor protein-containing cells or cell membrane fraction thereof.

Examples of the screening kit include as follows:

1. Reagent for Determining Ligand.

① Buffer for Measurement and Buffer for Washing.

The product wherein 0.05% of bovine serum albumin

(manufactured by Sigma) is added to Hanks' Balanced Salt Solution (manufactured by Gibco).

This may be sterilized by filtration through

a membrane filter with a 0.45 μ m pore size, and stored at 4°C or may be prepared upon use.

- 1 6 4 -

② Sample of G Protein Conjugated Receptor Protein.

CHO cells in which a G protein coupled receptor protein is expressed are subcultured at the rate of 5×10^2 cells/well in a 12-well plate and cultured at 37°C with a 5% CO₂ and 95% air atmosphere for two days to prepare the sample.

③ Labeled Ligand.

The ligand which is labeled with commercially available [³H], [¹²⁵I], [¹⁴C], [³⁵S], etc.

The product in a state of an aqueous solution is stored at 4°C or at -20°C and, upon use, diluted to 1 μ M with a buffer for the measurement.

④ Standard Ligand Solution.

Ligand is dissolved in PBS containing 0.1% of bovine serum albumin (manufactured by Sigma) to make 1 mM and stored at -20°C.

2. Method of the Measurement.

① CHO cells are cultured in a 12-well tissue culture plate to express G protein coupled receptor proteins. The G protein coupled receptor protein-expressing CHO cells are washed with 1 ml of buffer for the measurement twice. Then 490 μ l of buffer for the measurement is added to each well.

② Five μ l of a test compound solution of 10^{-3} to 10^{-10} M is added, then 5 μ l of a labeled ligand is added and is made to react at room temperature for one hour. For knowing the non-specific binding amount, 5 μ l of the ligand of 10^{-3} M is added instead of the test compound.

③ The reaction solution is removed from the well, which is washed with 1 ml of buffer for the measurement three times. The labeled ligand binding with the cells is dissolved in 0.2N NaOH-1% SDS and mixed with 4 ml of a liquid scintillator A (manufactured by Wako Pure Chemical, Japan).

④ Radioactivity is measured using a liquid scintillation counter (manufactured by Beckmann) and PMB (percent of maximum binding) is calculated by the following expression:

$$\text{PMB} = \frac{[(B - \text{NSB}) / (B_0 - \text{NSB})] \times 100}{35}$$

PMB: Percent of maximum binding

B: Value when a sample is added

NSB: Nonspecific binding

B₀: Maximum binding

5 The compound or a salt thereof obtained by the screening method or by the screening kit is a compound which inhibits the binding of a ligand with a G protein coupled receptor protein and, more particularly, it is a compound having a cell stimulating activity mediated via a G protein coupled receptor or a salt thereof (so-called "G protein coupled receptor agonist") or a compound having no said stimulating activity (so-called "G protein coupled receptor antagonist"). Examples of said compound are peptides, proteins, non-peptidic compounds, synthesized compounds, fermented products, etc. and the compound may be novel or known.

Said G protein coupled receptor agonist has the same physiological action as the ligand to the G protein coupled receptor protein has and, therefore, it is useful as a safe and less toxic pharmaceutical composition depending upon said ligand activity.

On the other hand, said G protein coupled receptor antagonist is capable of inhibiting the physiological activity of the ligand to the G protein coupled receptor protein and, there fore, it is useful as a safe and less toxic pharmaceutical composition for inhibiting said ligand activity.

It is also strongly suggested that agonists and/or antagonists related to the receptor encoded by pMAH2-17 obtained in Example 19 and/or the receptor encoded by pMAH2-17 obtained in Example 21 would be useful in therapeutic or prophylactic treatment of diseases or syndromes in connection with purine ligand compounds or related analogues. It is expected that the agonists of the receptor encoded by pMAH2-17 and/or of the receptor encoded by pMAH2-17 are useful as an immunomodulator or an antitumor agent, in addition they are

5 useful in therapeutically or prophylactically treating hypertension, diabetes, cystic fibrosis, etc. It is still expected that the antagonists of the receptor encoded by pMAH2-17 and/or of the receptor encoded by pMAH2-17 are useful as hypotensive agents, analgesics, agents for therapeutically or prophylactically treating incontinence of urine, etc. With regard to purinoceptors, the mutation of conserved basic amino acid residues in the 6th or 7th putative transmembrane domain of purinoceptors introduces alteration into the receptor's responses to ATP (J. Biol. Chem., Vol. 270(9), pp. 4185-4188 (1995)). It is suggested that ATP is related to blood pressure control and circular systems via receptors (Circulation Research, Vol. 58(3), pp. 319-330 (1986)) and that ATP and purinoceptors are closely related (Am. Phys. Soc., pp. C577-C606 (1993)).

When the compound or the salt thereof obtained by the screening method or by the screening kit is used as the above-mentioned pharmaceutical composition, a conventional means may be applied therefor. The compound or the salt thereof may be orally, parenterally, by inhalation spray, rectally, or topically administered as pharmaceutical compositions or formulations (e.g. powders, granules, tablets, pills, capsules, injections, syrups, emulsions, elixirs, suspensions, solutions, etc.). For example, it may be used by an oral route as tablets (sugar-coated if necessary), capsules, elixirs, microcapsules, etc. or by a parenteral route as injections such as an aseptic solution or a suspension in water or in other pharmaceutically acceptable liquid. The pharmaceutical compositions or formulations may comprise at least one such compound alone or in admixture with pharmaceutically acceptable carriers, adjuvants, vehicles, excipients and/or diluents. The pharmaceutical compositions can be formulated in accordance with conventional methods. For example, said compound or the salt thereof is mixed in a unit dose form which is required for preparing a generally approved pharmaceutical preparations together with a

physiologically acceptable carriers, flavoring and/or perfuming agents (fragrances), fillers, vehicles, antiseptics, stabilizers, binders, etc. whereupon the preparation can be manufactured. An amount of the effective component in those preparations is to be in such an extent that the suitable dose within an indicated range is achieved.

Examples of the additives which can be admitted in the tablets, capsules, etc. are binders such as gelatin, corn starch, tragacanth and gum arabicum; fillers such as crystalline cellulose; swelling agents such as corn starch, gelatin and alginic acid; lubricants such as magnesium stearate; sweetening agents such as sucrose, lactose and saccharine; preservatives such as parabens and sorbic acid; antioxidants such as ascorbic acid, α -tocopherol and cysteine; fragrances such as peppermint, akamono oil and cherry; disintegrants; buffering agents; etc. Other additives may include mannitol, maltitol, dextran, agar, chitin, chitosan, pectin, collagen, casein, albumin, synthetic or semi-synthetic polymers, glyceride, lactide, etc. When the unit form of the preparation is a capsule, a liquid carrier such as fat/oil may be further added besides the above-mentioned types of materials. The aseptic composition for injection may be formulated by a conventional technique or practice for the preparations such as that the active substance in a vehicle such as water for injection is dissolved or suspended in a naturally occurring plant oil such as sesame oil and palm oil.

Examples of an aqueous liquid for the injection are a physiological saline solution and isotonic solutions containing glucose and other auxiliary agents (e.g. D-sorbitol, D-mannitol, sodium chloride, etc.) wherein a suitable auxiliary solubilizers such as alcohol (e.g. ethanol, etc.), polyalcohol (e.g. propylene glycol, polyethylene glycol, etc.), nonionic surface-active agent (e.g. Polysorbate 80TM, HCO-50, etc.), etc. may be jointly used. In the case of the oily liquid, sesame oil, soybean oil, etc. may be exemplified wherein benzyl benzoate, benzyl alcohol, etc. may be jointly used as auxiliary solubilizers.

In addition, buffers (e.g. phosphate buffer, sodium acetate buffer, etc.), analgesic agents (e.g. benzalkonium chloride, procaine hydrochloride, etc.), stabilizers (e.g. human serum albumin, polyethylene glycol, etc.), stabilizers (e.g. benzyl alcohol, phenol, etc.), antioxidants, etc. may be compounded therewith too. The prepared injection solution is filled in suitable ampoules. The formulation prepared as such is safe and less toxic and, therefore, it can be administered to warm-blooded mammals such as rats, rabbits, sheep, swines, cattle, cats, dogs, monkeys, human being, etc.

Dose levels of said compound or the salt thereof may vary depending upon the symptom. Specific dose levels for any particular patient will be employed depending upon a variety of factors including the activity of specific compounds employed, the age, body weight, general health, sex, diet, time of administration, route of administration, rate of excretion, drug combination, and the severity of the particular disease undergoing therapy. In the case of oral administration, it is usually about 0.1-100 mg, preferably about 1.0-50 mg or, more preferably, about 1.0-20 mg per day for adults (as 60 kg). When it is administered parenterally, its dose at a time may vary depending upon the object to be administered, organs to be administered, symptoms, administering methods, etc. The term "parenteral" as used herein includes subcutaneous injections, intravenous, intramuscular, intraperitoneal injections, or infusion techniques. In the case of injections, it is usually convenient to give by an intravenous route in an amount of about 0.01-30 mg, preferably about 0.1-20 mg or, more preferably, about 0.1-10 mg per day to adults (as 60 kg). In the case of other animals, the dose calculated for 60 kg may be administered as well.

(6) Manufacture of Antibody or Antiserum against the G Protein Coupled Receptor Protein of the Present Invention,
Its Peptide Fragment or Its Salt.
Antibodies (e.g. polyclonal antibody and monoclonal antibody) and antisera against the G protein coupled receptor

protein or salt thereof of the present invention or against the peptide fragment of the G protein coupled receptor protein or salt thereof of the present invention may be manufactured by antibody or antiserum-manufacturing methods per se known to those of skill in the art or methods similar thereto, using the G protein coupled receptor protein or its salt of the present invention or the peptide fragment of the G protein coupled receptor protein or its salt of the present invention.

For example, monoclonal antibodies can be manufactured by the method as given below.

[Preparation of Monoclonal Antibody]

(a) Preparation of Monoclonal Antibody-Producing Cells.

The G protein coupled receptor protein of the present invention or its salt or the peptide fragment of the G protein coupled receptor protein of the present invention or its salt (hereinafter, may be abbreviated as the "G protein coupled receptor protein") is administered to warm-blooded animals either solely or together with carriers or diluents to the site where the production of antibody is possible by the administration. In order to potentiate the antibody productivity upon the administration, complete Freund's adjuvants or incomplete Freund's adjuvants may be administered. The administration is usually carried out once every two to six weeks and two to ten times in total. Examples of the applicable warm-blooded animals are monkeys, rabbits, dogs, guinea pigs, mice, rats, sheep, goats and chickens and the use of mice and rats is preferred.

In the preparation of the cells which produce monoclonal antibodies, an animal wherein the antibody titer is noted is selected from warm-blooded animals (e.g. mice) immunized with antigens, then spleen or lymph node is collected after two to five days from the final immunization and antibody-producing cells contained therein are fused with myeloma cells to give monoclonal antibody-producing hybridomas. Measurement of the antibody titer in antisera may, for example, be carried out by reacting a labeled

G protein coupled receptor protein (which will be mentioned later) with the antiserum followed by measuring the binding activity of the labeling agent with the antibody.

The operation for fusing may be carried out, for example, by a method of Kohler and Milstein (Nature, 256, 495, 1975). Examples of the fusion accelerator are polyethylene glycol (PEG), Sendai virus, etc. and the use of PEG is preferred.

Examples of the myeloma cells are NS-1, P3U1, SP2/0 AP-1, etc. and the use of P3U1 is preferred. The preferred fusion ratio of the numbers of antibody-producing cells used (spleen cells) to the numbers of myeloma cells is within a range of about 1:1 to 20:1. When PEG (preferably, PEG 1000 to PEG 6000) is added in a concentration of about 10-80% followed by incubating at 20-40°C (preferably, at 30-37°C) for one to ten minutes, an efficient cell fusion can be carried out.

Various methods may be applied for screening a hybridoma which produces anti-G protein coupled receptor antibody. For example, a supernatant liquid of hybridoma culture is added to a solid phase (e.g. microplate) to which the G protein coupled receptor protein antigen is adsorbed either directly or with a carrier, then anti-immunoglobulin antibody (anti-mouse immunoglobulin antibody is used when the cells used for the cell fusion are those of mouse) which is labeled with a radioactive substance, an enzyme or the like, or protein A is added thereto and then anti-G protein coupled receptor monoclonal antibodies bound on the solid phase are detected; or a supernatant liquid of the hybridoma culture is added to the solid phase to which anti-immunoglobulin or protein A is adsorbed, then the G protein coupled receptor labeled with a radioactive substance or an enzyme is added and anti-G protein coupled receptor monoclonal antibodies bonded with the solid phase is detected.

Selection and cloning of the anti-G protein coupled receptor monoclonal antibody-producing hybridoma may be carried out by methods per se known to those of skill in the art or methods similar thereto. Usually, it is carried out in a

- 1 7 1 -

medium for animal cells, containing HAT (hypoxanthine, aminopterin and thymidine). With respect to a medium for the selection, for the cloning and for the growth, any medium may be used so far as hybridoma is able to grow therein. Examples of the medium are an RPMI 1640 medium (Dainippon Pharmaceutical Co., Ltd., Japan) containing 1-20% (preferably 10-20%) of fetal calf serum (FCS), a GIT medium (Wako Pure Chemical, Japan) containing 1-20% of fetal calf serum and a serum-free medium for hybridoma culturing (SFM-101; Nissui Seiyaku, Japan). The culturing temperature is usually 20-40°C and, preferably, about 37°C. The culturing time is usually from five days to three weeks and, preferably, one to two weeks. The culturing is usually carried out in 5% carbon dioxide gas. The antibody titer of the supernatant liquid of the hybridoma culture may be measured by the same manner as in the above-mentioned measurement of the antibody titer of the anti-G protein coupled receptor in the antiserum.

The cloning can be usually carried out by methods known per se such as techniques in semi-solid agar and limiting dilution. The cloned hybridoma is preferably cultured in modern serum-free culture media to obtain optimal amounts of antibody in supernatants. The target monoclonal antibody is also preferably obtained from ascitic fluid derived from a mouse, etc. injected intraperitoneally with live hybridoma cells.

(b) Purification of the Monoclonal Antibody.

Like in the separation/purification of conventional polyclonal antibodies, the separation/purification of the anti-G protein coupled receptor monoclonal antibody may be carried out by methods for separating/purifying immunoglobulin (such as salting-out, precipitation with an alcohol, isoelectric precipitation, electrophoresis, adsorption/adsorption using ion exchangers such as DEAE, ultracentrifugation, gel filtration, specific purifying methods in which only an antibody is collected by treatment with an active adsorbent (such as an antigen-binding solid

- 1 7 2 -

phase, protein A or protein G) and the bond is dissociated whereupon the antibody is obtained.

The G protein coupled receptor antibody of the present invention which is manufactured by the aforementioned method (a) or (b) is capable of specifically recognizing G protein coupled receptors and, accordingly, it can be used for a quantitative determination of the G protein coupled receptor in test liquid samples and particularly for a quantitative determination by sandwich immunoassays.

Thus, the present invention provides, for example, the following methods:

- (i) a quantitative determination of a G protein coupled receptor in a test liquid sample, which comprises
 - (a) competitively reacting the test liquid sample and a labeled G protein coupled receptor with an antibody which reacts with the G protein coupled receptor of the present invention, and
 - (b) measuring the ratio of the labeled G protein coupled receptor binding with said antibody; and
 - (ii) a quantitative determination of a G protein coupled receptor in a test liquid sample, which comprises
 - (a) reacting the test liquid sample with an antibody immobilized on an insoluble carrier and a labeled antibody simultaneously or continuously, and
 - (b) measuring the activity of the labeling agent on the insoluble carrier
- wherein one antibody is capable of recognizing the N-terminal region of the G protein coupled receptor while another antibody is capable of recognizing the C-terminal region of the G protein coupled receptor.

When the monoclonal antibody of the present invention recognizing a G protein coupled receptor (hereinafter, may be referred to as "anti-G protein coupled receptor antibody") is used, G protein coupled receptors can be measured and, moreover, can be detected by means of a tissue staining, etc. as well. For such an object, antibody molecules per se may be used or F(ab')₂, Fab' or Fab fractions of the antibody

molecule may be used too. There is no particular limitation for the measuring method using the antibody of the present invention and any measuring method may be used so far as it relates to a method in which the amount of antibody, antigen or antibody-antigen complex, depending on or corresponding to the amount of antigen (e.g. the amount of G protein coupled receptor, etc.) in the liquid sample to be measured, is detected by a chemical or a physical means and then calculated using a standard curve prepared by a standard solution containing the known amount of antigen. For example, nephrometry, competitive method, immunometric method and sandwich method are suitably used and, in terms of sensitivity and specificity, the sandwich method which will be described herein later is particularly preferred.

Examples of the labeling agent used in the measuring method using the labeling substance are radioisotopes, enzymes, fluorescent substances, luminescent substances, colloids, magnetic substances, etc. Examples of the radioisotope are [125 I], [131 I], [3 H] and [14 C]; preferred examples of the enzyme are those which are stable and with big specific activity, such as β -galactosidase, β -glucosidase, alkali phosphatase, peroxidase and malate dehydrogenase; examples of the fluorescent substance are fluorescamine, fluorescein isothiocyanate, etc.; and examples of the luminescent substance are luminol, luminol derivatives, luciferin, lucigenin, etc. Further, a biotin-avidin system may also be used for binding an antibody or antigen with a labeling agent.

In an insolubilization (immobilization) of antigens or antibodies, a physical adsorption may be used or a chemical binding which is usually used for insolubilization or immobilization of proteins or enzymes may be used as well. Examples of the carrier are insoluble polysaccharides such as agarose, dextran and cellulose; synthetic resins such as polystyrene, polyacrylamide and silicone; glass; etc.

In a sandwich (or two-site) method, the test liquid is made to react with an insolubilized anti-G protein coupled

receptor antibody (the first reaction), then it is made to react with a labeled anti-G protein coupled receptor antibody (the second reaction) and the activity of the labeling agent on the insoluble carrier is measured whereupon the amount of the G protein coupled receptor in the test liquid can be determined. The first reaction and the second reaction may be conducted reversely or simultaneously or they may be conducted with an interval. The type of the labeling agent and the method of insolubilization (immobilization) may be the same as those mentioned already herein. In the immunoassay by means of a sandwich method, it is not always necessary that the antibody used for the labeled antibody and the antibody for the solid phase is one type or one species but, with an object of improving the measuring sensitivity, etc., a mixture of two or more antibodies may be used too.

In the method of measuring G protein coupled receptors by the sandwich method of the present invention, the preferred anti-G protein coupled receptor antibodies used for the first and the second reactions are antibodies wherein their sites binding to the G protein coupled receptors are different each other. Thus, the antibodies used in the first and the second reactions are those wherein, when the antibody used in the second reaction recognizes the C-terminal region of the G protein coupled receptor, then the antibody recognizing the site other than C-terminal regions, e.g. recognizing the N-terminal region, is preferably used in the first reaction.

The anti-G protein coupled receptor antibody of the present invention may be used in a measuring system other than the sandwich method such as a competitive method, an immunometric method and a nephrometry. In a competitive method, an antigen in the test solution and a labeled antigen are made to react with an antibody in a competitive manner, then an unreacted labeled antigen (F) and a labeled antigen binding with an antibody (B) are separated (i.e. B/F separation) and the labeled amount of any of B and F is measured whereupon the amount of the antigen in the test

solution is determined. With respect to a method for such a reaction, there are a liquid phase method in which a soluble antibody is used as the antibody and the B/F separation is conducted by polyethylene glycol, a second antibody to the above-mentioned antibody, etc.; and a solid phase method in which an immobilized antibody is used as the first antibody or a soluble antibody is used as the first antibody while an immobilized antibody is used as the second antibody.

In an immunometric method, an antigen in the test solution and an immobilized antigen are subjected to a competitive reaction with a certain amount of a labeled antibody followed by separating into solid and liquid phases; or the antigen in the test solution and an excess amount of labeled antibody are made to react, then a immobilized antigen is added to bind an unreacted labeled antibody with the solid phase and separated into solid and liquid phases. After that, the labeled amount of any of the phases is measured to determine the antigen amount in the test solution.

In a nephrometry, the amount of insoluble sediment which is produced as a result of the antigen-antibody reaction in a gel or in a solution is measured. Even when the antigen amount in the test solution is small and only a small amount of the sediment is obtained, a laser nephrometry wherein scattering of laser is utilized can be suitably used.

In applying each of those immunological measuring methods (immunoassays) to the measuring method of the present invention, it is not necessary to set up any special condition, operation, etc. therefor. A measuring system (assay system) for G protein coupled receptor may be constructed taking the technical consideration of the persons skilled in the art into consideration in the conventional conditions and operations for each of the methods. With details of those conventional technical means, a variety of reviews, reference books, etc. may be referred to. They are, for example, Hiroshi Irie (ed): "Radioimmunoassay" (Kodansha, Japan, 1974); Hiroshi Irie (ed): "Radioimmunoassay; Second Series" (Kodansha, Japan, 1979); Eiji Ishikawa et al. (ed): "Enzyme Immunoassay" (Igaku Shoin,

Japan, 1978); Eiji Ishikawa et al. (ed): "Enzyme Immunoassay" (Second Edition) (Igaku Shoin, Japan, 1982); Eiji Ishikawa et al. (ed): "Enzyme Immunoassay" (Third Edition) (Igaku Shoin, Japan, 1987); "Methods in Enzymology" Vol. 70 (Immunochemical Techniques (Part A)); *ibid.* Vol. 73 (Immunochemical Techniques (Part B)); *ibid.* Vol. 74 (Immunochemical Techniques (Part C)); *ibid.* Vol. 84 (Immunochemical Techniques (Part D: Selected Immunoassays)); *ibid.* Vol. 92 (Immunochemical Techniques (Part E: Monoclonal Antibodies and General Immunoassay Methods)); *ibid.* Vol. 121 (Immunochemical Techniques (Part I: Hybridoma Technology and Monoclonal Antibodies)) (Academic Press); etc.

(7) Preparation of Animals Having the G Protein Coupled

Receptor Protein-Encoding DNA of the Present Invention.

It is possible to prepare transgenic animals expressing G protein coupled receptors using G protein coupled receptor protein-encoding DNA. Examples of the animals are warm-blooded mammals such as rats, rabbit, sheep, swines, cattle, cats, dogs and monkeys.

In transferring the G protein coupled receptor protein-encoding DNA to the aimed animal, it is generally advantageous that said DNA is used by ligating with a site at the downstream of a promoter which is capable of expressing in animal cells. For example, when G protein coupled receptor protein DNA is to be transferred to a rabbit, a gene construct ligated with a site at the downstream of various promoters which are capable of expressing the G protein coupled receptor protein DNA derived from an animal compatible to the animal in animal host cells is subjected to a microinjection to the fertilized ovum (oosperm) of the aimed animal (e.g. fertilized ovum (embryo) of rabbit) whereupon the transgenic animal which produces the G protein coupled receptor protein in a high amount can be prepared.

Examples of the promoters used are promoters derived from virus and ubiquitous expression promoters such as metallothionein promoters may be used but, preferably,

enolase gene promoters and NGF gene promoters capable of specifically expressing in brain are used.

Transfer of the G protein coupled receptor protein DNA at a fertilized ovum cell stage is secured in order that the DNA can be present in all of embryonal cells and body somatic cells of an aimed animal. The fact that the G protein coupled receptor protein is present in the fertilized ovum cells of the produced transgenic animal after the DNA transfer means that all progeny of the produced transgenic animal have the G protein coupled receptor protein in all of their embryonal cells and somatic cells. Descendants (offsprings) of the animal of this type which inherited the gene have the G protein coupled receptor protein in all of their embryonal cells and somatic cells.

The transgenic animal to which the G protein coupled receptor protein DNA is transferred can be subjected to a mating and a breeding for generations under a common breeding circumstance as the animal holding said DNA after confirming that the gene can be stably retained. Moreover, male and female animals having the desired DNA are mated to give a homozygote having the transduced gene in both homologous chromosomes and then those male and female animals are mated whereby it is possible to breed for generations so that all descendants have said DNA.

The animal to which the G protein coupled receptor protein DNA is transferred highly expresses the G protein coupled receptor protein and, accordingly, it is useful as the animal for screening for an agonist or an antagonist to said G protein coupled receptor protein.

The DNA-transferred animal can be used as a cell source for a tissue culture. For example, DNA or RNA in the tissue of the DNA-transferred mouse is directly analyzed or protein tissues expressed by gene are analyzed whereupon the G protein coupled receptor protein can be analyzed.

Cells of the G protein coupled receptor protein-containing tissue are cultured by standard tissue culture techniques whereupon it is possible to study the function of the cells

which are usually difficult to culture (e.g. those derived from brain and peripheral tissues) using the resulting culture. By using said cells, it is also possible to select the pharmaceuticals which can potentiate, for example, the functions of various tissues. Moreover, if a cell strain with a high expression is available, it is possible to separate and purify G protein coupled receptor proteins therefrom.

As such, the amount of G protein coupled receptor proteins can now be determined with a high precision using the anti-G protein coupled receptor antibody of the present invention.

(8) Antisense Oligonucleotides Capable of Inhibiting Replication of G Protein Coupled Receptor Protein Gene

In another aspect of the present invention, antisense oligonucleotides (nucleic acids) capable of inhibiting the replication or expression of G protein coupled receptor protein gene may be designed and synthesized based on information on the nucleotide sequences of cloned and determined G protein coupled receptor protein-encoding DNAs. Such an antisense oligonucleotide (nucleic acid) is capable of hybridizing with RNA of G protein coupled receptor protein genes to inhibit the synthesis or function of said RNA or of modulating the expression of a G protein coupled receptor protein gene via the interaction with G protein coupled receptor protein-related RNA. Oligonucleotides complementary to, and specifically hybridizable with, selected sequences of G protein coupled receptor protein-related RNA are useful in controlling or modulating the expression of a G protein coupled receptor protein gene *in vitro* and *in vivo*, and in treating or diagnosing disease states of suspected animals. The term "corresponding" means homologous to or complementary to a particular sequence of the nucleotide sequence or nucleic acid including the gene. As between nucleotides (nucleic acids) and peptides (proteins), "corresponding" usually refers to amino acids of a peptide (protein) in an order derived from the sequence of a nucleotides (nucleic acids) or its complement.

The G protein coupled receptor protein gene 5' end hairpin loop, 5' end 6-base-pair repeats, 5' end untranslated region, polypeptide translation initiation codon, protein coding region, ORF translation initiation codon, 3' untranslated region, 3' end palindromic region, and 3' end hairpin loop may be selected as preferred targets though any region may be a target among G protein coupled receptor protein genes. The relationship between the target and oligonucleotides complementary to at least a portion of the target, specifically hybridizable with the target, is denoted as "antisense".

The antisense oligonucleotides may be polydeoxynucleotides containing 2-deoxy-D-ribose, polyribonucleotides containing D-ribose, any other type of polynucleotide which is an N-glycoside of a purine or pyrimidine base, or other polymers containing nonnucleotide backbones (e.g., protein nucleic acids and synthetic sequence-specific nucleic acid polymers commercially available) or nonstandard linkages, providing that the polymers contain nucleotides in a configuration which allows for base pairing and base stacking such as is found in DNA and RNA. They may include double- and single-stranded DNA, as well as double- and single-stranded RNA and DNA:RNA hybrids, and also include, as well as unmodified forms of the polynucleotide or oligonucleotide, known types of modifications, for example, labels which are known to those skilled in the art, "caps", methylation, substitution of one or more of the naturally occurring nucleotides with analogue, internucleotide modifications such as, for example, those with uncharged linkages (e.g., methyl phosphonates, phosphotriesters, phosphoramidates, carbamates, etc.) and with charged linkages or sulfur-containing linkages (e.g., phosphorothioates, phosphorodithioates, etc.), those containing pendant moieties, such as, for example, proteins (including nucleases, nuclease inhibitors, toxins, antibodies, signal peptides, poly-L-lysine, etc.) and saccharides (e.g., monosaccharides, etc.), those with intercalators (e.g., acridine, psoralen, etc.), those containing chelators (e.g., metals, radioactive metals, boron, oxidative metals, etc.),

those containing alkylators, those with modified linkages (e.g., alpha anomeric nucleic acids, etc.). The terms "nucleoside", "nucleotide" and "nucleic acid" will include those moieties which contain not only the known purine and pyrimidine bases, but also other heterocyclic bases which have been modified. Such modifications include methylated purines and pyrimidines, acylated purines and pyrimidines, or other heterocycles. Modified nucleosides or nucleotides will also include modifications on the sugar moiety, e.g., wherein one or more of the hydroxyl groups are replaced with halogen, aliphatic groups, or are functionalized as ethers, amines, or the like.

The antisense nucleic acid of the present invention is RNA, DNA or a modified nucleic acid. Examples of modified nucleic acid are, but not limited to, degradation-resistant sulfurized and thiophosphate derivatives of nucleic acids, and poly- or oligonucleoside amides. Preferred design modifications of the antisense nucleic acids of the present invention are modifications that are designed to:

- (1) increase the intracellular stability of the nucleic acid;
 - (2) increase the cellular permeability of the nucleic acid;
 - (3) increase the affinity of the nucleic acid for the target sense strand; or
 - (4) decrease the toxicity (if any) of the nucleic acid.
- Many such modifications are known to those skilled in the art, as described in J. Kawakami et al., Pharm Tech Japan, Vol. 8, pp.247, 1992; Vol. 8, pp.395, 1992; S. T. Crooke et al., ed., Antisense Research and Applications, CRC Press, 1993; etc. The nucleic acids may contain altered or modified sugars, bases or linkages, be delivered in specialized systems such as liposomes, microspheres or by gene therapy, or may have attached moieties. Such attached moieties include polycationic moieties such as polylysine that act as charge neutralizers of the phosphate backbone, or hydrophobic moieties such as lipids (e.g., phospholipids, cholesterol, etc.) that enhance interaction with cell membranes or increase uptake of the nucleic acid. Preferred lipids that may be attached are

cholesterols or derivatives thereof (e.g., cholesterol chloroformate, cholic acid, etc.). The moieties may be attached at the 3' or 5' ends of the nucleic acids, and also may be attached through a base, sugar, or internucleoside linkage. Other moieties may be capping groups specifically placed at the 3' or 5' ends of the nucleic acids to prevent degradation by nuclease such as exonuclease, RNase, etc. Such capping groups include, but are not limited to, hydroxyl protecting groups known to those skilled in the art, including glycols such as polyethylene glycols, tetraethylene glycol and the like.

The inhibitory activity of antisense nucleic acids can be examined using the transformant (or transfectant) of the present invention, the in vitro and in vivo gene expression system of the present invention, or the in vitro and in vivo translation system of G protein coupled receptor proteins. The nucleic acid can be placed in the cell through any number of ways known per se.

In the specification and drawings of the present application, the abbreviations used for bases (nucleotides), amino acids and so forth are those recommended by the IUPAC-IUB Commission on Biochemical Nomenclature or those conventionally used in the art. Examples thereof are given below.

Amino acids for which optical isomerism is possible are, unless otherwise specified, in the L form.

DNA : Deoxyribonucleic acid
 cDNA: Complementary deoxyribonucleic acid

A : Adenine

T : Thymine

G : Guanine

C : Cytosine

RNA : Ribonucleic acid

mRNA : Messenger ribonucleic acid

dATP: Deoxyadenosine triphosphate

dTTP: Deoxythymidine triphosphate

dGTP: Deoxyguanosine triphosphate
 dCTP: Deoxycytidine triphosphate
 ATP : Adenosine triphosphate
 EDTA: Ethylenediamine tetraacetic acid
 SDS : Sodium dodecyl sulfate
 EIA: Enzyme Immunoassay

G, Gly: Glycine (or Glycyl)
 A, Ala: Alanine (or Alanyl)
 V, Val: Valine (or Valyl)
 L, Leu: Leucine (or Leucyl)
 I, Ile: Isoleucine (or Isoleucyl)
 S, Ser: Serine (or Seryl)
 T, Thr: Threonine (or Threonyl)
 C, Cys: Cysteine (or Cysteinyl)
 M, Met: Methionine (or Methionyl)
 E, Glu: Glutamic acid (or Glutamyl)
 D, Asp: Aspartic acid (or Aspartyl)
 K, Lys: Lysine (or Lysyl)
 R, Arg: Arginine (or Arginyl)
 H, His: Histidine (or Histidyl)
 F, Phe: Phenylalanine (or Phenylalanyl)
 Y, Tyr: Tyrosine (or Tyrosyl)
 W, Trp: Tryptophan (or Tryptophanyl)
 P, Pro: Proline (or Prolyl)
 N, Asn: Asparagine (or Asparaginyl)
 Q, Gln: Glutamine (or Glutaminyl)
 NVal: Norvaline (or Norvalyl)
 pGlu: Pyroglutamic acid (or Pyroglutamyl)
 Bzl: γ -Butyrolactone- γ -carbonyl
 Kpc: 2-Ketopiperidiny-6-carbonyl
 Otc: 3-Oxopiperhydro-1,4-thiazin-5-carbonyl
 Me: Methyl
 Et: Ethyl
 Bu: Butyl
 Ph: Phenyl
 TC: Thiazolidiny-4(R)-carboxamide

The transformant *Escherichia coli*, designated INVa F'/p19P2, which is obtained in the Example 3 mentioned herein below, is on deposit under the terms of the Budapest Treaty from August 9, 1994, with the National Institute of Bioscience and Human-Technology (NIBH), Agency of Industrial Science and Technology, Ministry of International Trade and Industry, Japan and has been assigned the Accession Number FERM BP-4776. It is also on deposit from August 22, 1994 with the Institute for Fermentation, Osaka, Japan (IFO) and has been assigned the Accession Number IFO 15739.

The transformant *Escherichia coli*, designated INVa F'/pG3-2, which is obtained in the Example 4 mentioned herein below, is on deposit under the terms of the Budapest Treaty from August 9, 1994, with NIBH and has been assigned the Accession Number FERM BP-4775. It is also on deposit from August 22, 1994 with IFO and has been assigned the Accession Number IFO 15740.

The transformant *Escherichia coli*, designated INVa F'/p63A2, which is obtained in the Example 5 mentioned herein below, is on deposit under the terms of the Budapest Treaty from August 9, 1994, with NIBH and has been assigned the Accession Number FERM BP-4777. It is also on deposit from August 22, 1994 with IFO and has been assigned the Accession Number IFO 15738.

The transformant *Escherichia coli*, designated JM109/phGR3, which is obtained in the Example 6 mentioned herein below, is on deposit under the terms of the Budapest Treaty from September 27, 1994, with NIBH and has been assigned the Accession Number FERM BP-4807. It is also on deposit from September 22, 1994 with IFO and has been assigned the Accession Number IFO 15748.

The transformant *Escherichia coli*, designated JM109/p3H2-17, which is obtained in the Example 7 mentioned herein below, is on deposit under the terms of the Budapest Treaty from September 27, 1994, with NIBH and has been assigned the Accession Number FERM BP-4806. It is also on deposit from September 22, 1994 with IFO and has been assigned the Accession

Number IFO 15747.

The transformant *Escherichia coli*, designated JM109/p3H2-34, which is obtained in the Example 8 mentioned herein below, is on deposit under the terms of the Budapest Treaty from October 12, 1994, with NIBH and has been assigned the Accession Number FERM BP-4828. It is also on deposit from October 12, 1994 with IFO and has been assigned the Accession Number IFO 15749.

The transformant *Escherichia coli*, designated JM109/pMD4, which is obtained in the Example 9 mentioned herein below, is on deposit under the terms of the Budapest Treaty from November 11, 1994, with NIBH and has been assigned the Accession Number FERM BP-4888. It is also on deposit from November 17, 1994 with IFO and has been assigned the Accession Number IFO 15765.

The transformant *Escherichia coli*, designated JM109/pMGR20, which is obtained in the Example 10 mentioned herein below, is on deposit under the terms of the Budapest Treaty from December 15, 1994, with NIBH and has been assigned the Accession Number FERM BP-4937. It is also on deposit from December 14, 1994 with IFO and has been assigned the Accession Number IFO 15773.

The transformant *Escherichia coli*, designated JM109/pMJ10, which is obtained in the Example 12 mentioned herein below, is on deposit under the terms of the Budapest Treaty from December 15, 1994, with NIBH and has been assigned the Accession Number FERM BP-4936. It is also on deposit from December 16, 1994 with IFO and has been assigned the Accession Number IFO 15784.

The transformant *Escherichia coli*, designated JM109/pMH28, which is obtained in the Example 14 mentioned herein below, is on deposit under the terms of the Budapest Treaty from January 13, 1995, with NIBH and has been assigned the Accession Number FERM BP-4970. It is also on deposit from January 20, 1995 with IFO and has been assigned the Accession Number IFO 15791.

The transformant *Escherichia coli*, designated

JM109/pMN7, which is obtained in the Example 16 mentioned herein below, is on deposit under the terms of the Budapest Treaty from February 22, 1995, with NIBH and has been assigned the Accession Number FERM BP-5011. It is also on deposit from February 27, 1995 with IFO and has been assigned the Accession Number IFO 15803.

The transformant *Escherichia coli*, designated

JM109/pS38, which is obtained in the Example 17 mentioned herein below, is on deposit under the terms of the Budapest Treaty from October 27, 1994, with NIBH and has been assigned the Accession Number FERM BP-4856. It is also on deposit from October 25, 1994 with IFO and has been assigned the Accession Number IFO 15754.

The transformant *Escherichia coli*, designated

JM109/pMAH2-17, which is obtained in the Example 19 mentioned herein below, is on deposit under the terms of the Budapest Treaty from April 7, 1995, with NIBH and has been assigned the Accession Number FERM BP-5073. It is also on deposit from March 31, 1995 with IFO and has been assigned the Accession Number IFO 15813.

The transformant *Escherichia coli*, designated

JM109/pMN128, which is obtained in the Example 20 mentioned herein below, is on deposit under the terms of the Budapest Treaty from March 17, 1995, with NIBH and has been assigned the Accession Number FERM BP-5039. It is also on deposit from March 22, 1995 with IFO and has been assigned the Accession Number IFO 15810.

The transformant *Escherichia coli*, designated

JM109/pAH2-17, which is obtained in the Example 21 mentioned herein below, is on deposit under the terms of the Budapest Treaty from July 20, 1995, with NIBH and has been assigned the Accession Number FERM BP-5168. It is also on deposit from July 14, 1995 with IFO and has been assigned the Accession Number IFO 15856.

Each SEQ ID NO set forth in the SEQUENCE LISTING of the specification refers to the following sequence:

[SEQ ID NO: 24] is a partial amino acid sequence of the human pituitary gland-derived G protein coupled receptor protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2,

[SEQ ID NO: 25] is a partial amino acid sequence of the human pituitary gland-derived G protein coupled receptor protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2,

[SEQ ID NO: 26] is an entire amino acid sequence of the human pituitary gland-derived G protein coupled receptor protein encoded by the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2,

[SEQ ID NO: 27] is a partial amino acid sequence of the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein encoded by the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein cDNA fragment having a nucleotide sequence (SEQ ID NO: 32), derived based upon the nucleotide sequences of the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein cDNA fragments each included in pG3-2 and pG1-10,

[SEQ ID NO: 28] is a partial amino acid sequence of the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein encoded by pS38,

[SEQ ID NO: 29] is a nucleotide sequence of the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2,

[SEQ ID NO: 30] is a nucleotide sequence of the human pituitary gland-derived G protein coupled receptor protein cDNA fragment included in p19P2,

[SEQ ID NO: 31] is an entire nucleotide sequence of the human pituitary gland-derived G protein coupled receptor protein cDNA included in p19P2,

[SEQ ID NO: 32] is a nucleotide sequence of the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein cDNA, derived based upon the nucleotide sequences of the mouse pancreatic β -cell line, MIN6-derived G protein coupled receptor protein cDNA fragments each

included in pG3-2 and pG1-10,
[SEQ ID NO: 33] is a nucleotide sequence of the mouse
pancreatic β -cell line, MIN6-derived G protein cDNA included
in p5638,

5 [SEQ ID NO: 34] is a partial amino acid sequence of the human
amygdaloid nucleus-derived G protein coupled receptor protein
encoded by the cDNA fragment included in p63A2,

[SEQ ID NO: 35] is a partial amino acid sequence of the human
amygdaloid nucleus-derived G protein coupled receptor protein
encoded by the cDNA fragment included in p63A2,

[SEQ ID NO: 36] is a nucleotide sequence of the human
amygdaloid nucleus-derived G protein coupled receptor protein
cDNA fragment included in p63A2,

15 [SEQ ID NO: 37] is a nucleotide sequence of the human
amygdaloid nucleus-derived G protein coupled receptor protein
cDNA fragment included in p63A2,

[SEQ ID NO: 38] is a partial amino acid sequence encoded
by the mouse pancreatic β -cell line, MIN6-derived G protein
coupled receptor protein cDNA included in p3H2-17,

20 [SEQ ID NO: 39] is a full-length amino acid sequence encoded
by the open reading frame of the mouse pancreatic β -cell line,
MIN6-derived G protein coupled receptor protein cDNA included
in pMAH2-17,

[SEQ ID NO: 40] is a nucleotide sequence of the mouse
pancreatic β -cell line, MIN6-derived G protein coupled
receptor protein cDNA included in p3H2-17,

[SEQ ID NO: 41] is a nucleotide sequence of the mouse
pancreatic β -cell line, MIN6-derived G protein coupled
receptor protein cDNA included in pMAH2-17,

30 [SEQ ID NO: 42] is a partial amino acid sequence encoded
by the mouse pancreatic β -cell line, MIN6-derived G protein
coupled receptor protein cDNA included in p3H2-34,

[SEQ ID NO: 43] is a nucleotide sequence of the mouse
pancreatic β -cell line, MIN6-derived G protein coupled
receptor protein cDNA fragment included in p3H2-34,

35 [SEQ ID NO: 44] is a partial amino acid sequence encoded
by the rabbit gastropyrolic part smooth muscle-derived G

protein coupled receptor protein cDNA included in pMD4,
[SEQ ID NO: 45] is a nucleotide sequence of the rabbit
gastropyrolic part smooth muscle-derived G protein coupled
receptor protein cDNA fragment included in pMD4,

5 [SEQ ID NO: 46] is an entire amino acid sequence
encoded by the mouse pancreatic β -cell line, MIN6-derived G
protein coupled receptor protein cDNA included in pMGR20,

10 [SEQ ID NO: 47] is a nucleotide sequence of the mouse
pancreatic β -cell line, MIN6-derived G protein coupled
receptor protein cDNA included in pMGR20,

[SEQ ID NO: 48] is a partial amino acid sequence encoded
by the rabbit gastropyrolic part smooth muscle-derived G
protein coupled receptor protein cDNA included in pM10,

15 [SEQ ID NO: 49] is a nucleotide sequence of the rabbit
gastropyrolic part smooth muscle-derived G protein coupled
receptor protein cDNA fragment included in pM10,

[SEQ ID NO: 50] is a partial amino acid sequence encoded
by the rabbit gastropyrolic part smooth muscle-derived G
protein coupled receptor protein cDNA included in pMAH28,

20 [SEQ ID NO: 51] is a nucleotide sequence of the rabbit
gastropyrolic part smooth muscle-derived G protein coupled
receptor protein cDNA fragment included in pMAH28,

25 [SEQ ID NO: 52] is a partial amino acid sequence encoded
by the rabbit gastropyrolic part smooth muscle-derived G
protein coupled receptor protein cDNA included in pMN7,

[SEQ ID NO: 53] is a nucleotide sequence of the rabbit
gastropyrolic part smooth muscle-derived G protein coupled
receptor protein cDNA fragment included in pMN7,

30 [SEQ ID NO: 54] is a partial amino acid sequence encoded
by the rabbit gastropyrolic part smooth muscle-derived G
protein coupled receptor protein cDNA included in pMN128,

[SEQ ID NO: 55] is a nucleotide sequence of the rabbit
gastropyrolic part smooth muscle-derived G protein coupled
receptor protein cDNA fragment included in pMN128,

35 [SEQ ID NO: 56] is a full-length amino acid sequence of the
human-derived G protein coupled receptor protein encoded
by the human-derived G protein coupled receptor protein cDNA

included in PHA2-17, and

(SEQ ID NO: 57) is a nucleotide sequence of the human-derived G protein coupled receptor protein cDNA included in PHA2-17.

EXAMPLES

Described below are working examples of the present invention which are provided only for illustrative purposes, and not to limit the scope of the present invention. In light of the present disclosure, numerous embodiments within the scope of the claims will be apparent to those of ordinary skill in the art.

Example 1

Preparation of Synthetic DNA Primer for Amplifying DNA Coding for G Protein Coupled Receptor Protein

A comparison of deoxyribonucleotide sequences coding for the known amino acid sequences corresponding to or near the first membrane-spanning domain each of human-derived TRH receptor protein (HTRHR), human-derived RANTES receptor protein (L10918, HUMRANTES), human Burkitt's lymphoma-derived unknown ligand receptor protein (X68149, HSBRL1A), human-derived somatostatin receptor protein (L14856, HUMSOMAT), rat-derived μ -opioid receptor protein (U02083, RNU02083), rat-derived κ -opioid receptor protein (U00442, U00442), human-derived neuromedin B receptor protein (M73482, HUMNMBR), human-derived muscarinic acetylcholine receptor protein (X15266, HSHM4), rat-derived adrenaline α_1 B receptor protein (L08609, RATAADRE01), human-derived somatostatin 3 receptor protein (M96738, HUMSSTR3X), human-derived C_5 a receptor protein (HUMCSAAR), human-derived unknown ligand receptor protein (HUMRDC1A), human-derived unknown ligand receptor protein (M84605, HUMOPIODRE) and rat-derived adrenaline α_2 B receptor protein (M91466, RATA2BAR) was made. As a result, highly homologous regions or parts were found (Figure 1).

Further, a comparison of deoxynucleotide sequences coding for the known amino acid sequences corresponding to or near the sixth membrane-spanning domain each of mouse-derived

unknown ligand receptor protein (M80481, MUSGIR), human-derived bombesin receptor protein (L08893, HUMBOMB3S), human-derived adenosine A2 receptor protein (S46950, S46950), mouse-derived unknown ligand receptor protein (D21061, MUSGPCR), mouse-derived TRH receptor protein (S43387, S43387), rat-derived neuromedin K receptor protein (J05189, RATNEURA), rat-derived adenosine A1 receptor protein (M69045, RATALARA), human-derived neurokinin A receptor protein (M57414, HUMNEKAR), rat-derived adenosine A3 receptor protein (M94152, RATADENREC), human-derived somatostatin 1 receptor protein (M81829, HUMSR11A), human-derived neurokinin 3 receptor protein (S86390, S86371S4), rat-derived unknown ligand receptor protein (X61496, RNCGPCR), human-derived somatostatin 4 receptor protein (L07061, HUMSSTR4Z) and rat-derived GnRH receptor protein (M31670, RATGNRHA) was made. As a result, highly homologous regions or parts were found (Figure 2).

The aforementioned abbreviations in the parentheses are identifiers (reference numbers) which are indicated when GenBank/EMBL Data Bank is retrieved by using DNASIS Gene/Protein Sequencing Data Base (CD019, Hitachi Software Engineering, Japan) and are usually called "Accession Numbers" or "Entry Names". HTRHR is, however, the sequence as disclosed in Japanese Unexamined Patent Publication No. 286986/1993 (EPA 638645).

Specifically, it was planned to incorporate mixed bases relying upon the base regions that were in agreement with cDNAs coding for a large number of receptor proteins in order to enhance base agreement of sequences with as many receptor cDNAs as possible even in other regions. Based upon these sequences, the degenerate synthetic DNA having a nucleotide sequence represented by SEQ ID NO: 1 which is complementary to the homologous nucleotide sequence of Figure 1 and the degenerate synthetic DNA having a nucleotide sequence represented by SEQ ID NO: 2 which is complementary to the homologous nucleotide sequence of Figure 2 were produced. Nucleotide synthesis was carried out by a DNA synthesizer.

[Synthetic DNAs]

5'-CGTGG (G or C) C (A or C) T (G or C) (G or C) TGGGCAAC

(A, G, C or T) (C or T) CCTG-3'

(SEQ ID NO: 1)

5 5'-GT (A, G, C or T) G (A or T) (A or G) (A or G) GGCA

(A, G, C or T) CCAGCAGA (G or T) GGCAA-3'

(SEQ ID NO: 2)

The parentheses indicate the incorporation of a plurality of bases, leading to multiple oligonucleotides in the primer preparation. In other words, nucleotide residues in parentheses of the aforementioned DNAs were incorporated in the presence of a mixture of plural bases at the time of synthesis.

Example 2

Isolation of Human Somatostatin Receptor Protein-Encoding DNA,

Human D5 Dopamine Receptor Protein-Encoding DNA, and Rat

Somatostatin Receptor Protein-Encoding DNA

(1) Amplification of DNA by Polymerase Chain Reaction (PCR)

CDNAs (QuickClone, CLONTECH Laboratories, Inc.)

prepared from human brain amygdaloid nucleus, human pituitary gland and rat brain each in an amount of 1 ng as templates, the synthetic DNA primers prepared in Example 1 each in an amount of 1 μ M, 2.5 mM dNTPs (deoxyribonucleoside triphosphates), and 2.5 units of Taq DNA polymerase (Takara Shuzo Co., Japan) were mixed together with a buffer attached to the enzyme kit such that the total amount was 100 μ l. The polymerase chain reaction was carried out by using a Thermal Cycler manufactured by Perkin-Elmer Co. One cycle was set to include 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min.. Totally this one cycle was repeated 30 times to amplify DNAs.

Amplification of DNAs was confirmed by 1.2% agarose electrophoresis [Figure 17].

(2) Isolation of Amplified DNA and Analysis of DNA Sequence
By using a TA Cloning Kit (Invitrogen Co.), the DNA amplified by the PCR was inserted into a plasmid vector, PCRTM II.

The DNA was transfected into E. coli attached to the kit to form an amplified DNA library. Colonies formed by the transformants were selected under guidance based on the activity of β -galactosidase on X-gal (5-bromo-4-chloro-3-indolyl- β -D-galactoside)-added LB (Luria-Bertani) plates in order to separate only white colonies in which DNA fragments are inserted. They were cultured in an LB culture medium to which ampicillin was added and plasmid DNAs were prepared with an automatic plasmid extracting machine (Kurabo Co., Japan).

An aliquot of the DNA thus prepared was further digested with EcoRI to confirm DNA fragments that were inserted, and a DNA yield each of clones was compared with a marker. An aliquot of the plasmid DNA thus prepared was treated with RNase, extracted with phenol/chloroform, precipitated in ethanol, and the resulting product was then reacted for sequencing by using a Dyedexy terminator cycle sequencing kit (Applied Biosystems Co.).

Sequencing was carried out by using a 370A fluorescent automatic sequencer manufactured by Applied Biosystems Co. The nucleotide sequences obtained were analyzed by using DNASIS (Hitachi Software Engineering, Japan). The nucleotide sequences obtained are shown in Figures 18, 19, 20 and 21. From these Figures and the results of homology retrieval, it was learned that the DNAs obtained were DNAs encoding human somatostatin receptor protein [Figures 18 and 19], human D5 dopamine receptor protein [Figure 20] and rat somatostatin receptor protein [Figure 21] that can be classified each into a group of G protein coupled receptor proteins.

In Figure 18 as described herein, the nucleotide sequence of the DNA is in agreement with the nucleotide sequence encoding somatostatin receptor (HUMSOMAT) and the clone, A58, is a human somatostatin receptor cDNA. The underlined part represents the 5' side synthetic DNA primer

used for the PCR. Thus, even when parts of the nucleotide sequence are mismatched, amplification is effected to a sufficient degree by the PCR.

It will be understood from Figure 19 that the clone, A58 is in good agreement with the nucleotide sequence coding for the human somatostatin receptor (HUMSOMAT) even when the sequencing is carried out from the opposite side. The underlined part represents the 3' side synthetic DNA primer used for the PCR. In this figure, the nucleotide sequences are mismatched to some extent even in the portions other than the primer portion presumably due to base substitution at the time of PCR or due to partial deviation in the sequencing reaction. It can be confirmed via sequencing of chains complementary thereto as required.

In Figure 20 as described herein, the nucleotide sequence of the DNA is in good agreement with a nucleotide sequence coding for the human D5 dopamine receptor (HUMDRD5A) except the primer portion (underlined). It was learned that the clone, 57-A-2, is a human D5 dopamine receptor cDNA.

In Figure 21 as described herein, the DNA is in good agreement with a nucleotide sequence coding for the rat somatostatin receptor (RNU04738) except the primer portion (underlined). It was learned that the clone, B54, is a rat somatostatin receptor cDNA.

Example 3

Isolation of Human Pituitary Gland-Derived G Protein Coupled Receptor Protein-Encoding DNA

(1) Amplification of Receptor cDNA by PCR Using Human Pituitary Gland-Derived cDNA

By using human pituitary gland-derived cDNA (QuickClone, CLONTECH Laboratories, Inc.) as a template, PCR amplification using the DNA primers synthesized in Example 1 was carried out. The composition of the reaction solution consisted of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 1 μ M, 1 ng of the template cDNA, 0.25 mM dNTPs, 1 μ l of Tag DNA polymerase and

a buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 95 $^{\circ}$ C for 1 min., 55 $^{\circ}$ C for 1 min. and 72 $^{\circ}$ C for 1 min. was repeated 30 times by using a Thermal Cycler (Perkin-Elmer Co.). Prior to adding Tag DNA polymerase, the remaining reaction solution was mixed and was heated at 95 $^{\circ}$ C for 5 minutes and at 65 $^{\circ}$ C for 5 minutes. The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(2) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products were separated by using a 0.8% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned into the Plasmid vector, PCR II (TM represents registered trademark). The recombinant vectors were introduced into *E. coli* INVa F' competent cells (Invitrogen Co.) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothstick to obtain transformant *Escherichia coli* INVa F'/p19P2.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNA thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNA was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a Dye-deoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a

fluorescent automatic sequencer, and the data of the nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan). The underlined portions represent regions corresponding to the synthetic primers [Figures 22 and 23].

Homology retrieval was carried out based upon the determined nucleotide sequences [Figures 22 and 23].

As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid, p19P2, possessed by the transformant *Escherichia coli* INVα F'/p19P2. To further confirm this

fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequences were converted into amino acid sequences [Figures 22 and 23], and homology retrieval was carried out in view of hydrophobicity plotting [Figures 24 and 25] and at the amino acid sequence level to find homology relative to neuropeptide Y receptor proteins [Figure 26].

Example 4

Isolation of Mouse Pancreas-Derived G Protein Coupled Receptor Protein-Encoding DNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Mouse Pancreatic β-Cell Strain, MIN6 and Synthesis of cDNA

A total RNA was prepared from the mouse pancreatic β-cell strain, MIN6 (Jun-ichi Miyazaki et al., Endocrinology, Vol. 127, No. 1, p.126-132) according to the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J., 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μg of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to reaction with mouse Moloney leukemia virus (MMLV) reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μl of TE buffer (10 mM Tris-HCl at pH8.0, 1 mM EDTA at pH8.0).

(2) Amplification of Receptor cDNA by PCR Using MIN6-Derived cDNA and Sequencing

By using, as a template, 5 μl of cDNA prepared from the mouse pancreatic β-cell strain, MIN6 in the above step (1), PCR amplification using the DNA primers synthesized in Example 1 was carried out under the same conditions as in Example 3(2). The resulting PCR product was subcloned into the plasmid vector, PCRTM II, in the same manner as in Example 2 to obtain a plasmid, pg3-2. The plasmid pg3-2 was transfected into *E. coli* INVα F' to obtain transformed *Escherichia coli* INVα F'/pg3-2.

By using, as a template, 5 μl of the cDNA prepared from the mouse pancreatic β-cell strain, MIN6, PCR amplification using DNA primers as disclosed in Libert F. et al., "Science, 244:569-572, 1989", i.e., a degenerate synthetic primer represented by the following sequence:

5'-CTGTC (C or T) G (C or T) (G or C) AT (C or T) GCITT
(G or T) GA (C or T) (A or C) G (G or C) TAC-3'
(SEQ ID NO: 60)

wherein I is inosine; and
a degenerate synthetic primer represented by the following sequence:

5'-A (G or T) G (A or T) AG (A or T) AGGCACGACGACGAI
(G or C) (A or G) (C or T) GAA-3'
(SEQ ID NO: 61)

wherein I is inosine,
was carried out under the same conditions as in Working Example 1. The resulting PCR product was subcloned into the plasmid vector, PCRTM II, in the same manner as described in Example 3(2) to obtain a plasmid, pg1-10.

The reaction for determining the nucleotide sequence (sequencing) was carried out with a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNA was decoded with the fluorescent automatic sequencer (ABI Co.), and the data

of the nucleotide sequence obtained were analyzed with DNASIS (Hitachi System Engineering Co., Japan).

Figure 27 shows a mouse pancreatic β -cell strain MIN6-derived G protein coupled receptor protein-encoding DNA and an amino acid sequence encoded by the isolated DNA based upon the nucleotide sequences of plasmids pG3-2 and pG1-10 which are held by the transformant Escherichia coli INVa F'/pG3-2. The underlined portions represent regions corresponding to the synthetic primers.

Homology retrieval was carried out based upon the determined nucleotide sequence [Figure 27]. As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment obtained. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequence was converted into an amino acid sequence [Figure 27], hydrophobicity plotting was carried out to confirm the presence of six hydrophobic regions [Figure 28]. Upon comparing the amino acid sequence with that of p19P2 obtained in Example 3, furthermore, a high degree of homology was found as shown in [Figure 61]. As a result, it is strongly suggested that the G protein coupled receptor proteins encoded by pG3-2 and pG1-10 recognize the same ligand as the G protein coupled receptor protein encoded by p19P2 does while the animal species from which the receptor proteins encoded by pG3-2 and pG1-10 are derived is different from that from which the receptor protein encoded by p19P2 is.

Example 5

Isolation of Human Amygdaloid Nucleus-Derived G Protein Coupled Receptor Protein-Encoding DNA

(1) Amplification of Receptor cDNA by PCR using Human Amygdaloid Nucleus-Derived cDNA

By using an amplified human amygdala-derived cDNA (QuickClone, CLONTECH Laboratories, Inc.) as a template, PCR amplification using the DNA primers synthesized in Example 1 was carried out. The composition of the reaction solution

consisted of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of $1 \mu\text{M}$, 1 ng of the template cDNA, 0.25 mM dNTPs, $1 \mu\text{l}$ of Taq DNA polymerase and a buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be $100 \mu\text{l}$. The cycle for amplification including 95°C for 1 min. , 55°C for 1 min. and 72°C for 1 min. was repeated 30 times by using a Thermal Cycler (Perkin-Elmer Co.). Prior to adding Taq DNA polymerase, the remaining reaction solution was mixed and was heated at 95°C for 5 minutes and at 65°C for 5 minutes. The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(2) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products were separated by using a 0.8% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, pCRTM II. The recombinant vectors were introduced into E. coli INVa F' competent cells (Invitrogen Co.) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothstick to obtain transformant Escherichia coli INVa F'/pG3A2.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNA thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNA was further processed with RNase, extracted with phenol/chloroform, and

precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer, and the data of the nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan).

Homology retrieval was carried out based upon the determined nucleotide sequences (Figures 29 and 30).

As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid, p63A2 possessed by the transformant *Escherichia coli* INVα F' / p63A2. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequences were converted into amino acid sequences (Figures 29 and 30), and homology retrieval was carried out in view of hydrophobicity plotting (Figures 31 and 32) and at the amino acid sequence level to find homology relative to mouse GIR [Figure 33].

Example 6

20 Cloning of Human Pituitary Gland-Derived G Protein Coupled Receptor Protein cDNA

(1) Cloning of cDNA Comprising Whole Coding Regions for Receptor Protein from Human Pituitary Gland-Derived

cDNA Library

The DNA library constructed by Clontech Co. wherein λ gt11 phage vector is used (CLONTECH Laboratories, Inc.; CLH L1139b) was employed as a human pituitary gland-derived cDNA library. The human pituitary gland cDNA library (2×10^6 pfu (plaque forming units)) was mixed with *E. coli* Y1090 treated with magnesium sulfate, and incubated at 37°C for 15 minutes followed by addition of 0.5% agarose (Pharmacia Co.) LB. The *E. coli* was plated onto a 1.5% agar (Wako-Junyaku Co.) LB plate (containing 50 μg/ml of ampicillin). A nitrocellulose filter was placed on the plate on which plaques were formed and the plaque was transferred onto the filter. The filter was denatured with an alkali and

then heated at 80 °C for 3 hours to fix DNAs.

The filter was incubated overnight at 42 °C together with the probe mentioned herein below in a buffer containing 50% formamide, 5 x SSPE (20 x SSPE (pH 7.4) is 3 M NaCl, 0.2 M NaH₂PO₄·H₂O, 25 mM EDTA), 5 X Denhardt's solution (Nippon Gene, Japan), 0.1% SDS and 100 μg/ml of salmon sperm DNA for hybridization.

The probe used was obtained by cutting the DNA fragment inserted in the plasmid, p19P2, obtained in Working Example 3, with *Eco*RI, followed by recovery and labelling by incorporation of [³²P]dCTP (Dupont Co.) with a random prime DNA labelling kit (Amasham Co.).

It was washed with 2 x SSC (20 x SSC is 3 M NaCl, 0.3 M sodium citrate), 0.1% SDS at 55 °C for 1 hour and, then, subjected to an autoradiography at -80 °C to detect hybridized plaques.

In this screening, hybridization signals were recognized in three independent plaques. Each DNA was prepared from the three clones. The DNAs digested with *Eco*RI were subjected to an agarose electrophoresis and were analyzed by the southern blotting using the same probe as the one used in the screening. Hybridizing bands were identified at about 0.7kb, 0.8 kb and 2.0kb, respectively. Among them, the DNA fragment corresponding to the band at about 2.0kb (λ hGR3) was selected. The λ hGR3-derived *Eco*RI fragment with a hybridizable size was subcloned to the *Eco*RI site of the plasmid, pUC18, and *E. coli* JM109 was transformed with the plasmid to obtain transformant *E. coli* JM109/pHGR3. A restriction enzyme map of the plasmid, pHGR3, was prepared relying upon a restriction enzyme map deduced from the nucleotide sequence as shown in Example 3.

As a result, it was learned that it carried a full-length receptor protein-encoding DNA which was predicted from the receptor protein-encoding DNA as shown in Example 3.

(2) Sequencing of Human Pituitary Gland-Derived Receptor Protein cDNA.

Among the EcoRI fragments inserted in the plasmid, phGR3, obtained in the above step (1), the from EcoRI to NheI nucleotide sequence with about 1330bp that is considered to be a receptor protein-coding region was sequenced. Concretely speaking, by utilizing restriction enzyme sites that exist in the EcoRI fragments, unnecessary parts were removed or necessary fragments were subcloned in order to prepare template plasmids for analyzing the nucleotide sequence.

The reaction for determining the nucleotide sequence (sequencing) was carried out with a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNA was decoded with the fluorescent automatic sequencer (ABI Co.), and the data of the nucleotide sequence obtained were analyzed with DNASTIS (Hitachi System Engineering Co., Japan).

Figure 34 shows a nucleotide sequence of from immediate after the EcoRI site up to the NheI site encoded by phGR3. The nucleotide sequence of the human pituitary gland-derived receptor protein-encoding DNA corresponds to the nucleotide sequence of from 118th to 123rd nucleotides [Figure 34]. An amino acid sequence of the receptor protein that is encoded by the nucleotide sequence is shown in Figure 34. Figure 36 shows the results of hydrophobicity plotting based upon the amino acid sequence.

(3) Northern Hybridization with Human Pituitary Gland-Derived Receptor Protein-Encoding phGR3

Northern blotting was carried out in order to detect the expression of phGR3-encoded human pituitary gland-derived receptor proteins in the pituitary gland at a mRNA level. Human pituitary gland mRNA (2.5 μ g, Clontech Co.) was used as a template mRNA and the same as the probe used in Working Example 5 was used as a probe. Nylon membrane (Pall Biodyne, U.S.A.) was used as a filter for northern blotting and migration of the mRNA and adsorption (sucking) thereof with the blotting filter was carried out according to the method as

disclosed in Molecular Cloning, Cold Spring Harbor Laboratory Press, 1989.

The hybridization was effected by incubating the above-mentioned filter and probe in a buffer containing 50% formamide, 5 x SSPE, 5 x Denhardt's solution, 0.1% SDS and 100 μ g/ml of salmon sperm DNA overnight at 42 °C. The filter was washed with 0.1 x SSC, 0.1% SDS at 50 °C and, after drying with an air, was exposed to an X-ray film (XAR5, Kodak) for three days at -80 °C. The results were as shown in Figure 35 from which it is considered that the receptor gene encoded by phGR3 is expressed in the human pituitary gland.

Example 7

Cloning of Mouse Pancreatic β -Cell Strain, MIN6-Derived G Protein Coupled Receptor Protein cDNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Mouse Pancreatic β -Cell Strain, MIN6 and Synthesis of cDNA

A total RNA was prepared from the mouse pancreatic β -cell strain, MIN6 (Jun-ichi Miyazaki et al., Endocrinology, Vol. 127, No. 1, p.126-132) according to the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J., 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to reaction with MMLV reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE.

(2) Amplification of Receptor cDNA by PCR Using MIN6-Derived cDNA and Sequencing

By using, as a template, 5 μ l of cDNA prepared from the mouse pancreatic β -cell strain, MIN6 in the above step (1), PCR amplification using the DNA primers synthesized in Example 1 was carried out. A reaction solution was

composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Taq DNA polymerase and 10 μ l of 10x buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min. was repeated 30 times by using a Thermal Cycler (Perkin-Elmer Co.). Prior to adding Taq DNA polymerase, the remaining reaction solution was mixed and was heated at 95 °C for 5 minutes and at 65 °C for 5 minutes. The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted CDNA Region

The PCR products obtained in the above step (2) were separated by using a 0.8% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, PCRTM II. The recombinant vectors were introduced into *E. coli* JM109 competent cells (Takara Shuzo Co., Japan) to produce transformants. Then, transformant clones having a CDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG (Isopropylthio- β -D-galactoside) and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothpick to obtain transformant *Escherichia coli* JM109/p3H2-17.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNAs thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNAs was

further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer, and the data of the nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan).

Homology retrieval was carried out based upon the determined nucleotide sequence [Figure 37]. As a result, it was learned that a novel G protein coupled receptor protein was encoded by the CDNA fragment insert in the plasmid possessed by the transformant *Escherichia coli* JM109/p3H2-17. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequence were converted into an amino acid sequence [Figure 37], and homology retrieval was carried out in view of hydrophobicity plotting [Figure 38] and at the amino acid sequence level to find homology relative to chicken ATP receptor (P34996), human somatostatin receptor subtype 3 (A46226), human somatostatin receptor 4 (JN0605) and bovine neuropeptide Y receptor (S26787) [Figure 39]. Abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are usually called "Accession Numbers".

Example 8

Cloning of Mouse Pancreatic β -Cell Strain, MIN6-Derived G Protein Coupled Receptor Protein cDNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Mouse Pancreatic β -Cell Strain, MIN6 and Synthesis of cDNA

A total RNA was prepared from the mouse pancreatic β -cell strain, MIN6 (Jun-ichi Miyazaki et al., Endocrinology, Vol. 127, No. 1, p.126-132) according to the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J., 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected

to reaction with MMLV reverse transcriptase (BRL Co.)

in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE.

(2) Amplification of Receptor cDNA by PCR Using MIN6-Derived cDNA and Sequencing

By using, as a template, 5 μ l of cDNA prepared from the mouse pancreatic β -cell strain, MIN6, in the above step (1), PCR amplification using the DNA primers synthesized in Example 1 was carried out. A reaction solution was composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Taq DNA polymerase and 10 μ l of 10 \times buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 $^{\circ}$ C for 30 sec., 45 $^{\circ}$ C for 1 min. and 60 $^{\circ}$ C for 3 min. was repeated 30 times by using a Thermal Cycler (Perkin-Elmer Co.). Prior to adding Taq DNA polymerase, the remaining reaction solution was mixed and was heated at 95 $^{\circ}$ C for 5 minutes and at 65 $^{\circ}$ C for 5 minutes. The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products obtained in the above step (2) were separated with a 0.8% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, PCRTM II. The recombinant vectors were introduced into

E. coli JM109 competent cells (Takara Shuzo Co., Japan)

to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothstick to obtain transformant Escherichia coli JM109/p3H2-34.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNAs thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNAs was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer, and the data of the nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan).

Homology retrieval was carried out based upon the determined nucleotide sequence [Figure 40]. As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid possessed by the transformant Escherichia coli

JM109/p3H2-34. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequence were converted into an amino acid sequence (Figure 40), and homology retrieval was carried out in view of hydrophobicity plotting (Figure 41) and at the amino acid sequence level to find homology relative to human somatostatin receptor subtype 2 (B41795) and rat-derived ligand unknown receptor (A39297) (Figure 42). Abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are usually called "Accession Numbers" or "Entry Names".

- 2 0 7 -

Example 9Cloning of Rabbit Gastropyrolic Part Smooth Muscle-Derived G Protein Coupled Receptor Protein CDNA(1) Preparation of Poly(A)⁺ RNA Fraction from Rabbit

5 Gastropyrolic Part Smooth Muscle and Synthesis of cDNA

A total RNA was prepared from rabbit gastropyrolic part smooth muscles by the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J. 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to reaction with MMLV reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE (Tris-EDTA solution).

(2) Amplification of Receptor cDNA by PCR Using Rabbit

20 Gastropyrolic Part Smooth Muscle-Derived cDNA and Sequencing

By using, as a template, 1 μ l of cDNA prepared from the rabbit gastropyrolic part smooth muscle in the above step (1), PCR amplification using the DNA primers synthesized in Example 1 was carried out. A reaction solution was composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Taq DNA polymerase and 10 μ l of buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min. was repeated 25 times by using a Thermal Cycler (Perkin-Elmer Co.). The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

- 2 0 8 -

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products obtained in the above step (2) were separated with a 1.0% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, PCRTM II. The recombinant vectors were introduced into *E. coli* JM109 competent cells (Takara Shuzo Co., Japan) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothpick to obtain transformant Escherichia coli JM109/pMD4.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNAs thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNAs was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer, and the data of the nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan). The determined nucleotide sequence was as shown in Figure 43. It was learned from Figure 43 that the cloned cDNA fragment was amplified from both sides with only the synthetic DNA primer having a nucleotide sequence represented by SEQ ID NO: 1 as synthesized in Example 1.

Homology retrieval was carried out based upon the

determined nucleotide sequence [Figure 43]. As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid possessed by the transformant *Escherichia coli* JM109/pMD4. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequence were converted into an amino acid sequence [Figure 43], and homology retrieval was carried out in view of hydrophobicity plotting [Figure 44] and at the amino acid sequence level to find homology relative to rat ligand-unknown receptor protein (A35639) [Figure 45]. Abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are usually called "Accession Numbers".

Example 10

Cloning of cDNA Comprising Whole Coding Regions for Receptor Protein from Mouse Pancreatic β -Cell Strain, MIN6-Derived cDNA Library

(1) Cloning of cDNA Comprising Whole Coding Regions for Receptor Protein from Mouse Pancreatic β -Cell Strain, MIN6-Derived cDNA Library

SuperscriptTM Lambda System (BRL, Cat. 8256) distributed by BRL Co. and GigaPack II Gold (Stratagene, Cat. 200215) distributed by Stratagene Co. were used to construct MIN6-derived cDNA libraries. By using the above kits, a MIN6 cDNA library with 2.2×10^6 pfu (plaque forming units) was constructed from 10μ g of MIN6 poly(A)⁺ RNA. The cDNA library was mixed with *E. coli* Y1090⁺ treated with magnesium sulfate, and incubated at 37 °C for 15 minutes followed by addition of 0.5% agarose (Pharmacia Co.) LB. The *E. coli* was plated onto a 1.5% agar (Wako-Junyaku Co.) LB plate (containing 50 μ g/ml of ampicillin). A nitrocellulose filter was placed on the plate on which plaques were formed and the plaque was transferred onto the filter. The filter was denatured with an alkali and then heated at 80 °C for 3 hours to fix DNAs.

The filter was incubated overnight at 42 °C together

with the probe mentioned herein below in a buffer containing 50% formamide, 5 x SSPE, 5 x Denhardt's solution, 0.1% SDS and 100 μ g/ml of salmon sperm DNA for hybridization.

The probe used was obtained by cutting the DNA fragment inserted in the plasmid, p3H2-34, obtained in Working Example 8, with EcoRI, followed by recovery and labeling by incorporation of [³²P]dCTP (Dupont Co.) with a random prime DNA labelling kit (Amasham Co.).

It was washed with 2 x SSC (150 mM NaCl and 15 mM sodium citrate), 0.1% SDS at 55 °C for 1 hour and, then, subjected to an autoradiography at -80 °C to detect hybridized plaques.

In this screening, hybridization signals were recognized in two independent plaques. Each DNA was prepared from the two clones. The DNAs digested with SalI and NotI were subjected to an agarose electrophoresis and were analyzed. Inserted fragments were identified at about 2.0kb and 3.0kb, respectively. Between them, the DNA fragment corresponding to the band at about 3.0kb (λ No.20) was selected. The λ No.20-derived NotI-SalI fragment with about 3.0kb was subcloned into the NotI-SalI site of the plasmid, pBluescriptTM II SK(+), and *E. coli* JM109 was transformed with the plasmid to obtain a transformant *E. coli* JM109/pMGR20. A restriction enzyme map of the plasmid, pMGR20, was prepared relying upon a restriction enzyme map deduced from the nucleotide sequence as shown in Working Example 8. As a result, it was learned that it carried a full-length receptor protein-encoding DNA which was predicted from the receptor protein-encoding DNA as shown in Working Example 8.

(2) Sequencing of MIN6-Derived Receptor Protein Full-Length cDNA

Among the NotI-SalI fragments inserted in the plasmid, pMGR20, obtained in the above step (1), the nucleotide sequence with total 1607bp, including not only a region that is considered to be a receptor protein-coding region (ORF) but

also a neighboring region thereof was sequenced. Concretely speaking, by utilizing restriction enzyme sites that exist in the NotI-SalI fragments, unnecessary parts were removed or necessary fragments were subcloned in order to prepare template plasmids for analyzing the nucleotide sequence thereof. As for the nucleotide sequences of part of the regions, primers for sequencing were synthesized based upon the nucleotide sequences that were determined already and used to make confirmation.

The reaction for determining the nucleotide sequence (sequencing) was carried out with a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNA was decoded with the fluorescent automatic sequencer (ABI Co.), and the data of the nucleotide sequence obtained were analyzed with DNASIS (Hitachi System Engineering Co., Japan).

Figure 46 shows a nucleotide sequence around an open reading frame (ORF) of a mouse galanin receptor protein encoded by the cDNA insert in pMGR20. The nucleotide sequence of mouse galanin receptor protein-encoding DNA corresponds to from the 481st to 1525th nucleotides of the nucleotide sequence in Figure 46. The nucleotide sequence was converted into an amino acid sequence [Figure 46] and hydrophobicity plotting was carried out [Figure 47]. Since the amino acid sequence [Figure 46] has 92% homology to the human-derived galanin receptor protein at the amino acid sequence level [Figure 48], it was learned that the cDNA insert in the pMGR20 is a mouse-derived galanin receptor protein-encoding cDNA.

Example 11

Preparation of Synthetic DNA Primer for Amplifying G Protein Coupled Receptor Protein-Encoding DNA

Highly homologous parts were found by comparing nucleotide sequences corresponding to or near the third membrane-spanning domain [2C and 3D in Figure 4] and the sixth membrane-spanning domain [6C of Figure 6] among known G protein coupled receptors, i.e., rat-derived angiotensin II

receptor protein (L32840), rat-derived angiotensin II receptor protein (X64052), rat-derived angiotensin receptor protein subtype (M90065), human-derived angiotensin Ia receptor protein (M91464), rat-derived cholecystokinin_A receptor protein (M88096), rat-derived cholecystokinin_B receptor protein (M99418), human-derived cholecystokinin_B receptor protein (L04473), mouse-derived low-affinity interleukin 8 receptor protein (M73969), human-derived high-affinity interleukin 8 receptor protein (X65858), mouse-derived C5a anaphylatoxin receptor protein (S46655), human-derived N-formyl peptide receptor protein (M60626), etc.

The aforementioned abbreviations in parentheses are reference numbers that are indicated when the GenBank/EMBL data base is retrieved, and are usually called "Accession Numbers".

It was planned to incorporate mixed bases relying upon the base regions that were in agreement with a large number of receptor protein cDNAs in order to enhance base agreement of sequences with as many receptor cDNAs as possible even in other regions. Based upon these sequences, the degenerate synthetic DNA (3D of Figure 4) having a nucleotide sequence represented by SEQ ID NO: 3 which is complementary to the homologous nucleotide sequence of Figure 4 and the degenerate synthetic DNA (nucleotide sequence complementary to 6C of Figure 6) having a nucleotide sequence represented by SEQ ID NO: 4 were produced. Nucleotide synthesis was carried out by a DNA synthesizer.

[Synthetic DNAs]

5'-CTCGC (G or C) GC (C or T) (A or C) TT (A or G) G
(C or T) ATGGA (C or T) CGTAT-3' (SEQ ID NO:3)

5'-CATGT (A or G) G (T or A) AGGATTCAG (G or C) A
(A or C) AT (A or G) A (A or G)(A or G) AA-3' (SEQ ID NO:4)

The parentheses indicate the incorporation of a plurality of bases, leading to multiple oligonucleotides in the primer preparation. In other words, nucleotide residues in parentheses of the aforementioned DNAs were incorporated in the presence of a mixture of plural bases at the time of synthesis, provided that I denotes inosine.

Example 12

Cloning of Rabbit Gastropyloric Part Smooth Muscle-Derived G Protein Coupled Receptor Protein cDNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Rabbit Gastropyloric Part Smooth Muscle and Synthesis of cDNA

A total RNA was prepared from rabbit gastropyloric part smooth muscles by the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J. 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to reaction with MMLV reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE.

(2) Amplification of Receptor cDNA by PCR Using Rabbit Gastropyloric Part Smooth Muscle-Derived cDNA and Sequencing

By using, as a template, 1 μ l of cDNA prepared from the rabbit gastropyloric part smooth muscle in the above step (1), PCR amplification using the DNA primer having a nucleotide sequence represented by SEQ ID NO: 3 and the DNA primer having a nucleotide sequence represented by SEQ ID NO: 4 synthesized in Example 11 was carried out.

A reaction solution was composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Tag DNA polymerase

and 10 μ l of buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min. was repeated 25 times by using a Thermal Cycler (Perkin-Elmer Co.). The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products obtained in the above step (2) were separated with a 1.0% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, pCR™ II. The recombinant vectors were introduced into *E. coli* JM109 competent cells (Takara Shuzo Co., Japan) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothstick to obtain transformant *Escherichia coli* JM109/pMJ10.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNAs thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNAs was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer, and the data of the

nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan). The determined nucleotide sequence was as shown in Figure 49.

Homology retrieval was carried out based upon the determined nucleotide sequence [Figure 49]. As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid possessed by the transformant *Escherichia coli* JM109/pMJ10. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequence were converted into an amino acid sequence [Figure 49], and homology retrieval was carried out in view of hydrophobicity plotting [Figure 50] and at the amino acid sequence level to find homology relative to human ligand unknown receptor protein (B42009), human N-formyl peptide receptor protein (JC2014), rabbit N-formyl peptide receptor protein (A46520), mouse C5a anaphylatoxin receptor protein (A46525) and bovine neuropeptide Y receptor protein (S28787) [Figure 51]. Abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR/Swiss-PROT and are usually called "Accession Numbers".

Example 13

Preparation of Synthetic DNA Primer for Amplifying G Protein Coupled Receptor Protein-Encoding DNA

A comparison of nucleotide sequences coding for regions corresponding to or near the third membrane-spanning domain among known G protein coupled receptors, i.e., mouse-derived α -opioid receptor protein (L11064), mouse-derived δ -opioid receptor protein (L11065), rat-derived μ -opioid receptor protein (D16349), mouse-derived bradykinin B2 receptor protein (X69676), rat-derived bradykinin B2 receptor protein (M59967), mouse-derived bombesin receptor protein (M55328), human-derived neuromedin B receptor protein (M73482), human-derived gastrin releasing peptide receptor protein (M73481), human-derived bombesin receptor protein subtype 3 (L08893), mouse-derived substance K receptor protein

(X62933), mouse-derived substance P receptor protein (X62934), rat-derived neurokinin 3 receptor protein (J05189), rat-derived endothelin receptor protein (M60786), rat-derived ligand unknown receptor proteins (L04672), (X61496), (X59249) and (L09249), mouse-derived ligand unknown receptor protein (P30731), human-derived ligand unknown receptor proteins (M31210) and (U03642), etc. was made. In particular, the degenerate DNA primer having a nucleotide sequence (3B in Figure 3; SEQ ID NO: 6) with highly common bases (highly homologous nucleotides) was synthesized to enhance base agreement of sequences with as many receptor cDNAs as possible even in other regions on the basis of nucleotide sequence regions that were in agreement with a large number of receptor cDNAs. Nucleotide synthesis was carried out by a DNA synthesizer.

The nucleotide sequence represented by SEQ ID NO: 6

is:

5'-CTGAC (C or T) G (C or T) TCTT (A or G)(G or C) I
(A or G)(C or T) TGAC (A or C) G (A, C or G) TAT-3'

The parentheses indicate the incorporation of a plurality of bases, leading to multiple oligonucleotides in the primer preparation. In other words, nucleotide residues in parentheses of the aforementioned DNAs were incorporated in the presence of a mixture of plural bases at the time of synthesis, provided that I denotes inosine.

Furthermore, a comparison of nucleotide sequences coding for regions corresponding to or near the sixth membrane-spanning domain among known G protein coupled receptors, i.e., mouse-derived α -opioid receptor protein (L11064), mouse-derived δ -opioid receptor protein (L11065), rat-derived μ -opioid receptor protein (D16349), mouse-derived bradykinin B2 receptor protein (X69676), rat-derived bradykinin B2 receptor protein (M59967), mouse-derived bombesin receptor protein (M55328), human-derived neuromedin B receptor protein

(M73482), human-derived gastrin releasing peptide receptor protein (M73481), human-derived bombesin receptor protein subtype 3 (L08893), mouse-derived substance K receptor protein (X62933), mouse-derived substance P receptor protein (X62934), rat-derived neurokinin 3 receptor protein (J05189), rat-derived endochelin receptor protein (M60786), rat-derived ligand unknown receptor proteins (L04672), (X61496), (X59249) and (L09249), mouse-derived ligand unknown receptor protein (P30731), human-derived ligand unknown receptor proteins (M31210) and (U03642), etc. was made. In particular, the degenerate DNA primer having a nucleotide sequence (SEQ ID NO: 8) which is complementary to the nucleotide sequence (6A in Figure 5) with highly common bases (highly homologous nucleotides) was synthesized to enhance base agreement of sequences with as many receptor cDNAs as possible even in other portions on the basis of base regions that are in agreement with a large number of receptor cDNAs.

The nucleotide sequence represented by SEQ ID NO: 8 is:

20 5'-GATGTG (A or G) TA (A or G) GG (G or C)(A or G)
ICCAACAGATC (A or G) (C or T) AAA-3'

The parentheses indicate the incorporation of a plurality of bases, leading to multiple oligonucleotides in the primer preparation. In other words, nucleotide residues in parentheses of the aforementioned DNAs were incorporated in the presence of a mixture of plural bases at the time of synthesis, provided that I denotes inosine.

The aforementioned abbreviations in parentheses are reference numbers indicated when the GenBank/EMBL data base is retrieved and are usually called "Accession Numbers".

Example 14

Cloning of Rabbit Gastropyloric Part Smooth Muscle-Derived G Protein Coupled Receptor Protein cDNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Rabbit

Gastropyloric Part Smooth Muscle and Synthesis of cDNA

A total RNA was prepared from rabbit gastropyloric part smooth muscles by the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J. 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to reaction with MMLV reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE.

(2) Amplification of Receptor cDNA by PCR Using Rabbit Gastropyloric Part Smooth Muscle-Derived cDNA and Sequencing

By using, as a template, 1 μ l of cDNA prepared from the rabbit gastropyloric part smooth muscle in the above step (1), PCR amplification using the DNA primer having a nucleotide sequence represented by SEQ ID NO: 6 and the DNA primer having a nucleotide sequence represented by SEQ ID NO: 8 synthesized in Example 13 was carried out.

A reaction solution was composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Taq DNA polymerase and 10 μ l of buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min. was repeated 25 times by using a Thermal Cycler (Perkin-Elmer Co.). The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products obtained in the above step (2) were separated by using a 1.0% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were heat-melted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, PCRTM II. The recombinant vectors were introduced into E. coli JM109 competent cells (Takara Shuzo Co., Japan) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothpick to obtain transformant Escherichia coli JM109/pMH28.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with an automatic plasmid extracting machine (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNAs thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNAs was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer, and the data of the nucleotide sequences obtained were read by using DNASIS (Hitachi System Engineering Co., Japan). The determined nucleotide sequence was as shown in Figure 52.

Homology retrieval was carried out based upon the determined nucleotide sequence [Figure 52]. As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid possessed by the transformant Escherichia coli

JM109/pMH28. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequence were converted into an amino acid sequence [Figure 52], and homology retrieval was carried out in view of hydrophobicity plotting [Figure 53] and at the amino acid sequence level to find homology relative to mouse IL-8 receptor protein (P35343), human somatostatin receptor protein 1 (A41795) and human somatostatin receptor protein 4 (A47457) [Figure 54].

The aforementioned abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR or SWISS-PROT and are usually called "Accession Numbers".

Example 15

Preparation of Synthetic DNA Primer for Amplifying G Protein Coupled Receptor Protein-Encoding DNA

A comparison of nucleotide sequences coding for regions corresponding to or near the second membrane-spanning domain among known G protein coupled receptors, i.e., human-derived galanin receptor (HUMGALREC), rat-derived α -1B-adrenergic receptor (RATADRLB), human-derived β -1-adrenergic receptor (HUMADRL1), rabbit-derived IL-8 receptor (RABIL8RSB), human-derived opioid receptor (HUMOPIDRE), bovine-derived substance K receptor (BTSKR), human-derived somatostatin receptor-2 (HUMSTR12A), human-derived somatostatin receptor-3 (HUMSTR3Y), human-derived gastrin receptor (HUMGARE), human-derived cholecystokinin A receptor (HUMCKAR), human-derived dopamine receptor-D5 (HUMDLB), human-derived serotonin receptor 5HT1E (HUM5HT1E), human-derived dopamine receptor D4 (HUMD4C), mouse-derived serotonin receptor-2 (MMSERO), rat-derived α -1A-adrenergic receptor (RATADRA1A), rat-derived histamine H2 receptor (S57565), etc. was made. In particular, the degenerate DNA primer having a nucleotide sequence (T2A in Figure 7, SEQ ID NO: 10) with highly common bases (highly homologous nucleotides) was synthesized to enhance base agreement of sequences with as many receptor cDNAs as possible even in other regions on the basis of nucleotide sequence

regions that were in agreement with a large number of receptor cDNAs. Nucleotide synthesis was carried out by a DNA synthesizer.

The nucleotide sequence represented by SEQ ID NO: 10

5 is:

5'-GYCACCAACN₂WSTTCATCCTSWN₂HCTG-3'

wherein S represents G or C; Y represents C or T; W represents A or T; H represents A, C or T and N₂ represents I.

The parentheses indicate the incorporation of a plurality of bases, leading to multiple oligonucleotides in the primer preparation. In other words, nucleotide residues in parentheses of the aforementioned DNAs were incorporated in the presence of a mixture of plural bases at the time of synthesis, provided that I denotes inosine.

Furthermore, a comparison of nucleotide sequences

coding for regions corresponding to or near the seventh membrane-spanning domain among known G protein coupled receptors, i.e., human-derived galanin receptor (HUMGALAREC), rat-derived A1 adenosine receptor (RAT1ADREC), porcine-derived angiotensin receptor (PIGA2R), rat-derived serotonin receptor (RAT5HTRTC), human-derived dopamine receptor (S58541),

human-derived gastrin releasing peptide receptor (HUMGRPR), mouse-derived GRP/bombesin receptor (MUSGRPBOM), rat-derived mouse-derived angiotensin receptor (RRVT1AIR), vascular type 1 angiotensin receptor (RRVT1AIR), human-derived muscarinic acetylcholine receptor (HSHM4), human-derived β -1 adrenergic receptor (HUMDRB1),

human-derived gastrin receptor (HUMGARE), rat-derived

cholecystokinin receptor (RATCCKAR), rat-derived ligand unknown receptor (S59748), human-derived somatostatin receptor (HUMST28A), rat-derived ligand unknown receptor (RNGPROCR), mouse-derived somatostatin receptor 1 (MUSSRI1A), human-derived α -A1-adrenergic receptor (HUMA1AADR), mouse-derived delta-opioid receptor (S66181), human-derived somatostatin

receptor-3 (HUMSSTR3Y), etc. was made. In particular, the degenerate DNA primer having a nucleotide sequence (T7A in Figure 8, SEQ ID NO: 11) with highly common bases (highly homologous nucleotides) was synthesized to enhance base agreement of sequences with as many receptor cDNAs as possible even in other regions on the basis of nucleotide sequence regions that were in agreement with a large number of receptor cDNAs. Nucleotide synthesis was carried out by a DNA synthesizer.

The nucleotide sequence represented by SEQ ID NO: 11

is:

5'-ASN₂SAN₂RAAGSARTAGAN₂GAN₂RGGRTT-3'

wherein R represents A or G; S represents G or C and N₂ represents I.

The parentheses indicate the incorporation of a

plurality of bases, leading to multiple oligonucleotides in the primer preparation. In other words, nucleotide residues in parentheses of the aforementioned DNAs were incorporated in the presence of a mixture of plural bases at the time of synthesis, provided that I denotes inosine.

The aforementioned abbreviations in parentheses are reference numbers indicated when the GenBank/EMBL data base retrieved and are usually called "Accession Numbers".

Example 16

Cloning of Rabbit Gastropyloric Part Smooth Muscle-Derived G Protein Coupled Receptor Protein cDNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Rabbit Gastropyloric Part Smooth Muscle and Synthesis of cDNA

A total RNA was prepared from rabbit gastropyloric part smooth muscles by the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J. 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the

poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to reaction with MMLV reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE.

(2) Amplification of Receptor cDNA by PCR Using Rabbit Gastropyloic Part Smooth Muscle-Derived cDNA and Sequencing

By using, as a template, 1 μ l of cDNA prepared from the rabbit gastropyloic part smooth muscle in the above step (1), PCR amplification using the DNA primer having a nucleotide sequence represented by SEQ ID NO: 10 and the DNA primer having a nucleotide sequence represented by SEQ ID NO: 11 synthesized in Example 15 was carried out.

A reaction solution was composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Taq DNA polymerase and 10 μ l of buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min. was repeated 25 times with a Thermal Cycler (Perkin-Elmer Co.). The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products obtained in the above step (2) were separated with a 1.4% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were eluted electrophoretically, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.),

the recovered DNAs were subcloned to the plasmid vector, PCRTM II. The recombinant vectors were introduced into *E. coli* JM109 competent cells (Takara Shuzo Co., Japan) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothpick to obtain 100 transformant clones.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with the automatic plasmid extracting machine PI-100 (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNA thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNA was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer.

Homology retrieval was carried out based upon the determined nucleotide sequence by using DNASIS (Hitachi System Engineering Co., Japan). As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment insert in the plasmid possessed by the transformant *Escherichia coli* JM109/PMN7. Figure 56 and Figure 56 show the nucleotide sequences of the cDNA fragments. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan), the nucleotide sequences were converted into amino acid sequences [Figure 55] and [Figure 56], and hydrophobicity plotting was carried out [Figure 57]. As a result, the presence of hydrophobic domains which prove that it is a G protein coupled receptor protein were confirmed. Furthermore, homology retrieval was carried out at the amino acid sequence level to find that the DNAs were novel receptor proteins having 28% homology relative to rat-derived β_3 -adrenaline receptor protein (A41679), 29% homology relative to rat-derived serotonin (5-HT6) receptor protein (J05911),

27% homology relative to dog-derived histamine H_2 receptor protein (A39008), 27% homology relative to human-derived somatostatin receptor (type 4) protein (JN0605), 24% homology relative to human-derived dopamine D_1 receptor protein (S11377), 23% homology relative to rat-derived neurotensin receptor protein (JH0164), 31% homology relative to human-derived cholecystokinin B receptor protein (JC1352), and 30% homology relative to rat-derived gastrin receptor protein (JQ1614). The aforementioned abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR and are usually called "Accession Numbers".

Example 17

Amplification of Receptor cDNA by PCR Using MIN6-Derived cDNA and Sequencing

By using, as a template, 5 μ l of cDNA prepared from the mouse pancreatic β -cell strain, MIN6 in Working Example 4 (1), PCR amplification using the DNA primers synthesized in Example 4 (2) as disclosed in Libert F. et al., "Science, 244:569-572, 1989", i.e., a synthetic primer represented by the following sequence:

5'-CTGTC (C or T) G (C or T) (G or C) AT (C or T) GCIT
(G or T) GA (C or T) (A or C) G (G or C) TAC-3'

(SEQ ID NO: 60)

wherein I is inosine; and
a synthetic primer represented by the following sequence:

5'-A (G or T) G (A or T) AG (A or T) AGGCAGCCAGCAGAI
(G or C) (A or G) (C or T) GAA-3'

(SEQ ID NO: 61)

wherein I is inosine, was carried out under the same conditions as in Example 3 (1). The resulting PCR product was subcloned to the plasmid vector, PCRTM II, in the same manner as in Example 3 (2) to obtain a plasmid, p5S38. The plasmid p5S38 was transfected into *E. coli* JM109 to obtain

transformant *Escherichia coli* JM109/p5S38.

The reaction for determining the nucleotide sequence (sequencing) was carried out with a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNA was decoded with the fluorescent automatic sequencer (ABI Co.), and the data of the nucleotide sequence obtained were read with DNASIS (Hitachi System Engineering Co., Japan).

Figure 62 shows a mouse pancreatic β -cell strain MIN6-derived G protein coupled receptor protein-encoding DNA (SEQ ID NO: 33) and an amino acid sequence (SEQ ID NO: 28) encoded by the isolated DNA based upon the nucleotide sequence of plasmid, p5S38. The underlined portions represent regions corresponding to the synthetic primers.

Homology retrieval was carried out based upon the determined nucleotide sequence (Figure 62). As a result, it was learned that a novel G protein coupled receptor protein was encoded by the cDNA fragment obtained. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan), the nucleotide sequence was converted into an amino acid sequence (Figure 62), and hydrophobicity plotting was carried out to confirm the presence of four hydrophobic regions (Figure 64). Upon comparing the amino acid sequence with those encoded by p19P2 obtained in Example 3 (2) and encoded by pG3-2 obtained in Example 4 (2), furthermore, a high degree of homology was found as shown in Figure 63.

As a result, it is strongly suggested that the mouse pancreatic β -cell strain, MIN6-derived G protein coupled receptor protein encoded by p5S38 recognizes the same ligand as the human pituitary gland-derived G protein coupled receptor protein encoded by p19P2 does while the animal species from which the receptor protein encoded by p5S38 is derived is different from that from which the receptor protein encoded by p19P2 is. It is also strongly suggested that the mouse pancreatic β -cell strain, MIN6-derived G protein coupled receptor protein encoded by p5S38 recognizes the same ligand as the mouse pancreatic β -cell strain, MIN6-derived G protein coupled receptor proteins encoded by pG3-2 and pG1-10 do and

- 2 2 7 -

they are analogous receptor proteins one another (so-called "subtype").

Example 18

Northern Hybridization with cDNA Fragment Included in MIN6-Derived Receptor Protein-Encoding p3H2-17

Mouse cell line, MIN6, Neuro-2a, poly(A)⁺ RNA (2.5 μ g) and mouse brain, spleen, thymus and pancreas poly(A)⁺ RNAs (2.5 μ g) were used as poly(A) RNAs. The DNA fragment inserted into the plasmid, p3H2-17, obtained in Example 7 (3)

was recovered as a DNA fragment with about 400bp by cutting the plasmid with EcoRI and the resulting DNA fragment was labeled by incorporation of [³²P]dCTP (Dupont Co.) with a random prime DNA labeling kit (Amasham Co.). The about 400bp labeled DNA fragment was used as a probe for hybridization.

Nylon membrane (Pall Biodyne, U.S.A.) was used as a filter for northern blotting and migration of the poly(A)⁺ RNA and adsorption (sucking) thereof with the blotting filter was carried out according to the method as disclosed in Molecular Cloning, Cold Spring Harbor Laboratory Press, 1989.

The hybridization was carried out by incubating the above-mentioned filter and probe in a buffer containing 50% formamide, 5 x SSPE (20 x SSPE (pH 7.4) is 3 M NaCl, 0.2 M NaH₂PO₄·H₂O, 25 mM EDTA), 5 x Denhardt's solution (Nippon Gene, Japan), 0.1% SDS and 100 μ g/ml of salmon sperm DNA overnight at 42 °C. The filter was washed with 0.1 x SSC (20 x SSC is 3 M NaCl, 0.3 M sodium citrate), 0.1% SDS at 50 °C and, after drying with an air, was exposed to an X-ray film (XAR5, Kodak) for 15 days at -80 °C. The results were as shown in Figure 65.

It is considered from Figure 65 that mRNA for the receptor gene encoded by the cDNA fragment included in p3H2-17 is expressed in the cell line, MIN6, Neuro-2a, and the mouse brain, pancreas, spleen and thymus and especially expressed in the mouse spleen and thymus intensely. The MIN6 signal position hybridizable in the northern hybridization plotting is different from those of other organ cells.

- 2 2 8 -

Example 19

PCR Cloning of cDNA Comprising Whole Coding Regions of Receptor Proteins from Mouse Spleen, Thymus-Derived poly(A)⁺ RNA and Sequencing

(1) PCR Cloning of cDNA Comprising Whole Coding Region of Receptor Protein

In order to obtain a full-length open reading frame (coding region) of the receptor protein encoded by the cDNA fragment included in p3H2-17, PCR amplification was carried out by 5'RACE and 3'RACE wherein poly(A)⁺ RNA derived from mouse spleen and thymus was used.

Based on the nucleotide sequence of 3H2-17 which was disclosed, the following 4 primers were synthesized:

- (Nucleotide sequence of synthesized primer)
- ① 5'-TAGTGTGTGACTGCTGGCTG-3' (SEQ ID NO: 20)
 - ② 5'-AGTCTTGTGCTGCACAGCATCCACG-3' (SEQ ID NO: 21)
 - ③ 5'-CAGCCAGTAAGCATATGAGGCGACG-3' (SEQ ID NO: 22)
 - ④ 5'-ACAGAGCTGCTGGCCATCTGCGCACACA-3' (SEQ ID NO: 23)

The 5'RACE was carried out according to the protocol of 5'Amplicon Finder RACE kit from Clontech Co. (Clontech Co.).

In an embodiment, cDNA was prepared from 2 μ g each of poly(A)⁺ RNAs derived from mouse spleen and thymus by using the aforementioned primer ④ and ligated with an anchor attached to the 5'RACE kit. A mixture of a 1/200 amount of the cDNA thus prepared, the anchor and the aforementioned primer ④ was subjected to PCR using 4 polymerases, Taq (Takara, Japan), Ex Tag (Takara, Japan), Vent (New England Biolabs) and Pfu (Stratagene) under the following conditions: 96 °C for 30 sec., 60 °C for 60 sec., 72 °C for 90 sec. and 35 cycles. A 1/5 amount of the PCR product was subjected to agarose electrophoresis and stained with ethidium bromide (EtBr). The results are shown in Figure 66. The amplified DNA band

appeared at an about 1 kbp position and the isolated about 1 kbp DNA band which was synthesized from poly(A)⁺ RNAs derived from mouse spleen and thymus by the 5'RACE using Ex Taq polymerase was treated with SUPRECTM-01 (Takara, Japan) to recover cDNA.

The isolated DNA was subcloned into PCRTM II vector by using a TA Cloning Kit (Invitrogen Co.) and the vector was transfected into *E. coli* JM109 to obtain 3 transformant clones, N26, N64 and N75. The clone, N26, holds the thymus-derived cDNA which is amplified by the 5'RACE and the clone, N75, holds the spleen-derived cDNA which is amplified by the 5'RACE (Figure 68).

The 3'RACE was carried out according to the protocol of 3' RACE kit (GIBCO BRL Co.).

In an embodiment, cDNA was prepared from 1 μ g each of poly(A)⁺ RNAs derived from mouse spleen and thymus by using an adaptor primer attached to the 3' RACE kit. A mixture of the adaptor primer thus prepared and a 1/10 amount of cDNA which was prepared by using the aforementioned primer ① was subjected to 1st PCR using 4 polymerases, Tag (Takara, Japan), Ex Taq (Takara, Japan), Vent (NEB) and Pfu (Stratagene) under the following conditions: 96 °C for 30 sec., 55 °C for 60 sec., 72 °C for 120 sec. and 30 cycles. A mixture of a 1/50 amount of the 1st PCR product, the aforementioned primer ② and the adaptor primer was subjected to 2nd PCR using the aforementioned polymerases under the same conditions as aforementioned herein in the 5'RACE process. A 1/5 amount of the 2nd PCR product was subjected to agarose electrophoresis and stained with ethidium bromide. The results are shown in Figure 67.

The amplified DNA band appeared at an about 1 kbp position (which was synthesized from poly(A)⁺ RNAs derived from mouse thymus by the 3'RACE using Vent polymerase) and the amplified DNA band appeared at an about 1 kbp position (which was synthesized from poly(A)⁺ RNAs derived from mouse thymus by the 3'RACE using Pfu polymerase) were treated with

SUPRECTM-01 (Takara, Japan) to recover cDNA, respectively.

The isolated DNAs were treated with T4 polynucleotide kinase (Wako Pure Chemical Co., Japan) to add phosphate to the end thereof and the phosphorylated DNAs were ligated with pUC18 SmaI BAP (Pharmacia) by using DNA Ligation Kit (Takara, Japan) followed by transformation of *E. coli* JM109 to obtain 3 transformant clones, C2, C13 and C15. The clones, C13 and C15, hold the thymus-derived cDNA which is amplified by the 3'RACE and the clone, C2, holds the thymus-derived cDNA which is amplified by the 3'RACE (Figure 68).

Based on the nucleotide sequences of clones, N26, N64 and N75, which are considered to hold the N-terminal region of the open reading frame (ORF) of the cDNA fragment included in p3H2-17 and the nucleotide sequences of clones, C2, C13 and C15, which are considered to hold the C-terminal region of the open reading frame (ORF) of the cDNA fragment included in p3H2-17, the entire nucleotide sequence coding for the open reading frame and neighboring region of the receptor protein encoded by the cDNA included in p3H2-17 was determined.

To be more specific, sequencing was carried out with the primers used in the 5'RACE and 3'RACE or synthetic primers for sequencing by using a DyeDeoxy Terminator Cycle Sequencing Kit (ABI Co.), the nucleotide sequences were decoded by using a fluorescent automatic sequencer. The obtained data of the DNA were analyzed by DNASIS (Hitachi System Engineering Co., Japan).

PCR errors which presumably happen to occur upon PCR have been corrected by a way of thinking that, when nucleotides between two clones which are independently produced by PCR are identical (e.g. those between clones, N75 and N64, are identical) each other, the identical base is considered as correct. The determined nucleotide sequence is shown in Figure 69. The amino acid sequence is deduced based on the determined nucleotide sequence (Figure 69). Hydrophobicity plotting was carried out based on the deduced amino acid sequence (Figure 70). As a result, it was learned that it was a seven transmembrane G protein coupled receptor, as

it is suggested from the cDNA fragment included in p3H2-17.

Homology retrieval at the amino acid level indicates that it is homologous to mouse P_2 purinoceptor and chicken P_2 purinoceptor.

Further, the clone which are free of an error in the open reading frame (ORF) was selected and used to construct plasmids carrying the full-length ORF of the receptor protein encoded by p3H2-17. In an embodiment, the cDNA fragment held by the clone, N75, was digested with restriction enzymes, DraIII and EcoRI, to obtain cDNA fragments which are the N-terminal region of the receptor protein held by p3H2-17. The C-terminal cDNA fragment encoded by C13 was digested with restriction enzymes, DraIII and EcoRI, to delete 5'-side regions from the DraIII site of the C-terminal and the long fragment was obtained by the digestion of C13 with restriction enzymes, DraIII and EcoRI. The N75-derived N-terminal cDNA DraIII-EcoRI fragment was ligated with the long C13-derived DraIII-EcoRI fragment by using a DNA Ligation Kit (Takara, Japan) and transfected into *Escherichia coli* JM109 to obtain transformant *Escherichia coli* JM109/pMAH2-17.

(2) Electrophysiological Measurement of Receptor Encoded by pMAH2-17

The receptor encoded by pMAH2-17 was examined electrophysiologically in *Xenopus* oocytes. The ORF of the receptor encoded by pMAH2-17 was inserted into the XhoI-XbaI sites of pBluescript TM II SK(+) (Stratagene) with directing the sequence thereof downstream from T7 promoter. The resulting plasmid as a template was treated with a mCAP TM mRNA Capping kit (Stratagene) to produce cRNA of this receptor gene.

The cRNA was injected into *Xenopus* oocytes (50ng cRNA/50nl/oocyte), previously prepared according to the method disclosed in Nathan Dascal et al., Proc. Natl. Acad. Sci. USA, Vol. 90, pp.6596-6600 (1993). The cRNA-injected oocytes were incubated at 20 °C for 2 to 3 days and subjected to electrophysiological measurements. The measurement was carried

out with a microelectrode-applicable high input resistance amplifier (MEz-8300, Nippon Kodon, Co., Japan), and a voltage clamping amplifier (CEz-2/200, Nippon Kodon, Co., Japan). The initial membrane potential of oocytes was set to -60 mV and responses (current changes of the membrane) evoked by addition of ligands were recorded with a recorder (Thermal Array recorder, Nippon Kodon, Co., Japan) (Nathan Dascal et al., Proc. Natl. Acad. Sci. USA, Vol. 90, pp.6596-6600 (1993)).

Typical inward currents elicited upon activation of phospholipase C-coupled receptors were observed in oocytes injected with pMAH2-17 cRNA via stimulation by 10 μ M ATP (Figure 75). In contrast, such a current was not observed in oocytes injected with H_2O , instead of pMAH2-17 cRNA, by the ATP stimulation.

In conclusion, it is considered that the receptor encoded by pMAH2-17 cRNA is classified into a subtype within the ATP receptor, P_2 purinoceptor.

Example 20

Cloning of Rabbit Gastropyrolic Part Smooth Muscle-Derived G Protein Coupled Receptor Protein cDNA

(1) Preparation of Poly(A)⁺ RNA Fraction from Rabbit Gastropyrolic Part Smooth Muscle and Synthesis of cDNA

A total RNA was prepared from rabbit gastropyrolic part smooth muscles by the guanidine thiocyanate method (Kaplan B.B. et al., Biochem. J. 183, 181-184 (1979)) and, then, poly(A)⁺ RNA fractions were prepared with a mRNA purifying kit (Pharmacia Co.). Next, to 5 μ g of the poly(A)⁺ RNA fraction was added a random DNA hexamer (BRL Co.) as a primer, and the resulting mixture was subjected to the reaction with MMLV reverse transcriptase (BRL Co.) in the buffer attached to the MMLV reverse transcriptase kit to synthesize complementary DNAs. The reaction product was extracted with phenol/chloroform (1:1), precipitated in ethanol, and was then dissolved in 30 μ l of TE.

(2) Amplification of Receptor cDNA by PCR Using Rabbit Gastropyloric Part Smooth Muscle-Derived cDNA and Sequencing

By using, as a template, 1 μ l of cDNA prepared from the rabbit gastropyloric part smooth muscle in the above step (1), PCR amplification using the DNA primer having a nucleotide sequence represented by SEQ ID NO: 10 and the DNA primer having a nucleotide sequence represented by SEQ ID NO: 4 synthesized in Example 15 was carried out.

A reaction solution was composed of the synthetic DNA primers (SEQ: 5' primer sequence and 3' primer sequence) each in an amount of 100 pM, 0.25 mM dNTPs, 1 μ l of Taq DNA polymerase and 10 μ l of buffer attached to the enzyme kit, and the total amount of the reaction solution was made to be 100 μ l. The cycle for amplification including 96 °C for 30 sec., 45 °C for 1 min. and 60 °C for 3 min. was repeated 25 times by using a Thermal Cycler (Perkin-Elmer Co.). The amplified products were confirmed relying upon 1.2% agarose gel electrophoresis and ethidium bromide staining.

(3) Subcloning of PCR Product into Plasmid Vector and Selection of Novel Receptor Candidate Clone via Decoding Nucleotide Sequence of Inserted cDNA Region

The PCR products obtained in the above step (2) were separated by using a 1.0% low-melting temperature agarose gel, the band parts were excised from the gel with a razor blade, and were electro-eluted, extracted with phenol and precipitated in ethanol to recover DNAs. According to the protocol attached to a TA Cloning Kit (Invitrogen Co.), the recovered DNAs were subcloned to the plasmid vector, PCR™ II. The recombinant vectors were introduced into *E. coli* JM109 competent cells (Takara Shuzo Co., Japan) to produce transformants. Then, transformant clones having a cDNA-inserted fragment were selected in an LB agar culture medium containing ampicillin, IPTG and X-gal. Only transformant clones exhibiting white color were picked with a sterilized toothstick to obtain 100 transformant clones.

The individual clones were cultured overnight in an LB culture medium containing ampicillin and treated with the automatic plasmid extracting machine PI-100 (Kurabo Co., Japan) to prepare plasmid DNAs. An aliquot of the DNAs thus prepared was cut by EcoRI to confirm the size of the cDNA fragment that was inserted. An aliquot of the remaining DNAs was further processed with RNase, extracted with phenol/chloroform, and precipitated in ethanol so as to be condensed. Sequencing was carried out by using a DyeDeoxy terminator cycle sequencing kit (ABI Co.), the DNAs were decoded by using a fluorescent automatic sequencer.

Homology retrieval was carried out based upon the determined nucleotide sequence. As a result, it was learned that a novel G protein coupled receptor protein was been encoded by the cDNA fragment insert in the plasmid possessed by the transformant *Escherichia coli* JM109/pMN128. The nucleotide sequences of the cDNA fragments are shown in Figures 71 and 72. To further confirm this fact, by using DNASIS (Hitachi System Engineering Co., Japan) the nucleotide sequences were converted into amino acid sequences [Figure 71 and Figure 72], and homology retrieval was carried out in view of hydrophobicity plotting [Figure 73] and at the amino acid sequence level to find a novel receptor protein which has 27% homology relative to hamster-derived β_2 -adrenaline receptor protein (A03159), 20% homology relative to rat-derived bradykinin receptor (type B₂) protein (A41283), 24% homology relative to human-derived dopamine D₁ receptor protein (S11377) and 23% homology relative to human-derived blue sensitive opsin receptor protein (A03156). The aforementioned abbreviations in parentheses are reference numbers assigned when they are registered as data to NBRF-PIR and are usually called "Accession Numbers".

Example 21

Cloning of cDNA Comprising Whole Coding Regions for Receptor Protein from Human-Derived DNA Library

The DNA library constructed by Clontech wherein

5 λ gt11 phage vector is used (Clontech Laboratories, Inc.; CLH L1008b) was employed as a human placenta-derived cDNA library. The human placenta cDNA library (1×10^5 pfu (plaque forming units)) was thermally denatured. By using the human placenta-derived cDNA library, PCR amplification using the DNA primer having a nucleotide sequence represented by SEQ ID NO: 20 and the DNA primer having a nucleotide sequence represented by SEQ ID NO: 23 synthesized in Example 19 was carried out.

(Nucleotide sequence of synthesized primer)

15 ① 5'-TAGTGTGGAGTCTGTGCTGGCTG-3'

(SEQ ID NO: 20)

② 5'-ACAGAGACTGCTGGCCATCTCTGGCCACACA-3'

(SEQ ID NO: 23)

20 The isolated DNA was subcloned using a TA Cloning Kit (Invitrogen Co.) and sequencing was carried out. Figure 76 shows a nucleotide sequence of obtained cDNA fragment, p3H2-17. As a result, it was learned that p3H2-17 is highly homologous to the mouse purinoceptor cDNA fragment, p3H2-17. It is strongly suggested that the human-derived cDNA fragment is a partial nucleotide sequence of human purinoceptor.

25 Based on the nucleotide sequence of p3H2-17 which was sequenced, the following 2 primers were synthesized:

(Nucleotide sequence of synthesized primer)

③ 5'-ACAGCCATCTTCGCTGCCACAGCAT-3'

(SEQ ID NO: 58)

④ 5'-AGACAGTAGCAGCCGACGAGGCGACAAA-3'

(SEQ ID NO: 59)

5 The above synthetic 2 primers were each used in combination with λ gt 11 primers (Takara, Japan; catalogue 3864) for obtaining full-length human purinoceptor cDNA. Thus, using thermally denatured, human placenta-derived λ gt 11 cDNA libraries (Clontech; CLH L1008b), first PCR amplification using a combination of the DNA primer having a nucleotide sequence represented by SEQ ID NO: 20 with λ gt 11 Forward primer, of the DNA primer having a nucleotide sequence represented by SEQ ID NO: 20 with λ gt 11 Reverse primer, of the DNA primer having a nucleotide sequence represented by SEQ ID NO: 23 with λ gt 11 Forward primer, and of the DNA primer having a nucleotide sequence represented by SEQ ID NO: 23 with λ gt 11 Reverse primer was carried out with Ex Taq polymerase (Takara, Japan) (30 cycles; 95°C/30 seconds, 55°C/60 seconds, and 72°C/60 seconds), respectively.

15 Next, by using a 1/50 of the 1st PCR product, second PCR amplification was carried in the same manner as in the first PCR except for using the DNA primer having a nucleotide sequence represented by SEQ ID NO: 58 in place of SEQ ID NO: 20 and the DNA primer having a nucleotide sequence represented by SEQ ID NO: 59 in place of SEQ ID NO: 23 (30 cycles; 95°C/30 seconds, 65°C/60 seconds and 72°C/60 seconds). The amplified product DNA was subcloned using a TA Cloning Kit (Invitrogen Co.) and sequencing was carried out for three clones each of 5' and 3' sides (Figure 77).

25 Based on the amino acid sequence (Figure 77) deduced from the determined nucleotide sequence of human purinoceptor cDNA as shown in Figure 77, hydrophobicity plotting was carried out (Figure 78). As a result, it was learned that the human-derived receptor is a novel seven transmembrane G protein coupled receptor, similarly to the mouse type. It was also learned that the deduced amino acid sequence of human receptor has 87% homology relative to the amino acid sequence of mouse purinoceptor and its amino acid residues are well conserved (Figure 79).

35 Clones free of PCR errors which often occur in a PCR amplification were selected and restriction enzyme regions

comprising overlapping areas were obtained therefrom.

The restriction enzyme regions thus obtained were subjected to construction of plasmid pHAH2-17 having a full-length open reading frame of human purinoceptor cDNA. The plasmid pHAH2-17 is possessed by transformant Escherichia coli JM109/pHAH2-17.

The DNA primers of the present invention allow efficient amplification of DNAs that encode G protein coupled receptor proteins. This makes it possible to efficiently screen for the DNAs coding for G protein coupled receptor proteins and to accomplish the cloning.

The G protein coupled receptor protein of the present invention and their G protein coupled receptor protein-encoding DNA are advantageously useful in:

- ① determining ligands,
- ② obtaining antibodies and an antisera,
- ③ constructing systems for expressing recombinant receptor proteins,
- ④ investigating or developing receptor-binding assay systems and screening for pharmaceutical candidate compounds, by using the above expression system
- ⑤ designing drugs based upon comparisons with ligands and receptors having a structure similar or analogous thereto,
- ⑥ preparing probes and/or PCR primers in gene diagnosis, and
- ⑦ gene manipulating therapy.

In particular, discovering the structure and properties of the G protein coupled receptor will lead to the development of unique pharmaceuticals acting upon these systems.

The practice of the present invention will employ, otherwise indicated, conventional techniques of molecular biology, microbiology, recombinant DNA, pharmacology, immunology, bioscience, and medical technology, which are within the skill of the art. All patents, patent applications, and publications mentioned herein, both supra and infra, are hereby incorporated herein by reference.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

(A) NAME: Takeda Chemical Industries, Ltd.
 (B) STREET: 1-1, Doshomachi 4-chome, Chuo-ku
 (C) CITY: Osaka-shi
 (D) STATE: Osaka
 (E) COUNTRY: Japan
 (F) POSTAL CODE (ZIP): 541

(ii) TITLE OF INVENTION: G Protein Coupled Receptor Protein, Production, And Use Thereof

(iii) NUMBER OF SEQUENCES: 61

(iv) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
 (B) COMPUTER: IBM PC compatible
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA

(iii) FEATURES: N is A, G, C, or T

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

CGTGGSCWTS STGGGCAACN YCCTG 25

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA

(iii) FEATURES: N is A, G, C, or T

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GTNGWRGGC ANCCAGCAGA KGGCAAA 27

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear
 (ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA
 (iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

CTCGCGGCTM TNRGATGCA YCGNTAT 27

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

CATGTRGAG GGAACGAGS AMANBARRAA 30

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

CTGACGCTC TNRSNRYTGA CMGYTAC 27

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

CTGACGCTC TNRSNRYTGA CMGYTAT 27

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

CTCGCGGCTM TNRGATGCA YCGNTAC 27

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

GATGTRTAR GGSNCCAC AGANGRYAA 30

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

- 2 4 1 -

- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:
GATGTGRTAR GGRNCCAC AGANGYCAA 30
- (2) INFORMATION FOR SEQ ID NO: 10:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA
- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:
GYCACCACN WSTCATCCT SWNHCTG 27
- (2) INFORMATION FOR SEQ ID NO: 11:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA
- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:
ASNSANRAG SARTAGANGA NRGRTT 27
- (2) INFORMATION FOR SEQ ID NO: 12:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA
- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:
TGNTSSTWMT NGSNGTKGTN GGNAA 25

- 2 4 2 -

- (2) INFORMATION FOR SEQ ID NO: 13:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA
- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:
AYCKGTAYCK GTCCANKGN ATKGC 25
- (2) INFORMATION FOR SEQ ID NO: 14:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA
- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:
CATKCCSTG GASAGNTAYN TRGC 24
- (2) INFORMATION FOR SEQ ID NO: 15:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA
- (iii) FEATURES: N is inosine
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:
GWWGGSACK CAGCASANGG CRAA 24
- (2) INFORMATION FOR SEQ ID NO: 16:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18

(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: 15th N is A, G, C, or T
6th, 9th, 10th & 12th Ns are inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

ARYTNGCINN TNGCNGAY 18

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: 13th, 15th, 16th & 18th Ns are
each A, G, C, or T
1st, 4th, 6th Ns are inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

NGGNANCCAR CANAMRRRA A 21

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

GCCTSVNTRN SATGASTGTG GANMGNT 27

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(iii) FEATURES: N is inosine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

GAMSTGMYN ANRTGMAAG GNAACCA 27

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

TACTGTGTG AGTCGTGTG CTGACGC 27

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

AGCTTTGCT GCCACAGCA TCCAGCG 27

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Single
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
Synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

CAAGCCAGTA AGGCTATGAA GGCACGACAG 30

- 2 4 6 -

1 5 10 15
 Ser Tyr Val Arg Val Ser Val Lys Leu Arg Asn Arg Val Val Pro Gly 30
 Cys Val Thr Gln Ser Gln Ala Asp Trp Asp Arg Ala Arg Arg Arg Arg 45
 Thr Phe Cys Leu Leu Val Val Val Val Val Val 55

(2) INFORMATION FOR SEQ ID NO: 26:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 370
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

Met Ala Ser Ser Thr Thr Arg Gly Pro Arg Val Ser Asp Leu Phe Ser 15
 Gly Leu Pro Pro Ala Val Thr Thr Pro Ala Asn Gln Ser Ala Glu Ala 30
 Ser Ala Gly Asn Gly Ser Val Ala Gly Ala Asp Ala Pro Ala Val Thr 45
 Pro Phe Gln Ser Leu Gln Leu Val His Gln Leu Lys Gly Leu Ile Val 60
 Leu Leu Tyr Ser Val Val Val Val Val Gly Leu Val Gly Asn Cys Leu 80
 Leu Val Leu Val Ile Ala Arg Val Arg Arg Leu His Asn Val Thr Asn 95
 Phe Leu Ile Gly Asn Leu Ala Leu Ser Asp Val Leu Met Cys Thr Ala 110
 Cys Val Pro Leu Thr Leu Ala Tyr Ala Phe Glu Pro Arg Gly Trp Val 125
 Phe Gly Gly Gly Leu Cys His Leu Val Phe Phe Leu Gln Pro Val Thr 140
 Val Tyr Val Ser Val Phe Thr Leu Thr Thr Ile Ala Val Asp Arg Tyr 160
 Val Val Leu Val His Pro Leu Arg Arg Ile Ser Leu Arg Leu Ser 175
 Ala Tyr Ala Val Leu Ala Ile Trp Ala Leu Ser Ala Val Leu Ala Leu

- 2 4 5 -

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 31
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

ACAGGACCTG CTGGGCCATC CTGGGACAC A 31

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 91
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

Leu Val Leu Val Ile Ala Arg Val Arg Arg Leu His Asn Val Thr Asn 15
 Phe Leu Ile Gly Asn Leu Ala Leu Ser Asp Val Leu Met Cys Thr Ala 30
 Cys Val Pro Leu Thr Leu Ala Tyr Ala Phe Glu Pro Arg Gly Trp Val 45
 Phe Gly Gly Gly Leu Cys His Leu Val Phe Phe Leu Gln Pro Val Thr 60
 Val Tyr Val Ser Val Phe Thr Leu Thr Thr Ile Ala Val Asp Arg Tyr 80
 Val Val Leu Val His Pro Leu Arg Arg Ile 90
 Gly Leu Leu Leu Val Thr Tyr Leu Leu Pro Leu Leu Val Ile Leu Leu

180 185 190
 Pro Ala Ala Val His Thr Tyr His Val Glu Leu Lys Pro His Asp Val
 195 200 205
 Arg Leu Cys Glu Glu Phe Trp Gly Ser Gln Glu Arg Gln Arg Leu
 210 215 220
 Tyr Ala Trp Gly Leu Leu Val Thr Tyr Leu Leu Pro Leu Leu Val
 225 230 235 240
 Ile Leu Leu Ser Tyr Val Arg Val Ser Val Lys Leu Arg Asn Arg Val
 245 250 255
 Val Pro Gly Cys Val Thr Gln Ser Gln Ala Asp Trp Asp Arg Ala Arg
 260 265 270
 Arg Arg Arg Thr Phe Cys Leu Leu Val Val Val Val Phe Ala
 275 280 285
 Val Cys Trp Leu Pro Leu His Val Phe Asn Leu Leu Arg Asp Leu Asp
 290 295 300
 Pro His Ala Ile Asp Pro Tyr Ala Phe Gly Leu Val Gln Leu Leu Cys
 305 310 315 320
 His Trp Leu Ala Met Ser Ser Ala Cys Tyr Asn Pro Phe Ile Tyr Ala
 325 330 335
 Trp Leu His Asp Ser Phe Arg Glu Glu Leu Arg Lys Leu Leu Val Ala
 340 345 350
 Trp Pro Arg Lys Ile Ala Pro His Gly Gln Asn Met Thr Val Ser Val
 355 360
 Val Ile
 370

(2) INFORMATION FOR SEQ ID NO: 27:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 206
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

Leu Val Leu Val Ile Ala Arg Val Arg Arg Leu Tyr Asn Val Thr Asn
 1 5 10 15
 Phe Leu Ile Gly Asn Leu Ala Leu Ser Asp Val Leu Met Cys Thr Ala
 20 25 30
 Cys Val Pro Leu Thr Leu Ala Tyr Ala Phe Glu Pro Arg Gly Trp Val

35 40 45
 Phe Gly Gly Gly Leu Cys His Leu Val Phe Phe Leu Gln Ala Val Thr
 50 55 60
 Val Tyr Val Ser Val Phe Thr Leu Thr Thr Ile Ala Val Asp Arg Tyr
 65 70 75 80
 Val Val Leu Val His Pro Leu Arg Arg Arg Ile Ser Leu Arg Leu Ser
 85 90 95
 Ala Tyr Ala Val Leu Ala Ile Trp Val Leu Ser Ala Val Leu Ala Leu
 100 105 110
 Pro Ala Ala Val His Thr Tyr His Val Glu Leu Lys Pro His Asp Val
 115 120 125
 Arg Leu Cys Glu Glu Phe Trp Gly Ser Gln Glu Arg Gln Arg Leu
 130 135 140
 Tyr Ala Trp Gly Leu Leu Val Thr Tyr Leu Leu Pro Leu Leu Val
 145 150 155 160
 Ile Leu Leu Ser Tyr Ala Arg Val Ser Val Lys Leu Arg Asn Arg Val
 165 170 175
 Val Pro Gly Arg Val Thr Gln Ser Gln Ala Asp Trp Asp Arg Ala Arg
 180 185 190
 Arg Arg Arg Thr Phe Cys Leu Leu Val Val Val Val Val
 195 200 205

(2) INFORMATION FOR SEQ ID NO: 28:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 126
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

Val Val Leu Val His Pro Leu Arg Arg Ile Ser Leu Arg Leu Ser
 1 5 10 15
 Ala Tyr Ala Val Leu Gly Ile Trp Ala Leu Ser Ala Val Leu Ala Leu
 20 25 30
 Pro Ala Ala Val His Thr Tyr His Val Glu Leu Lys Pro His Asp Val
 35 40 45
 Ser Leu Cys Glu Glu Phe Trp Gly Ser Gln Glu Arg Gln Arg Ile
 50 55 60
 Tyr Ala Trp Gly Leu Leu Leu Gly Thr Tyr Leu Leu Pro Leu Leu Ala

65 70 75 80
 Ile Leu Leu Ser Tyr Val Arg Val Ser Val Lys Leu Arg Asn Arg Val 95
 85
 Val Pro Gly Ser Val Thr Gln Ser Gln Ala Asp Trp Asp Arg Ala Arg 110
 100 105
 Arg Arg Arg Thr Phe Cys Leu Leu Val Val Val Val Val 125
 115

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 273
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

CTGTGTCTGG TGATCGCGG GGTGCGCGG CTGCACACG TGACGAACCTT CCTCATCGGC 60
 AACCTGGCCT TGTCGACCT GCTCATGTC ACAGCCTGG TGCCGCTCAC GCTGACCTAT 120
 GCCTTCGAGC CACGCGCTG GGTGTTCGG GCGCGCTGT GCCACCTGGT CTCTCTCTG 180
 CAGCCGGTCA CCGTCTATGT GTCCGTGTTT ACCTCACCA CCATCGCACT GGACCGGTAC 240
 GTCTGCTGG TGCACCGCT GAGCGCGGC ATC 273

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 177
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

GGCTGTCTG TGATCACCTA CCTGCTCCCT CTGCTGGTCA TCCTCTCTG TTAGGTCCG 60
 GTCTCAGTCA AGCTCCGCAA CCGGTGGTG CCGGCTGGG TGACCCAGAG CAGGCGGAC 120
 TGGACCGCG CTCGCGCGCG GCGACCTTC TGCTGTCTG TGSTGTCTG GTGTG 177

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1110
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

ATGGCTCAT CGACCACTCG GGGCCCCAGG GTTCTGACT TATTTTCTGG GTCGCCCGC 60
 GCGGTCAAA CTCGCGCAA CCAGAGCGCA CAGGCTCGG CCGGCAACGG GTCGGTGGCT 120
 GCGCGGAGC CTCAGCGGT CAGCCCTTC CAGAGCTGC AGCTGTGCA TCAGTGAAG 180
 GGGCTGATCG TGCTGCTCTA CAGGTGCTG GTGTGCTGG GGTGTGTGG CACTGCTG 240
 CTGTGTCTGG TGATCGCGG GGTGCGCGG CTGCACAAAG TGACGAACCTT CCTCATCGC 300
 AACCTGGCT TGTCGACCT GCTCATGTC ACCGCTCGG TGCCGCTCAC GTCGCTCAT 360
 GCCTTCGAGC CAGCGGCTG GGTGTTCGG GCGGCTGTG GCCACCTGGT CTCTCTCTG 420
 CAGCGGTCA CCGTCTATGT GTCGGTGTTT AGCTCACCA CCATCGCACT GGACCGGTAC 480
 GTGTGTCTGG TGCACCGCTT GAGCGCGCG ATCTGCTGC GCTCAGCGC CTACGCTGTG 540
 CTGGCCATCT GGGGCTGTC CCGGTGCTG GCGTCCCGG CCGCGGTGCA CACTATCAC 600
 GTGGAGCTCA AGCGCGACA CGTGCCTTC TCGAGGAGT TCTGGGCTC CCAGAGCGG 660
 CAGCGCGAGC TCTACGCTG GGGGCTGCTG CTGGTCACTT ACTGTCTCC TCTGTGCTG 720
 ATCTCTCTGT CTTAGCTCG GGTGTCACTG AGCTCCGCA ACCGCTGGT GCGGGCTGC 780
 GTGACCCAGA CCGAGCGCA CTGGGACCG GCTCGGCGC GCGCACTT CTGCTGTCTG 840
 GTGTGTCTG TGSTGTGTT CCGCTCTGC TGSTGCTGC TGCAGTCTT CAACCTGTG 900
 CCGGACCTCG ACCCCACGC CATCGACCTT TACGCTTTG GGTGTGTGCA GCTGTCTGC 960
 CACTGTCTG CCATGAGTTC GGCCTGTAC AACCCCTTCA TCTAGGCTG GCTGACGAC 1020
 AGCTTCGCG AGGAGCTGC CAACTGTTG GTGCTTGGC CCGCAAGAT AGCCCCCAT 1080
 GGCCAGAATA TGACCGTAG CGTGTCTATC 1110

(2) INFORMATION FOR SEQ ID NO: 32:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 618
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

CTGCTGCTGCG TGATGCGCGG GATGCGCGCG CTGTACAGG TGACGATTT CTTATCGCG 60
AAGCTGGCCT TGTCCGAGT GCTCATGTC ACCGCTGCG TGCCGCTCAC GCTGCGCTAT 120
GCCTTCAGC CAGCGGCGTG GATGTTGGC GCGGCGCTGT GCCACTGCT GTTCTTCTG 180
CAGCGGTCA CGGTCTATGT GTGGGTGTT ACCTACACA CCATCGCAT GAGCCGCTAC 240
GTGCTGCTG TGACCCCGCT GAGCGGCGC ATCTGCTGC GCTTCAGCG CTACGCTG 300
CTGGCATCT GAGTGTCTG CCGGCTGCTG GCGTCCCG CCGCGTGA CACTATAC 360
GTGAGCTCA AGCCGACGA CGTGGGCTC TGCGAGAGT TCTGGGCTC CCAGAGCGC 420
CAGCGGAGC TTAACCTG GGGGCTGCTG CTGTCACT ACTTCTCCC TCTGCTGCT 480
ATCTCTCTGT CTACGCGCG GATGCTAGT AGCTTCGCA ACCGGTGT GCGGCGCGC 540
GTGACCCAGA GCCAGCGCA CTGGAGCGG GCTGCGCGC GCGGCACTT CTGCTTCTG 600
GTGCTGCTG TGCTGCTG 618

(2) INFORMATION FOR SEQ ID NO: 33:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 378
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

GTGCTTCTG TGACCCCGCT ACCTGCGCG ATTTCATCA GCTACAGCG CTACGCGTG 60
CTGGCATCT GAGCTTATC TGAGTGTG GCGCTGCGG CCGGCGTGA CACTATCAT 120
GTGAGCTCA AGCCGACGA CGTGGGCTC TGCGAGAGT TCTGGGCTC GCAGAGCGC 180

CAGCCGACA TCTAGCCTG GGGGCTGCTT CTGGGCACT ATTGCTCCC CTGCTGCC 240
ATCTCTCTGT CTACGCTAG GGTGTGAGT AGCTACAGA ACCGCGTGT GCTGGCAGC 300
GTGACCCAGA GTCAAGCTGA CTGGAGCGA GCGGCTGCC GCGCACTT CTGCTGCTG 360
GTGCTGCTG TGCTGCTG 378

(2) INFORMATION FOR SEQ ID NO: 34:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 70
(B) TYPE: Amino acid
(C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

Val Cys His Val Ile Phe Lys Asn Gln Arg Met His Ser Ala Thr Ser 15
1
Leu Phe Ile Val Asn Leu Ala Val Ala Asp Ile Met Ile Thr Leu Ile 25
20
Asn Thr Pro Phe Thr Leu Val Arg Phe Val Asn Ser Thr Trp Ile Phe 35
30
Gly Lys Gly Met Cys His Val Ser Arg Phe Ala Gln Tyr Cys Ser Leu 45
50
His Val Ser Ala Leu Thr 55
60
His Val Ser Ala Leu Thr 65
70

(2) INFORMATION FOR SEQ ID NO: 35:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 71
(B) TYPE: Amino acid
(C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

Glu Pro Ala Asp Leu Phe Trp Lys Asn Leu Asp Leu Pro Thr Phe Ile 15
1
Leu Leu Asn Ile Leu Pro Leu Leu Ile Ile Ser Val Ala Tyr Val Arg 25
20
Val Thr Lys Lys Leu Trp Leu Cys Asn Met Ile Val Asp Val Thr Thr 35
40
Glu Gln Tyr Phe Ala Leu Arg Pro Lys Lys Lys Thr Ile Lys Met 55
60

Leu Met Leu Val Val Leu
65 70

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 210
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:

GTCTGTCATG TCATCTTCAA GAACGACGGA ATGCACTCGG CCACCAAGCCT CTTTCATGTC 60
AACTGGCAG TTGCGGACAT AATGATCAG CTGCTCAACA CCCCCTTCAC TTGGTTCCG 120
TTTGTGAACA GCACATGGAT ATTTGGGAG GGCATGTGCC ATGTCAGCCG CTTTGCCCAAG 180
TACTGTCAC TGCACGTCTC AGCACTGACA 210

(2) INFORMATION FOR SEQ ID NO: 37:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 213
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

GAGCCAGCTG ACCTCTTCTG GAAGAACCTG GACTTGCCCA CTTTCATCTT GTCACATC 60
CTGCCCTCC TCATCATCTC TGTGGCCTAC GTTGGTGTA CCAAGAACT GTGGCTGTGT 120
AATATGATTG TCGATGTGAC CACAGAGCAG TACTTTGCC TGGCGCCCAA AAAGAAGAG 180
ACCATCAAGA TGTGTATGCT GGTGTAGTC CTC 213

(2) INFORMATION FOR SEQ ID NO: 38:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 115
(B) TYPE: Amino acid
(C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:

Ala Ser Trp His Lys Arg Gly Arg Ala Ala Trp Val Val Cys 15
1 5 10
Gly Val Val Trp Leu Ala Val Thr Ala Gln Cys Leu Pro Thr Ala Val 30
20 25 30
Phe Ala Ala Thr Gly Ile Gln Arg Asn Arg Thr Val Cys Tyr Asp Leu 45
35 40 45
Ser Pro Pro Ile Leu Ser Thr Arg Tyr Leu Pro Tyr Gly Met Ala Leu 60
50 55 60
Thr Val Ile Gly Phe Leu Leu Pro Phe Ile Ala Leu Leu Ala Cys Tyr 80
65 70 75
Cys Arg Met Ala Arg Arg Leu Cys Arg Gln Asp Gly Pro Ala Gly Pro 95
85 90 95
Val Ala Gln Glu Arg Arg Ser Lys Ala Ala Arg Met Ala Val Val Val 110
100 105 110
Ala Ala Val 115

(2) INFORMATION FOR SEQ ID NO: 39:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 328
(B) TYPE: Amino acid
(C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 39:

Met Glu Gln Asp Asn Gly Thr Ile Gln Ala Pro Gly Leu Pro Pro Thr 15
1 5 10
Thr Cys Val Tyr Arg Glu Asp Phe Lys Arg Leu Leu Thr Pro Val 30
20 25 30
Tyr Ser Val Val Leu Val Val Gly Leu Pro Leu Asn Ile Cys Val Ile 45
35 40 45
Ala Gln Ile Cys Ala Ser Arg Arg Thr Leu Thr Arg Ser Ala Val Tyr 60
50 55 60
Thr Leu Asn Leu Ala Leu Ala Asp Leu Met Tyr Ala Cys Ser Leu Pro 80
65 70 75 80

Leu Leu Ile Tyr Asn Tyr Ala Arg Gly Asp His Trp Pro Phe Gly Asp
85 90 95
Leu Ala Cys Arg Phe Val Arg Phe Phe Tyr Ala Asn Leu His Gly
100 105 110
Ser Ile Leu Phe Leu Thr Cys Ile Ser Phe Gln Arg Tyr Leu Gly Ile
115 120 125
Cys His Pro Leu Ala Ser Trp His Lys Arg Gly Arg Arg Ala Ala
130 135 140
Trp Val Val Cys Gly Val Val Trp Leu Ala Val Thr Ala Gln Cys Leu
145 150 155 160
Pro Thr Ala Val Phe Ala Ala Thr Gly Ile Gln Arg Asn Arg Thr Val
165 170 175
Cys Tyr Asp Leu Ser Pro Pro Ile Leu Ser Thr Arg Tyr Leu Pro Tyr
180 185 190
Gly Met Ala Leu Thr Val Ile Gly Phe Leu Leu Pro Phe Ile Ala Leu
195 200 205
Leu Ala Cys Tyr Cys Arg Met Ala Arg Arg Leu Cys Arg Gln Asp Gly
210 215 220
Pro Ala Gly Pro Val Ala Gln Gln Arg Arg Ser Lys Ala Ala Arg Met
225 230 235 240
Ala Val Val Val Ala Ala Val Phe Ala Ile Ser Phe Leu Pro Phe His
245 250 255
Ile Thr Lys Thr Ala Tyr Leu Ala Val Arg Ser Thr Pro Gly Val Ser
260 265 270
Cys Pro Val Leu Gln Thr Phe Ala Ala Ala Tyr Lys Gly Thr Arg Pro
275 280 285
Phe Ala Ser Val Asn Ser Val Leu Asp Pro Ile Leu Phe Tyr Phe Thr
290 295 300
Gln Gln Lys Phe Arg Arg Gln Pro His Asp Leu Leu Gln Arg Leu Thr
305 310 315 320
Ala Lys Trp Gln Arg Gln Arg Val
325

(2) INFORMATION FOR SEQ ID NO: 40:

(1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 345
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(11) MOLECULE TYPE: cDNA
(1x) FEATURE
(C) IDENTIFICATION METHOD: S
(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 40:
GCTTCCTGGC ACAAAGCTGG AGTGGCCGT GCTGCTGGG TAGTGTGG AGTGTGGC 60
CTGGCTGTGA CAGCCAGTGG CCGCCCAAG GAGTCTTTG CTGCCAGAG CATCAGGCG 120
AAGCCACTG TGTGTACGA CCGAGCCCA CCATCTGT CTACTGTCTA CCGCCCTAT 180
GGTATGGCC TCAGGTATC CGGCTTCTTG CTGCCCTTCA TACCTTACT GGTCTGTAT 240
TGTCCATGG CCCGCCGCT GTGTGCCAG GATGGCCAG CAGTCTGT GCCCAAGAG 300
CGGCCAGCA AGCGGCTCG TATGGCTGG GTGTGGCAG CTGTC 345

(2) INFORMATION FOR SEQ ID NO: 41:

(1) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 984
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear
(11) MOLECULE TYPE: cDNA
(1x) FEATURE
(C) IDENTIFICATION METHOD: S
(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 41:

ATGAGCAGG ACAAAGCAG CATCAGGCT CCAAGCTTG CCGCCAGAC CTGGCTTAC 60
CGTAGAGATT TGAGGCACT GGTGCTAAC CCGTATACT CGTGTGCTT GGTGTGGC 120
CTGCCACTGA ACATGTGGT CATGCCCCG ATGTGGCAT CCGCGGAGC CTGACCCGT 180
TCCGCTGTG ACACCTTGA CCGGCACTG GCGACTGA TGTATGCTG TTCACTACC 240
CTACTATCT ATTACTAGC CAGAGGGAG CACTGGCCT TGGAGACTT GGCCTGCCG 300
TTGTAGCT TCCCTTTCTA TGGCAATCTA CATTGACACA TCCGTTCCT CACTGCATT 360
AGCTTCAGC GCTACTGGG CATGTGCCA CCGCTGGCTT CTGGCACA GGTGAGGT 420
CGCCTGCTG CTGGGTAGT GTGTGAGTC GTGTGCTGG CTGTGACAC CAGTGGCTG 480
CCGAGGAGG TCTTGTGTC CAGAGGATC CAGGCAACC GCACTGTGT CTACGACTG 540
AGCCACCCA TCTGTCTAC TCGCTACTG CCGTATGTA TGGCCTTAC GGTATGGC 600
TTCTTGCTG CCTATATAC CTACTGCTT TGTATTTC GATGGCCG CCGCTGTGT 660

GCCCAGGATG GCCCAGCAGG TCCTGTGGCC CAAGAGCGGC GCAGCAAGGC GGCTCGTATG 720
 GCTGTGTGG TGGCAGCTGT CTTTGGCATT AGCTTCTTGC CTTTCCACAT CACCAAGACA 780
 GCCTACTGG CTGTGGCTC CAGCCCGGT GTCTCTTGGC CTGTCTGGA GACCTTGGCT 840
 GCTGCTACA AAGGCACTGG GCCTTGGCC AGTGTCAACA GTGTCTGGA CCCATTCTC 900
 TTCTACTCA CACAACAGAA GTTCCGGCGG CAACCCACG ATCTCTTACA GAGGCTACA 960
 GCCAAGTGGC AGAGGCAGAG AGTC 984

(2) INFORMATION FOR SEQ ID NO: 42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 128
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 42:

Ala Ala Met Ser Val Asp Arg Tyr Val Ala Ile Val His Ser Arg Arg 15
 1
 Ser Ser Ser Leu Arg Val Ser Arg Asn Ala Leu Leu Gly Val Gly Phe 30
 20
 Ile Trp Ala Leu Ser Ile Ala Met Ala Ser Pro Val Ala Tyr His Gln 45
 35
 Arg Leu Phe His Arg Asp Ser Asn Gln Thr Phe Cys Trp Glu Gln Trp 60
 50
 Pro Asn Lys Leu His Lys Lys Ala Tyr Val Val Cys Thr Phe Val Phe 80
 65
 Gly Tyr Leu Leu Pro Leu Leu Leu Ile Cys Phe Cys Tyr Ala Lys Val 95
 85
 Leu Asn His Leu His Lys Lys Leu Lys Asn Met Ser Lys Lys Ser Glu 110
 100
 Ala Ser Lys Lys Lys Thr Ala Gln Thr Val Leu Val Val Val Val 125
 115

(2) INFORMATION FOR SEQ ID NO: 43:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 384
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 43:

GCGCGCATGT CTGTGATCG CTAGTGGCC ATTGTCACT CGCGGGGCTC CTCTCCCTTC 60
 AGGTTGTCCT GCAAGCACT GCTGGGGGTG GCCTTCATCT GGGCGCTGTC CATCGGCATG 120
 GCCTGCGCGG TGGCCTTACCA CCAGCGTCTT TTCCATCGGG ACAGCAACCA GACCTTCTGC 180
 TGGGAGCAGT GGGCCACAA GCTCCACAG AAGCTTAGG TGGTGTGCAC TTCTGCTCTT 240
 GGGTACTTTC TGCCCTTACT GCTCATCTGC TTTTGCTATG CCAAGGTCTT TAATCATCTG 300
 CATAAAAGC TGAATAACAT GTCAAAAAG TCTGAAGCAT CCAAGAAAAA GACTGCACAG 360
 ACCGTCTCTG TGGTGGTTGT AGTA 384

(2) INFORMATION FOR SEQ ID NO: 44:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 71
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 44:

Val Leu Trp Phe Phe Gly Phe Ser Ile Lys Arg Thr Pro Phe Ser Val 15
 1
 Tyr Phe Leu His Leu Ala Ser Ala Asp Gly Ala Tyr Leu Phe Ser Lys 30
 20
 Ala Val Phe Ser Leu Leu Asn Ala Gly Gly Phe Leu Gly Thr Phe Ala 45
 35
 His Tyr Val Arg Ser Val Ala Arg Val Leu Gly Leu Cys Ala Phe Val 60
 50
 Ala Gly Val Ser Leu Leu Pro 70

(2) INFORMATION FOR SEQ ID NO: 45:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 215
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(c) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 45:

GTGCTCTGTGTTCTTGGGCTTCTCCATCAGAGACACCCCTTCTCCGCTCTA CTTCCTGCAC 60
 CTGGCCAGCG CCGAGCGCGC CTACCTCTTC AGCAAGGCGG TGTCTCCCT GCTGAAGCGC 120
 GCGGCGTTCC TGGGACACCTT CGCCCACTAT GTGGCAGCG TGGCCCGGGT GCTGGGCGCTC 180
 TGGCGCTTCG TGGCGGGCGT GAGCTCTCTG CCGGC 215

(2) INFORMATION FOR SEQ ID NO: 46:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 348

(B) TYPE: Amino acid

(C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 46:

Met Glu Leu Ala Met Val Asn Leu Ser Glu Gly Asn Gly Ser Asp Pro 15
 1 5 10
 Glu Pro Pro Ala Pro Glu Ser Arg Pro Leu Phe Gly Ile Gly Val Glu 30
 20 25 30
 Asn Phe Ile Thr Leu Val Val Phe Gly Leu Ile Phe Ala Met Gly Val 45
 35 40 45
 Leu Gly Asn Ser Leu Val Ile Thr Val Leu Ala Arg Ser Lys Pro Gly 60
 50 55 60
 Lys Pro Arg Ser Thr Thr Asn Leu Phe Ile Leu Asn Leu Ser Ile Ala 80
 65 70 75
 Asp Leu Ala Tyr Leu Leu Phe Cys Ile Pro Phe Gln Ala Thr Val Tyr 95
 85 90 95
 Ala Leu Pro Thr Trp Val Leu Gly Ala Phe Ile Cys Lys Phe Ile His 110
 100 105 110
 Tyr Phe Phe Thr Val Ser Met Leu Val Ser Ile Phe Thr Leu Ala Ala 125
 115 120 125
 Met Ser Val Asp Arg Tyr Val Ala Ile Val His Ser Arg Arg Ser Ser 140
 130 135 140
 Ser Leu Arg Val Ser Arg Asn Ala Leu Leu Gly Val Gly Phe Ile Trp 160
 145 150 155
 Ala Leu Ser Ile Ala Met Ala Ser Pro Val Ala Tyr His Gln Arg Leu 175
 165 170 175

Phe His Arg Asp Ser Asn Gln Thr Phe Cys Trp Glu Gln Trp Pro Asn 180
 185 190

Lys Leu His Lys Lys Ala Tyr Val Val Cys Thr Phe Val Phe Gly Tyr 195
 200 205

Leu Leu Pro Leu Leu Ile Cys Phe Cys Tyr Ala Lys Val Leu Asn 210
 215 220

His Leu His Lys Lys Leu Lys Asn Met Ser Lys Lys Ser Glu Ala Ser 225
 230 235 240

Lys Lys Lys Thr Ala Gln Thr Val Leu Val Val Val Val Phe Gly 245
 250 255

Ile Ser Trp Leu Pro His His Val Val Val His Leu Trp Ala Glu Phe Gly 260
 265 270

Ala Phe Pro Leu Thr Pro Ala Ser Phe Phe Phe Arg Ile Thr Ala His 275
 280 285

Cys Leu Ala Tyr Ser Asn Ser Ser Val Asn Pro Ile Ile Tyr Ala Phe 290
 295 300

Leu Ser Glu Asn Phe Arg Lys Ala Tyr Lys Gln Val Phe Lys Cys His 305
 310 315 320

Val Cys Asp Glu Ser Pro Arg Ser Glu Thr Lys Glu Asn Lys Ser Arg 325
 330 335

Met Asp Thr Pro Pro Ser Thr Asn Cys Thr His Val 340
 345

(2) INFORMATION FOR SEQ ID NO: 47:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1044

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Double

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(c) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 47:

ATGAAGCTCG CTATGCTGAA CCTAGTGAA GGAATGGGA GCGACCCAGA GCCCGCAGCC 60
 CCGAGTCCA GCGCGCTCTT CGGCAATGGC GTGGAGAACT TCATTACGCT GGTAGTCTT 120
 GGCTGATTT TCGGATGGG GGTCTGGGC AACACCTCG TGATACCGT GCTGGCGCC 180
 AGCAACGAG GCAACCCCG CAGACCAAC AACCTGTTA TCTCAATCT GAGCATGGA 240

- 2 6 2 -

- 2 6 1 -

GACCTGGCCT ACCTGCTCTT CTGCATCCCT TTTCAGGCCA CCGTGTATGC ACTGCCCCACC 300
 TGGGTGCTGG GGGCTTTCAT CTGCAGTTT ATACACTACT TCTTCACCGT GTCCATGCTG 360
 GTGACCATCT TCACCTGGC CGCATGTCT GTGGATCCCT ACGTGGCCAT TGTGCACTCG 420
 CGGGCTCCT CCTCCCTCAG GGTGTCCCGC AACGCACTGC TGGGGCTGGG CTTTCATCTGG 480
 GCGGTGTCCA TGGCCATGCC CTGCGCGGTG GCCTACCACC AGCGTCTTTT CCATGGGAC 540
 AGCAACCAGA CCTTCTGCTG GGAGCAGTGG CCCAACAGC TCCACAAGAA GGCTTACGTG 600
 GTGTGCACTT TCGTCTTGG GTACCTTCTG CCTTACTGC TCATCTGCTT TTGCTATGCC 660
 AAGGTCTTAA ATCATCTGCA TAAAGCTG TAAACATGT CAAAAAGTC TGAAGCATCC 720
 AAGAAAAGA CTGCACAGAC CGTCTGGTG GTGCTGTAG TATTGGCAT ATCTGGCTG 780
 CCCCATCAG TCGTCCACT CTGGGCTGAG TTGGAGCCT TCCCACTGAC GCCAGCTTCC 840
 TTCTTCTTCA GAATCACCGC CCATTGGCTG GCATACAGCA ACTCTCAGT GAACCCCATC 900
 ATATATGCC TTTCTCTAGA AAATTCGG AGGGGTACA AGCAAGTTT CAAGTGTAT 960
 GTTTGGGATG AATCTCACG CAGTGAACCT AAGGAAACA AGAGCCGGAT GGACACCCCG 1020
 CCATCCACCA ACTGCACCCA CGTG 1044

(2) INFORMATION FOR SEQ ID NO: 48:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 125
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 48:

Leu Leu Thr Leu His Pro Val Trp Ser Gln Lys His Arg Thr Ser His 15
 1
 Trp Ala Ser Arg Val Val Leu Gly Val Trp Leu Ser Ala Thr Ala Phe 20
 25
 Ser Val Pro Tyr Leu Val Phe Arg Glu Thr Tyr Asp Asp Arg Lys Gly 35
 40
 Arg Val Thr Cys Arg Asn Tyr Ala Val Ser Thr Asp Trp Glu Ser 50
 55
 Lys Glu Met Gln Thr Val Arg Gln Trp Ile His Ala Thr Cys Phe Ile 65
 70
 Ser Arg Phe Ile Leu Gly Phe Leu Leu Pro Phe Leu Val Ile Gly Phe 75
 80

85

90

95

Cys Tyr Glu Arg Val Ala Arg Lys Met Lys Glu Arg Gly Leu Phe Lys 110
 100
 Ser Ser Lys Pro Phe Lys Val Thr Met Thr Ala Val Ile 125
 115

(2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 377
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

CTTCTCACCC TTCACCACT GTGGTCCCA AAGCACCAG CCTCACACTG GCCTTCCAGA 60
 GTGCTCTGG GAGTCTGGCT CTCTGCCACT GCCTTCAGCG TGCCCTATTT GGTTTCAGG 120
 GAGACATATG ATGACCGTAA AGGAAGAGTG ACCTGCAGAA ATAACTACGC TGTGTCCACT 180
 GACTGGGAAA GCAAGAGAT GCAACACGTA AGACAATGGA TTCATGCCAC CTGTTTCATC 240
 AGCGCTTCA TACTGGGCTT CCTTCTGCTT TTCTTAGTCA TTGGCTTTTG TTATGAAGA 300
 GTAGCCCGCA AGATGAAGA GAGGGGCTC TTAAATCCA GCAACCCCTT CAAAGTCACG 360
 ATGACTGCTG TTATCTC 377

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 119
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 50:

Phe Lys Ile Val Lys Pro Leu Ser Thr Ser Phe Ile Gln Ser Val Asn 15
 1
 Tyr Ser Lys Leu Val Ser Leu Val Val Trp Leu Leu Met Leu Leu Leu 30
 20
 Ala Val Pro Asn Val Ile Leu Thr Asn Gln Arg Val Lys Asp Val Thr 45
 35

- 2 6 3 -

Gln Ile Lys Cys Met Glu Leu Lys Asn Glu Leu Gly Arg Gln Trp His
50 55 60
Lys Ala Ser Asn Tyr Ile Phe Val Gly Ile Phe Trp Leu Val Phe Leu
65 70 75 80
Leu Leu Ile Ile Phe Tyr Thr Ala Ile Thr Arg Lys Ile Phe Lys Ser
85 90 95
His Leu Lys Ser Arg Lys Asn Ser Ile Ser Val Lys Lys Lys Ser Ser
100 105 110
Arg Asn Ile Phe Ser Ile Val
115

(2) INFORMATION FOR SEQ ID NO: 51:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 357
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(11) MOLECULE TYPE: CDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 51:

TTCAAGATTG TGAAGCCCT TTCAAGCTC TTCATTCAGT CTGTGACTA CAGAAATC 60
GTCTGGCTGG TGGTGTGTT GCTCATCTC CTCCTGCGG TCCCAAGT CATTCAC 120
AACCAAGAG TTAAGACGT GACGACATA AATGCAATG AACTAATAA CGAATGGGC 180
CGCCAGTGGC ACAAGCCCT AAATACATC TTGTGGGCA TTCTGTGCT TGTGTCTT 240
TTGTAATCA TTTTACAC TGTATCAC AGAATAAT TTAAGTCCA CCGAATTC 300
AGAAAGATT CCATCTCGT CAATAAGAA TTAAGCCGA ACATCTCAG CATCTG 357

(2) INFORMATION FOR SEQ ID NO: 52:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 252
(B) TYPE: Amino acid
(C) TOPOLOGY: Linear

(11) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 52:

Val Asp Leu Leu Ala Ala Leu Thr Leu Met Pro Leu Ala Met Leu Ser
1 5 10 15

- 2 6 4 -

Ser Ser Ala Leu Phe Asp His Ala Leu Phe Gly Glu Val Ala Cys Arg
20 25 30
Leu Tyr Leu Phe Leu Ser Val Cys Phe Val Ser Leu Ala Ile Leu Ser
35 40 45
Val Ser Ala Ile Asn Val Glu Arg Tyr Tyr Val Val His Pro Met
50 55 60
Arg Tyr Glu Val Arg Met Lys Leu Gly Leu Val Ala Ser Val Leu Val
65 70 75 80
Gly Val Trp Val Lys Ala Leu Ala Met Ala Ser Val Pro Val Leu Gly
85 90 95
Arg Val Ser Trp Glu Glu Gly Pro Pro Ser Val Pro Pro Gly Cys Ser
100 105 110
Leu Gln Trp Ser His Ser Ala Tyr Cys Gln Leu Phe Val Val Phe
115 120 125
Ala Val Leu Tyr Phe Leu Leu Pro Leu Leu Ile Leu Val Val Tyr
130 135 140
Cys Ser Met Phe Arg Val Ala Arg Val Ala Ala Met Gln His Gly Pro
145 150 155 160
Leu Pro Thr Trp Met Glu Thr Pro Arg Gln Arg Ser Glu Ser Leu Ser
165 170 175
Ser Arg Ser Thr Met Val Thr Ser Ser Gly Ala Pro Gln Thr Thr Pro
180 185 190
His Arg Thr Phe Gly Gly Lys Ala Ala Val Val Leu Leu Ala Val
195 200 205
Gly Gly Gln Phe Leu Leu Cys Trp Leu Pro Tyr Phe Ser Phe His Leu
210 215 220
Tyr Val Ala Leu Ser Ala Gln Pro Ile Ala Ala Gly Gln Val Glu Asn
225 230 235 240
Val Val Thr Trp Ile Gly Tyr Phe Cys Phe Thr Ser
245 250

(2) INFORMATION FOR SEQ ID NO: 53:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 756
(B) TYPE: Nucleic acid
(C) STRANDEDNESS: Double
(D) TOPOLOGY: Linear

(11) MOLECULE TYPE: CDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

GTGGACCTGC TGGCTGCCCT GACCTCATG CCTCTGGCCA TGCTCTCCAG CTCGGCCCTC 60
TTTGACCAAG CCTCTTTGG GGAGTGGCC TCGCCCTCT ACTTGTCTT GAGCGTTCG 120
TTTGTACGC TGGCCATCCT CTCGGTGTCC GCCATCAATG TGGAGCGCTA CTATTATGTG 180
GTCCACCCCA TCGGTATGA GTTGGCATG AACTGGGGC TGGTGGCTC TGTGCTGTG 240
GGCGTGTGG TGAAGGCCCT GGCATGGCT TCTGTGCCAG TGTGGGAAG GGTGTCTCTGG 300
GAGGAGGCC CTCCTCAGTGT CCCCCAGGC TGTTCACCTCC AATGAGCCA CAGTGCCTAC 360
TGCCAGCTTT TCGTGTGTG CTTCGGGTC CTCTACTTC TGTGCCCTT GCTCTCTATC 420
CTTGTGTCT ACTGCAGCAT GTTCCGGGTG GCTGTGTGG CTGCCATGCA GCACGGCCG 480
CTGCCACGT GGATGGAGAC GCCCCGGCAA CGTCCGAGT CTCTCAGCAG CCCTCCACT 540
ATGTCACCA GCTCGGGGC CCCCAGACC ACCCTCACC GGAGCTTTGG CGGAGGGAAG 600
GCACAGTGG TCTCTCTGC TGTGGAGGA CAGTTCTCTG TCTGTGTGT GCTCTACTTC 660
TCCTTCACC TCTATGTGC CTGAGCGCT CAGCCCATG CAGCCGGGCA GTTGGAGAAC 720
GTGTGACCT GGATGGCTA CTCTGCTTC ACCTCC 756

(2) INFORMATION FOR SEQ ID NO: 54:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 263

(B) TYPE: Amino acid

(C) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

Ala Asp Val Leu Val Thr Ala Ile Cys Leu Pro Ala Ser Leu Leu Val 15
1

Asp Ile Thr Glu Ser Trp Leu Phe Gly His Ala Leu Cys Lys Val Ile 30
20

Pro Tyr Leu Gln Ala Val Ser Val Val Val Leu Thr Leu Ser 45
35

Ser Ile Ala Leu Asp Arg Tyr Tyr Ala Ile Cys His Pro Leu Leu Phe 60
50

Lys Ser Thr Ala Arg Arg Ala Arg Gly Ser Ile Leu Gly Ile Trp Ala 80
65 70 75

Val Ser Leu Ala Val Met Val Pro Gln Ala Ala Val Met Glu Cys Ser 90
85

Ser Val Leu Pro Glu Leu Ala Asn Arg Thr Arg Leu Leu Ser Val Cys 110
100

Asp Glu Arg Trp Ala Asp Asp Leu Tyr Pro Lys Ile Tyr His Ser Cys 125
115

Phe Phe Ile Val Thr Tyr Leu Ala Pro Leu Gly Leu Met Ala Met Ala 140
130

Tyr Phe Gln Ile Phe Arg Lys Leu Trp Gly Arg Gln Ile Pro Gly Thr 160
145

Thr Ser Ala Leu Val Arg Asn Trp Lys Arg Pro Ser Asp Gln Leu Asp 175
165

Asp Gln Gly Gln Leu Ser Ser Glu Pro Gln Pro Arg Ala Arg Ala 190
180

Phe Leu Ala Glu Val Lys Gln Met Arg Ala Arg Arg Lys Thr Ala Lys 205
195

Met Leu Met Val Leu Leu Val Phe Ala Leu Cys Tyr Leu Pro Ile 220
210

Ser Val Leu Asn Val Leu Lys Arg Val Phe Gly Met Phe Arg Gln Ala 240
235

Ser Asp Arg Glu Ala Ile Tyr Ala Cys Phe Thr Phe Ser His Trp Leu 255
245

Val Tyr Ala Asn Ser Ala Ala 260

(2) INFORMATION FOR SEQ ID NO: 55:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 789

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Double

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 55:

GCCGATGTC TGGTACAGC CATCTGCCTG CCGGCCAGTC TGCTGTAGA CATCAGGNA 60

TCCTGCTCT TTGGCCATGC CCTCTGCAAG GTCATCCCT ATCTACAGGC CGTGTCCGTG 120

- 2 6 7 -

TCAGTGTG TGCTGACTCT CAGCTCCATC GCCCTGACG GCTGTACGC CATCTGCAC 180
 CCGCTGTGT TCAAGACAC TGCCCGGCGC GCCCGGCGCT CCATCTCGG CATCTGGCG 240
 GTGTGCTGG CTGTGATGCT GCTTCAGGCT GCTGTACAG AGGTACACG CGTGCTGCC 300
 GAGCTGGCA ACCGACCGC CTCCTGCTT GTCTGTGAT AGGCTGGGC AGACACTTG 360
 TACCCACAG TCTACACAG CTGCTCTTC ATTGTACCT ACCTGACCC ACTGGGCTTC 420
 ATGCCATGG CTAATTCCA GATCTCCG AGCTCTGG GCCGCAGAT CCGCGGACC 480
 ACCTGGGCCC TGGTGGGCA CTGTAAGGG CCTCAGAC ACCTGACGA CAGAGCCAG 540
 GGCTGAGCT CAGAGCCCA GCGCGGGGCC CGGCGCTTC TGGCCGAGT GAAACAGATG 600
 CGAGCCCGA GAAAGACGC CAGATCTCT AGGTGTGTC TGCTGTCTT CGCTCTCTG 660
 TACCTGCCA TCACTCTCT CAGCTCTCT AGAGGCTCT TCGGATGTT CCGCAGACC 720
 AGCAGCCAG AGGCACTTA CGCTGCTTC ACCTCTCCC ACTGCTGCT GTACGCCAC 780
 AGCGCGGCC 789

(2) INFORMATION FOR SEQ ID NO: 56:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 328
 (B) TYPE: Amino acid
 (C) TOPOLOGY: Linear

(11) MOLECULE TYPE: Peptide

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 56:

Met Glu Trp Asp Asn Gly Thr Gly Gln Ala Leu Gly Leu Pro Pro Thr 1
 5 10 15
 Thr Cys Val Tyr Arg Glu Asn Phe Lys Gln Leu Leu Leu Pro Pro Val 20
 25 30
 Tyr Ser Ala Val Leu Ala Ala Gly Leu Pro Leu Asn Ile Cys Val Ile 35
 40 45
 Thr Gln Ile Cys Thr Ser Arg Arg Ala Leu Thr Arg Thr Ala Val Tyr 50
 55 60
 Thr Leu Asn Leu Ala Leu Ala Asp Leu Leu Tyr Ala Cys Ser Leu Pro 65
 70 75 80
 Leu Leu Ile Tyr Asn Tyr Ala Gln Gly Asp His Trp Pro Phe Gly Asp 85
 90 95
 Phe Ala Cys Arg Leu Val Arg Phe Phe Tyr Ala Asn Leu His Gly 100
 105 110

- 2 6 8 -

Ser Ile Leu Phe Leu Thr Cys Ile Ser Phe Gln Arg Tyr Leu Gly Ile 115
 120 125
 Cys His Pro Leu Ala Pro Trp His Lys Arg Gly Gly Arg Arg Ala Ala 130
 135 140
 Trp Leu Val Cys Val Thr Val Trp Leu Ala Val Thr Thr Gln Cys Leu 145
 150 155 160
 Pro Thr Ala Ile Phe Ala Ala Thr Gly Ile Gln Arg Asn Arg Thr Val 165
 170 175
 Cys Tyr Asp Leu Ser Pro Pro Ala Leu Ala Thr His Tyr Met Pro Tyr 180
 185 190
 Gly Met Ala Leu Thr Val Ile Gly Phe Leu Leu Pro Phe Ala Ala Leu 195
 200 205
 Leu Ala Cys Tyr Cys Leu Leu Ala Cys Arg Leu Cys Arg Gln Asp Gly 210
 215 220
 Pro Ala Glu Pro Val Ala Gln Glu Arg Arg Gly Lys Ala Ala Arg Met 225
 230 235 240
 Ala Val Val Val Ala Ala Ala Phe Ala Ile Ser Phe Leu Pro Phe His 245
 250 255
 Ile Thr Lys Thr Ala Tyr Leu Ala Val Gly Ser Thr Pro Gly Val Pro 260
 265 270
 Cys Thr Val Leu Glu Ala Phe Ala Ala Tyr Lys Gly Thr Arg Pro 275
 280 285
 Phe Ala Ser Ala Asn Ser Val Leu Asp Rto Ile Leu Phe Tyr Phe Thr 290
 295 300
 Gln Lys Lys Phe Arg Arg Arg Pro His Glu Leu Leu Gln Lys Leu Thr 305
 310 315 320
 Ala Lys Trp Gln Arg Gln Gly Arg 325

(2) INFORMATION FOR SEQ ID NO: 57:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 984
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Double
 (D) TOPOLOGY: Linear

(11) MOLECULE TYPE: cDNA

(ix) FEATURE

(C) IDENTIFICATION METHOD: S

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 57:

ATGGAAATGGG ACAATGGCAC AGGCCAGGCT CTGGGCTTGC CACCCACCAC CTGTGTCTAC 60
 CCGGAGAACT TCAGCAACT GCTGCTCCA CTTGTGTATT CGCGGTGCT GGGGGTGGC 120
 CTGCGCTGA ACATCTGTCT CATTACCCAG ATCTGCAGCT CCGCGGGGC CTGAGACCCGC 180
 AGGCGGTGT ACACCTTAA CTTGTCTTG GCTGACCTGC TATATGCCCTG CTCCCTGCCC 240
 CTGCTCATCT ACAACTATGC CCAAGGTGAT CACTGGCCCT TTGGGGACTT CGCCTGCCGC 300
 CTGTGCCCT TCTCTTCTA TGCCACCTG CAGGCGACA TCTCTTCTT CACTTGCAATC 360
 AGCTTCAGC GTTACCTGG CATCTGCCAC CGCTGGCCC CTTGGCACA ACCTGGGGGC 420
 CGCGGGCTG CTGGGTAGT GTGTGTAACC GTGTGGCTGG CGGTGACAAC CCAAGTGCCCTG 480
 CCCACAGCA TCTTCGCTGC CACAGGCATC CAGGTAACC GCATGTCTG CTATGACCTC 540
 AGCCGCGCTG CCTTGCCAC CCACTATATG CCTATGGCA TGGCTCTCAC TGTCTATCGC 600
 TTCTGTCTG CTTTGTCTGC CTGTGGGCC TGTACTGTC TCTGGGCTG CGGCTGTGTC 660
 CGCCAGGATG GCGCGGCAGA GCTGTGGCC CAGGAGCGG GTGGGAAGG GGGCCGCATG 720
 GCCGTGTGG TGGCTGTGTC CTTTGGCATC AGCTTCTGTC CTTTTCACAT CACCAAGACA 780
 GCCTACTGG CAGTGGGCTC GAGCGGGGC GTCCCTTGCA CTGTATTGGA GGCCTTTGCA 840
 GCGGCTTACA AAGGACCGG GCGTTTGGC AGTGCCAACA GCGTGGCTGA CCCCATCCTC 900
 TTCTACTTCA CCCAGAGAA GTTCCGCGG CGACCACATG AGCTCCTACA GAAACTCACA 960
 GCCAAATGGC AGAGCGAGG TCGC 984

(2) INFORMATION FOR SEQ ID NO: 58:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 26
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

- (ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 58:

ACAGCCATCT TCGCTGCCAC AGGCAT 26

(2) INFORMATION FOR SEQ ID NO: 59:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 29
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single

- (D) TOPOLOGY: Linear
 (ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 59:
 AGACAGTAGC AGCCAGCAG GGCAGCAA 29

(2) INFORMATION FOR SEQ ID NO: 60:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 27
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

- (ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA

- (iii) FEATURES: N is inosine

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 60:

CTGTGCGYSA TYGCNTYGA YMGSTAC 27

(2) INFORMATION FOR SEQ ID NO: 61:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 29
 (B) TYPE: Nucleic acid
 (C) STRANDEDNESS: Single
 (D) TOPOLOGY: Linear

- (ii) MOLECULE TYPE: Other nucleic acid
 Synthetic DNA

- (iii) FEATURES: N is inosine

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 61:

AKGWAGWAGG GCAGCCAGCA GANSRYGAA 29

CLAIMS

1. A DNA which comprises a nucleotide sequence represented by a SEQ ID NO selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 19.

2. A method for amplifying a DNA coding for a G protein coupled receptor protein by polymerase chain reaction techniques, which comprises:

(i) carrying out a polymerase chain reaction in the presence of a mixture of

① a DNA coding for a G protein coupled receptor protein, said DNA being capable of acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, DNA primers comprising a nucleotide

sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19; or

(ii) carrying out a polymerase chain reaction in the presence of a mixture of

① a DNA coding for G protein coupled receptor protein, said DNA being capable of acting as a template,

② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and

③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13.

3. A method for screening a DNA library for a DNA coding for a G protein coupled receptor protein, which comprises:

(i) carrying out a polymerase chain reaction in the presence of a mixture of

① said DNA library,

② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 3, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 5, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 6, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 7, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 10, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 14, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 16 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 18, and

③ at least one DNA primer selected from the group

consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 2, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 4, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 8, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 9, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 11, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 15, DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 17 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 19, under conditions to amplify selectively a template DNA coding for the G protein coupled receptor protein, contained in the DNA library and selecting said DNA; or

(ii) carrying out a polymerase chain reaction in the presence of a mixture of

- ① said DNA library
 - ② at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 1 and DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 12, and
 - ③ at least one DNA primer selected from the group consisting of DNA primers comprising a nucleotide sequence represented by SEQ ID NO: 13,
- under conditions to amplify selectively a DNA coding for the G protein coupled receptor protein, contained in the DNA library and selecting said DNA.

4. A DNA coding for a G protein coupled receptor protein or a fragment thereof, which is obtained by the method according to claim 2 to 3.

5. A G protein coupled receptor protein encoded by the DNA according to claim 4, a peptide segment or fragment thereof or a salt thereof.

6. A G protein coupled receptor protein comprising an amino acid sequence selected from the group consisting of

an amino acid sequence represented by SEQ ID NO: 24, an amino acid sequence represented by SEQ ID NO: 25, an amino acid sequence represented by SEQ ID NO: 26, an amino acid sequence represented by SEQ ID NO: 27, an amino acid sequence represented by SEQ ID NO: 28, an amino acid sequence represented by SEQ ID NO: 34, an amino acid sequence represented by SEQ ID NO: 35, an amino acid sequence represented by SEQ ID NO: 38, an amino acid sequence represented by SEQ ID NO: 39, an amino acid sequence represented by SEQ ID NO: 56, and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 38, SEQ ID NO: 39, or SEQ ID NO: 56; a peptide segment (or fragment) thereof, a modified peptide derivative thereof or a salt thereof.

7. The G protein coupled receptor protein according to claim 6, comprising an amino acid sequence selected from the group consisting of an amino acid sequence represented by SEQ ID NO: 38, an amino acid sequence represented by SEQ ID NO: 39, an amino acid sequence represented by SEQ ID NO: 56 and substantial equivalents to the amino acid sequence represented by SEQ ID NO: 38, SEQ ID NO: 39, or SEQ ID NO: 56.

8. The G protein coupled receptor protein according to claims 6 or 7, wherein said receptor is a purinoceptor.

9. The G protein coupled receptor protein according to any of claims 6 to 8, wherein an agonist to said receptor is useful as an immunomodulator or an antitumor agent, in addition it is useful in therapeutically or prophylactically treating hypertension, diabetes or cystic fibrosis, and an antagonist to said receptor is useful as a hypotensive agent, an analgesic, or an agent for therapeutically or prophylactically treating incontinence of urine.

10. A DNA which comprises a nucleotide sequence coding for a G protein coupled receptor protein of claim 6.

11. The DNA according to claim 10 comprising a nucleotide sequence coding for the G protein coupled receptor

Protein according to claim 7.

12. The DNA according to claim 11 comprising a nucleotide sequence represented by SEQ ID NO: 40, SEQ ID NO: 41, or SEQ ID NO: 57.

13. A transformant containing a vector comprising the DNA according to claim 4 or 10; or an expression system comprising an open reading frame (ORF) of DNA derived from a G protein coupled receptor protein DNA according to claim 4 or 10, wherein the ORF is operably linked to a control sequence compatible with a desired host cell.

14. A method for determining a ligand to the G protein coupled receptor protein according to any of claims 5 to 8, which comprises contacting

(i) at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of claims 5 to 8, peptide segments or salts thereof, and mixtures thereof, with

(ii) at least one compound to be tested and determining whether said compound to be tested bound to the component of (i).

15. A screening method for a compound capable of inhibiting the binding of a G protein coupled receptor protein according to any of claims 5 to 8 with a ligand, which comprises carrying out a comparison between:

(i) at least one case where said ligand is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of claims 5 to 8, peptide segments or salts thereof, and mixtures thereof,

and
(ii) at least one case where said ligand together with a compound to be tested is contacted with at least one component selected from the group consisting of G protein coupled receptor proteins or salts thereof according to any of claims 5 to 8, peptide segments or salts thereof, and mixtures thereof.

16. A compound which is determined through the method according to claim 15 or a salt thereof.

17. The compound according to claim 16, which is an agonist or antagonist to a G protein coupled receptor protein according to any of claims 5 to 8.

18. A ligand to a G protein coupled receptor protein according to any of claims 5 to 8, which is determined through the method according to claim 14.

1 / 7 9

2 / 7 9

FIGURE 1

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

Primer HS-1 CGTGGCCATCCTGGGCAACACCCCTG

G C G G C T

G

T

HTRH CCTGGGCATTGTAGGCAACATCATGGT
 HUMRANTES CATTGGCCTGGTTGAAACATCCTGGT
 HSLRIA CCTGGGCGTGATCGGCAACGTCCTGGT
 HUMSONAT GGTGGGGCTGGTGGGCAACGCCCTGGT
 RNU02083 AGTGGGCCCTCTTCGGAAACTTCCTGGT
 U00442 GGTGGGCTTAGTGGGCAATTCCCTGGT
 HUMNBR CGTGGGCTTGCTGGGCAACATCATGCT
 HSHM4 GGTACCATCATCGGCAACATCCTGGT
 RATAADRE01 CTTTGGCCATCGTGGGCAACATCTTGGT
 HUMSTR3X GGTGGGCTGCTGGGTAACCTCGCTGGT
 HUMCSAAR GGTGGGAGTGTGGGCAATGCCCTGGT
 HUMRDC1A CATGGGCATGATTGCCAACTCCGTGGT
 HUMOPIODRE CGTGGCGGTGCTCGGCAACCTCGTGGT
 RATA2BAR GCTGGCAGTGGCGGGCAACGTCGTGGT

FIGURE 2

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 3' SIDE PRIMER

Complementary Sequence TTTGCCATCTGCTGGATGCCCCACAAC
to Primer HS-2 C C TTT C G T

HUMGIR TTTGCCCTCTGCTGGTTCCTCTCAAC
 HUMBOMB3S TTTGCCCTCTGCTGGTTCCTCAATCAC
 S46950 TTTGCCCTCTGCTGGTGGCCCTACAC
 MUSGPCR TTTGCCCTGCTGCTGGTGGCCCTCTCAAC
 S43387 TTTGCCCTTTTATGGATGCCCTACAGG
 RATNEURA TTTGCCATCTGCTGGTGGCCCTATCAC
 RATALARA TTTGCCCTCAGCTGGTGGCGCTGCAT
 HUMOPIODRE TTTGCCATCTGCTGGTGGCCCTATCAC
 HUMNEKAR TTTGCCATCTGCTGGTGGCCCTACCAC
 RATADENREC TTTGCTTGTGCTGGTGGCTTTGTCC
 HUMSR11A TTTGTATCTGCTGGATGCCCTTTCTAC
 S8637154 TTTGTATCTGCTGGTGGCCCTATCAT
 RUCGPCR TTTGCCGCTGCTGGATGGCTTTTACC
 HUMSTR42 TTTGTGCTGCTGCTGGATGCCCTTTCTAC
 RATGNRHA TTTGCACACTGGTCGAAGCCAGACAAA

3 / 7 9

FIGURE 3

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

Primer 3A CTGACCGCTCTTACCTGACCGATAC
 T T GG GT A C
 G

Primer 3B CTGACCGCTCTTACCTGACCGATAT
 T T GG GT A C
 G

L11064 CTACCATGATGAGCGTGGACCGCTAC
 L11065 TTGACCATGATGAGTGTGACCGCTAC
 D16349 CTCTGACCATGAGCGTGGACCGCTAC
 X69676 CTGATGCTCGTGAATATGACCGCTAC
 M35328 CTACGGCACTGTCACTGACAGGTAC
 M73482 CTCACTGCCCTCAGCGCGACAGGTAC
 M73481 CTCACGGCGCTCTGGCAGACAGATAC
 L08893 TTACAATTCTCAGCGCTGACAGATAC
 X62933 ATGACCGCATCGCCGCTGACAGGTAC
 X62934 ATGACAACCTGTGGCTTTGACAGATAC
 J05189 ATGACAGCCATTGCACTGGACAGGTAT
 M60786 CTCTGGCTCTCAGTGTGGACAGGTAC
 L04672 CTCACTGCCCTCAGCATTTGACCGCTAC
 X61496 TTGCTGGCTATCACTGTGGACCGCTAC
 X59249 TTGCTGGCCATTGCTGTAGACCGATAC
 L09249 CTCACCTGCCCTCAGCATTTGACCGCTAC
 P30731 CTGACAGCTATGCACTGGACCGCTAC
 M31210 CTCTGCGCATCGCCATTGACCGCTAT
 U03642 CTCACGGCGCTCAGCTTGGACCGCTAC

4 / 7 9

FIGURE 4

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

Primer 3C CTCGCCCTATTAGCATGGACCGTTAC
 G CC G T T

Primer 3D CTCGCCCTATTAGCATGGACCGTTAT
 G CC G T T

L32840 ATTACCTGATGAGTGTGATAGGTAC
 X64052 CTCACGTGTCTCAGCATCGATCGCTAC
 M90065 CTCACGTGTCTCAGCATCGATCGCTAC
 M91464 CTCACGTGTCTCAGCATTTGATCGATAC
 M88096 CTGGTAGCCATCTCTGGAGAGATAT
 M99418 CTCGTGGCCATAGCCCTGGAGCGATAC
 L04473 CTCGTGGCCATCGCACTGGAGCGGTAC
 M73969 CTGGCTGCACTCAGTGTGGACCGTTAC
 X65858 TTGGCTGCACTCAGTGTGGACCGTTAC
 S46665 CTGGCTACCATTTAGTGGACCGCTTC
 M60626 ATGCGCTCATTTGCTCTGGACCGCTGT

5 / 7 9

6 / 7 9

FIGURE 5

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 3' SIDE PRIMER

Complementary Sequence TTTACCTCTGTTGGTGGCCCTACCATC
to Primer 6A GT TC T T

Complementary Sequence TTCACCTCTGTTGGTGGCCCTACCATC
to Primer 6B GT TC T T

L11064	TTGGTGGTGTGCTGGGCGCCCATCCACATC
L11065	TTTCATCATCTGTTGGACCCCATTCACATC
D16349	TTTATCGTCTGCTGGACCCCATCCACATC
X69876	TTTGCTGCTGTGGGTGGCTTTCCAGATC
M35328	TTTGCCCTCTGCTGGCTCCCCAACCATGTC
M73482	TTCACTCTCTGTTGGTTCCAAACCATC
M73481	TTGGCCCTCTGCTGGCTCCCCAATCATGTC
L08893	TTTGCCCTCTGCTGGTTGCCAATCACCTC
X62933	TTTGCCATCTGCTGGCTGCCCTACCACTC
X62934	TTGGCCATCTGCTGGCTGCCCTTCCACATC
J05189	TTTGCCATCTGCTGGCTGCCCTATCAGTG
M60786	TTGGCCCTGCTGGTTCCTCTTCACTTA
L04672	TTTGTCATCTGCTGGCTGCCCTACCAAGTG
X61496	TTTGCCGCTGCTGGATGCTTTTACCCTC
X59249	TTTGCCCTGCTGGCTGCCCTTGTCCATC
L09249	TTTGCCATCTGCTGGCTGCCCTACCAAGTG
P30731	TTTGCCCTCTGCTGGTTCCCTCTCAACTGC
M31210	TTTCATGCTGCTGGGACCGCTCTTCATC
U03642	TTTGCCCTGCTGGATGCTGCCCTACCACTG

FIGURE 6

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 3' SIDE PRIMER

Complementary Sequence TTTTCTTTTGGCTGGTTCCCTACCATG
to Primer 6C CC T G C T T

L32840	TTTCATCATTTGCTGGCTTCCCTTCCATGTT
X64052	TTCTTCTTTTCTGGGTCCCCACCAATA
M90065	TTCTTCTTTTCTGGGTCCCCACCAATA
M91464	TTTTTCTTTTCTGGATTCCCCACCAATA
M88096	TTCTCTGCTGGATGCCCATCTTCAGC
M99418	TTCTCTGCTGGCTGCCAGTGTACAGC
L04473	TTTTTCTGTTGGTTGCCAGTTTATAGT
M73969	TTCTGCTTTGCTGGCTGCCCTACACCTG
X65858	TTCTGCTTTGCTGGCTGCCCTACACCTG
S46665	TTCTTTATCTTCTGGCTGCCCTATCAGGTG
M60626	TTTTTCTCTGCTGGTCCCATATCAGGTG

7 / 7 9

8 / 7 9

FIGURE 7

FIGURE 8

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

COMPLEMENTARY OLIGODEOXYNUCLEOTIDE SEQUENCE TO 3' SIDE PRIMER

Primer T2A

GTCACCACTGTTCACTCTCACTG
C AC GT T A

Complementary Sequence
to Primer T7A T T C C C G G

HUNGALAREC ACCACCACTGTTCACTCTCACTG
RATADRALB CCCACCACTACTTATCGTCAACCTG
HUMADRB1 ACCACCACTGTTCACTCTCAACCTG
RAB1L8RSB GTACCGAGCTACCTGCTGAACCTG
HUMOPIDRE GTACCAACTCTTCTCTGTGAACCTG
BTSR GTACCAACTACTTCACTGTCAACCTG
HUMSRI2A ATCACCACATTTACATCTCAACCTG
HUMSTR3Y GTACCAAGCTTACATCTCAACCTG
HUNGARE GTACCAAGCTTCTCTCTCACTG
HUMCKAR GTACCAACATCTTCTCTCTCTCTG
HUMSHTR CCTTCACTACCTGATGCTGCTG
HUMD1B ATGACCAAGCTTTCATGCTGCTG
HUMSH1E CCTGCCACTACCTAATCTGTTCTG
HUMD4C CCCACCACTCTTCACTGCTGAACCTG
HUMSRI1A GCCACCACTATTCTGATGCTCACTT
RATADRALA GTACCACTATTTCATGCTGAACCTG
S57565 CTGACCAATGCTTCACTGCTGCTG

HUNGALAREC AATCTATCATTTATGCACTTCTCT
RATADRALB AACCCATGCTATGCTTCCGATC
PIGA2R AATCTCTCTTTATGCTTCTG666
RAT5HTRC AACCTATCATCTACCCGCTCTTATG
S58541 AACCCATCATTTATGCTTAACTGCT
HUMGRPR AACCTTTGCTCTACCTGCTGAGC
MUSGRBOM AACCTTTGCTCTTATCTGCTGAGC
RRV1A1IR AACCTCTGCTCTACGCTTCTG666
HUMADRB1 AACCCATCATCTACTGCGGAGCCCC
HSHM4 AACCCGATGCTATGCTGCTGCAAC
HUNGARE AACCCGCTGCTACTGCTGCTGATGCAAC
RATCKAR AACCCATCATCTATGCTTCAATGAAC
S59749 AATCCATGCTCTACACCTTGGCTG6C
HUMS128A AACCCGCTCTACG6CTTCTCTG
RNGPROCR AACCCATCTCTACG6CTTCTCTCC
MUS5RI1A AACCCATCTCTACG6CTTCTCTG
HUM1A1ADR AACCCGCTCATCTACCCGCTTCTGAGC
S66181 AACCCGCTCTCTACG6CTTCTCTGAGC
HUMSTR3Y AACCCATCTCTTATGCTTCTCTCC

9 / 7 9

1 0 / 7 9

FIGURE 9

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

Primer TM1-A2	TGTTGGTTATIGGIGTTGTIGGIAA CC GC C G
MUSB82R	TGGTGGTGGTGGTGGTGGTGGCAA
BTSKR	TGGTGTGGTGGCTGTGATGGCAA
BOVEETBR	TGTTCTGTCTGGCATCATCGGAAA
HUMNEUYREC	TGATCATTTCTTGGTGTCTCTGGAAA
MMSUBKREC	TGGTGTGGTGGCTGTAAACAGGCAA
HUMPE2R	TGTTCAITCTTCGGGGTGGTGGCAA
HUMPIR	TGTTCTGGCCGGTGTGGTGGCAA
HSU11053	TGTTCTGTGGGCTTGGTGGGCAA
RRHC3RA	TGGTATCTCTGGCTGTGGTGGGAA
HUMMR	TGTTATCTCTGGCCGTGGTGGGAA
MUSGRPBOX	TCATCGTGTAGGTCTTATTGGCAA
RATCHOLREC	TCCTTCTGATGAGTGTGGCGGAAA
RATCCKAR	TATTCCTTCTCAGTGTGGCGGGGAA

FIGURE 10

COMPLEMENTARY OLIGODEOXYNUCLEOTIDE SEQUENCE TO 3' SIDE PRIMER

Complementary Sequence to Primer TM3-B2	A T A C G A G
HUMCCKR	GCCATCGCACTGGAGCGGTACAG
HUMCCKBGR	GCCATCGCACTGGAGCGGTACAG
MMGMC5R	GCCATTGCGGTGGACAGGTACA
HUMV2R	GCCATGACGCTGGACCGCCACCG
RATNEURA	GCCATTGCAGTGGACAGGTA
DOGGSTRN	GCCATGCCCTGGAGCGGATACAG
RAT5HT5A	GCAATAGCTTTGGACCGCTACTGGT
MUSALP2ADA	GCCATTAGTCTGGACCGCTACTGGT
HUMADORA1X	GCAATTGCTGTGGACCGCTACC
HUMOPIDORE	GCCATCGCGGTGGACAGATACA
MUSGRPBOX	GCACTGTGAGTGTACAGGTACAAA
RATCCKAR	GCCATCTCTCTGGAGAGATATGG
HSTRHREC	GCCTTACCATTGAGAGGTACATA

1 3 / 7 9

1 4 / 7 9

FIGURE 13

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

Primer TM2F18	ARYTIGCIITIGCGAY
---------------	------------------

HUMTSX	AACCTGGCCTTGGCGAT
HUMNEKAR	AATCTGGCGCTGGCTGAC
HUMFNLP	AACCTGGCGCTGGCTGAC
HUMINTLEU8	AACCTAGCCTTGGCCGAC
HUM1AADR	AACCTGGCGCTGGCCGAC
HUM1L8RA	AACCTGGCCTTGGCCGAC
HSDD2	AGCCTCGCAGTGGCCGAC
HUMANTIR	AATTAGCACTGGCTGAC
HUMSOMAT	AACCTGGCGTAGCCGAC
HUMEL4REC	AGCTTGGCTGTGGCTGAT
HSTRHREC	AGCCTGGCAGTAGCTGAT
HSU07882	AACCTGGCCTTAGCCGAT

(R = A or G, Y = C or T, N = A, C, G or T, and
I = Inosine)

FIGURE 14

COMPLEMENTARY OLIGODEOXYNUCLEOTIDE SEQUENCE TO 3' SIDE PRIMER

Complementary Sequence to Primer TM6R21	TTYNNNTNGYTGGIITICCI
--	----------------------

HSBAR	TTCACCCCTCTGCTGGCTGCCC
HUMNEKAR	TTTGCCATCTGCTGGCTGCCC
HUMETN1R	TTTGCTCTTTGCTGGTTCCCT
HUMHISH2R	TTTCATCATCTGCTGGTTTCCC
HUM1AADR	TTGCTGCTCTGCTGGTTCCCT
HUM1L8RA	TTCTGCTTTGCTGGCTGCCC
HUMNMBR	TTTCATCTCTGTTGGTTTCCT
HUMNKIRX	TTGCCATCTGCTGGCTGCCC
HUMSUBPRA	TTGCCATCTGCTGGCTGCCC
HUMSH1DA	TTTATCATCTGCTGGCTGCCC
HUMPFPR2A	TTCTTCATCTGTTGGTTTCCC
HSDD2	TTTCATCATCTGCTGGCTGCCC
HUMNEUYREC	TTTGCACTCTGCTGGCTCCCT
HUM2XXX	TTTGCCCTCTGCTGGCTGCCC
HUMBK2A	TTTCATCATCTGCTGGCTGCCC
HUMFNLPX	TTCTTCATCTGTTGGTTTCCC
HUMSSTR3X	TTGCTGCTCTGCTGGATGCCC
HUMCCCKR	TTTTTCTGTGTTGGTTGCCA
HSNEURA	TTTGCTGCTGCTGGCTGCCC

(Y = C or T, N = A, C, G or T, and I = Inosine)

1 5 / 7 9

FIGURE 15

OLIGODEOXYNUCLEOTIDE SEQUENCE FOR 5' SIDE PRIMER

Primer S3A
 GCCTGTTAIGATGAGTGTGGAAGIT
 C G C TC C

HUMGALAREC CCCTGCCCCGATGTCCGTGACCCCT
 S70057 GCCTGTGGCCATCGCACTGGAGCCGT
 S67127 ACCCTGGCCCTCTTAAGTGTGACAGGT
 S44866 GTCTAATGCTCTGAGTATTGACAGAT
 HUMC5AAR TCCTGGCCACATCAGCGCCGACCGCT
 HUMANTIR TACTCAGGTGCTCAGCATTGATCGAT
 HUMBK2A TCCTGATGCTGGTGAAGCATGACCCCT
 HSNEURA ACGTGGCCAGCCTGAGTGTGGAGCCCT
 HUMGRPR CACTCAGCGCCCTCTGGCAGACAGAT
 HUMFSRS GCCTGACAGTCATGAGCGTGGACCCGT
 HUMIL8RA TGTGGCCCTGCACTCAGTGTGGAACCGTT
 HUMNEKAR CCAATGACCGCCATTGCTGCCGACAGGT

1 6 / 7 9

FIGURE 16

COMPLEMENTARY OLIGODEOXYNUCLEOTIDE SEQUENCE TO 3' SIDE PRIMER

Complementary Sequence TGGTTCCTACCACTTTATCATC
 to Primer S6A T T GG GT

HUMGALAREC TGGCTGCCGACACCATCATCCATCTC
 S70057 TGGTTGCCAGTTTATAGTGCCACACAG
 S67127 TGGTTCCTCTTCAATTTAAGCCGTATA
 S44866 TGGCTTCCCTTTCACCTCAGCAGGATT
 HUMC5AAR TGGTGGCCCTACCAAGTGAACGGGATA
 HUMANTIR TGGATTCCCCCAATAATTCACTTTT
 HUMBK2A TGGCTGCCCTTCAGATCAGACCTTC
 HSNEURA TGGACTCGGTTCTCTAATGACTTTTAC
 HUMGRPR TGGCTCCCAATCATGTCATCTACCTG
 HUMFSRS TGGCTGCCCTTCTTACCGGTCAACATC
 HUMIL8RA TGGCTGCCCTTACAACTGCTCTGCTG
 HUMNEKAR TGGCTGCCCTTACCACCTCTACTCATC

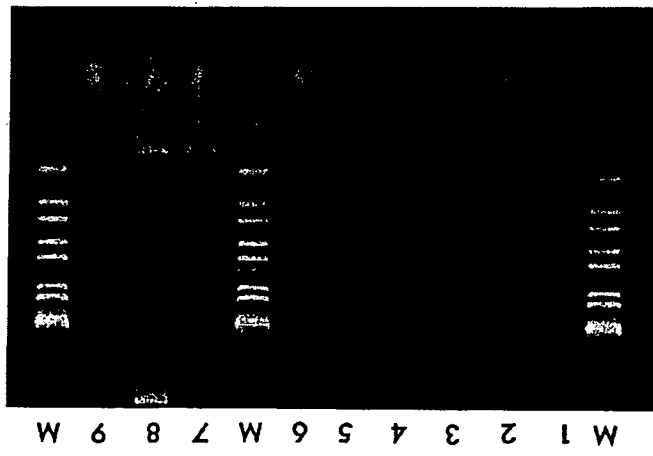


FIGURE 17

WO 96/05302

1 7 / 7 9

PCT/JP95/01599

A58-T7-2	GTGGCATGTGGGCAACCCCTGATCTTCGTCATCCTTCGCTACGC	X:::	GTGGGCTGTGGGCAACCCCTGATCTTCGTCATCCTTCGCTACGC	285	295	305	315	325	100
A58-T7-2	CAAGATGAAGAGCGCTAACCAACATCTACCTGCTCAACCTGCGTAGCCG	335	345	355	365	375	110	150	190
A58-T7-2	ACGAGCTCTTCATGCTTGAGCGCTTCCGCTTCGCTGCGTCCGCTTG	160	170	180	190	200	210	240	280
HUMSOMAT	ACGAGCTCTTCATGCTTGAGCGCTTCCGCTTCGCTGCGTCCGCTTG	385	395	405	415	425	435	445	455
A58-T7-2	CGCACTGCGCCCTTCGCGCTTCGCTGCTGCTGCTGCTGCTGCTGCTG	465	475	485	495	505	515	525	535
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	545	555	565	575	585	595	605	615
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	625	635	645	655	665	675	685	695
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	705	715	725	735	745	755	765	775
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	785	795	805	815	825	835	845	855
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	865	875	885	895	905	915	925	935
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	945	955	965	975	985	995	1005	1015
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1025	1035	1045	1055	1065	1075	1085	1095
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1105	1115	1125	1135	1145	1155	1165	1175
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1185	1195	1205	1215	1225	1235	1245	1255
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1265	1275	1285	1295	1305	1315	1325	1335
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1345	1355	1365	1375	1385	1395	1405	1415
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1425	1435	1445	1455	1465	1475	1485	1495
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1505	1515	1525	1535	1545	1555	1565	1575
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1585	1595	1605	1615	1625	1635	1645	1655
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1665	1675	1685	1695	1705	1715	1725	1735
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1745	1755	1765	1775	1785	1795	1805	1815
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1825	1835	1845	1855	1865	1875	1885	1895
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1905	1915	1925	1935	1945	1955	1965	1975
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	1985	1995	2005	2015	2025	2035	2045	2055
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2065	2075	2085	2095	2105	2115	2125	2135
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2145	2155	2165	2175	2185	2195	2205	2215
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2225	2235	2245	2255	2265	2275	2285	2295
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2305	2315	2325	2335	2345	2355	2365	2375
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2385	2395	2405	2415	2425	2435	2445	2455
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2465	2475	2485	2495	2505	2515	2525	2535
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2545	2555	2565	2575	2585	2595	2605	2615
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2625	2635	2645	2655	2665	2675	2685	2695
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2705	2715	2725	2735	2745	2755	2765	2775
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2785	2795	2805	2815	2825	2835	2845	2855
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2865	2875	2885	2895	2905	2915	2925	2935
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	2945	2955	2965	2975	2985	2995	3005	3015
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3025	3035	3045	3055	3065	3075	3085	3095
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3105	3115	3125	3135	3145	3155	3165	3175
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3185	3195	3205	3215	3225	3235	3245	3255
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3265	3275	3285	3295	3305	3315	3325	3335
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3345	3355	3365	3375	3385	3395	3405	3415
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3425	3435	3445	3455	3465	3475	3485	3495
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3505	3515	3525	3535	3545	3555	3565	3575
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3585	3595	3605	3615	3625	3635	3645	3655
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3665	3675	3685	3695	3705	3715	3725	3735
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3745	3755	3765	3775	3785	3795	3805	3815
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3825	3835	3845	3855	3865	3875	3885	3895
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3905	3915	3925	3935	3945	3955	3965	3975
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	3985	3995	4005	4015	4025	4035	4045	4055
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4065	4075	4085	4095	4105	4115	4125	4135
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4145	4155	4165	4175	4185	4195	4205	4215
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4225	4235	4245	4255	4265	4275	4285	4295
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4305	4315	4325	4335	4345	4355	4365	4375
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4385	4395	4405	4415	4425	4435	4445	4455
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4465	4475	4485	4495	4505	4515	4525	4535
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4545	4555	4565	4575	4585	4595	4605	4615
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4625	4635	4645	4655	4665	4675	4685	4695
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4705	4715	4725	4735	4745	4755	4765	4775
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4785	4795	4805	4815	4825	4835	4845	4855
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4865	4875	4885	4895	4905	4915	4925	4935
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	4945	4955	4965	4975	4985	4995	5005	5015
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5025	5035	5045	5055	5065	5075	5085	5095
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5105	5115	5125	5135	5145	5155	5165	5175
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5185	5195	5205	5215	5225	5235	5245	5255
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5265	5275	5285	5295	5305	5315	5325	5335
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5345	5355	5365	5375	5385	5395	5405	5415
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5425	5435	5445	5455	5465	5475	5485	5495
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5505	5515	5525	5535	5545	5555	5565	5575
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5585	5595	5605	5615	5625	5635	5645	5655
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5665	5675	5685	5695	5705	5715	5725	5735
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5745	5755	5765	5775	5785	5795	5805	5815
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5825	5835	5845	5855	5865	5875	5885	5895
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5905	5915	5925	5935	5945	5955	5965	5975
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	5985	5995	6005	6015	6025	6035	6045	6055
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6065	6075	6085	6095	6105	6115	6125	6135
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6145	6155	6165	6175	6185	6195	6205	6215
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6225	6235	6245	6255	6265	6275	6285	6295
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6305	6315	6325	6335	6345	6355	6365	6375
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6385	6395	6405	6415	6425	6435	6445	6455
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6465	6475	6485	6495	6505	6515	6525	6535
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6545	6555	6565	6575	6585	6595	6605	6615
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6625	6635	6645	6655	6665	6675	6685	6695
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6705	6715	6725	6735	6745	6755	6765	6775
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6785	6795	6805	6815	6825	6835	6845	6855
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6865	6875	6885	6895	6905	6915	6925	6935
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	6945	6955	6965	6975	6985	6995	7005	7015
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7025	7035	7045	7055	7065	7075	7085	7095
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7105	7115	7125	7135	7145	7155	7165	7175
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7185	7195	7205	7215	7225	7235	7245	7255
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7265	7275	7285	7295	7305	7315	7325	7335
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7345	7355	7365	7375	7385	7395	7405	7415
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7425	7435	7445	7455	7465	7475	7485	7495
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7505	7515	7525	7535	7545	7555	7565	7575
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7585	7595	7605	7615	7625	7635	7645	7655
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7665	7675	7685	7695	7705	7715	7725	7735
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7745	7755	7765	7775	7785	7795	7805	7815
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7825	7835	7845	7855	7865	7875	7885	7895
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7905	7915	7925	7935	7945	7955	7965	7975
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	7985	7995	8005	8015	8025	8035	8045	8055
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8065	8075	8085	8095	8105	8115	8125	8135
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8145	8155	8165	8175	8185	8195	8205	8215
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8225	8235	8245	8255	8265	8275	8285	8295
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8305	8315	8325	8335	8345	8355	8365	8375
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8385	8395	8405	8415	8425	8435	8445	8455
HUMSOMAT	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8465	8475	8485	8495	8505	8515	8525	8535
A58-T7-2	CGCCCTCAACATGTTCAACGAGTCTTCTGTCATCAGCGT	8545	8555	8565	8575	8585	8595	8605	

21/79

FIGURE 21

[illegible]

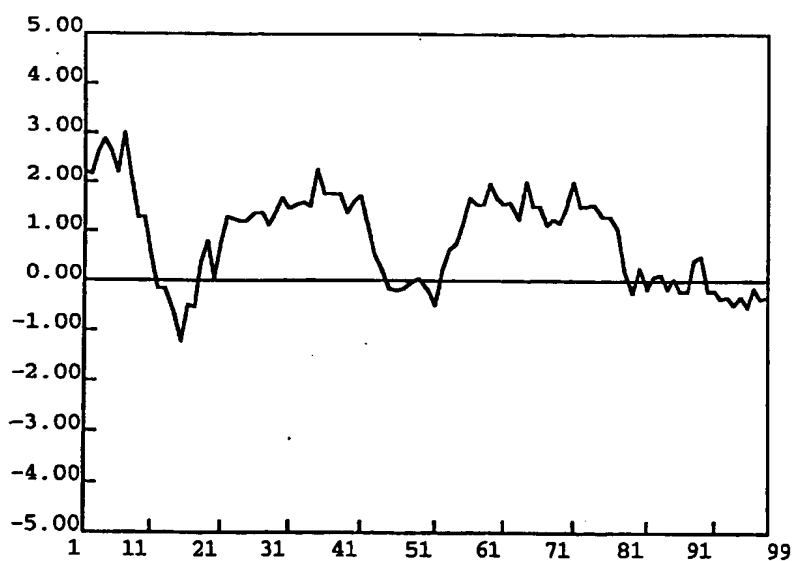
FIGURE 23

5'	GGC	CTG	CTG	CTG	GTC	ACC	TAC	CTG	CTC	CCT	CTG	CTG	GTC	ATC	CTC	CTG	TCT	TAC	54
	Gly	Leu	Leu	Leu	Val	Thr	Tyr	Leu	Leu	Pro	Leu	Leu	Val	Ile	Leu	Leu	Ser	Tyr	
	GTC	CGG	GTG	TCA	GTG	AAG	CTC	CGC	AAC	CCG	GTG	GTG	CCG	GTC	TGC	GTG	ACC	CAG	108
	Val	Arg	Val	Ser	Val	Lys	Leu	Arg	Asn	Pro	Val	Val	Pro	Val	Cys	Val	Thr	Gln	
	AGC	CAG	GCC	GAC	TGG	GAC	CGC	GCT	CGG	CGC	CGG	CGC	ACC	TTC	TGC	TTG	CTG	GTG	162
	Ser	Gln	Ala	Asp	Trp	Asp	Arg	Ala	Arg	Arg	Arg	Arg	Thr	Phe	Cys	Leu	Leu	Val	
	GTG	GTC	GTG	GTG	GTG	TTT	GCC	ATC	TGC	TGG	TTG	CCT	TAC	TAC					
	Val	Val	Val	Val	Val	Phe	Ala	Ile	Cys	Trp	Leu	Pro	Tyr	Tyr					

23 / 79

PCT/JP95/01599

FIGURE 24



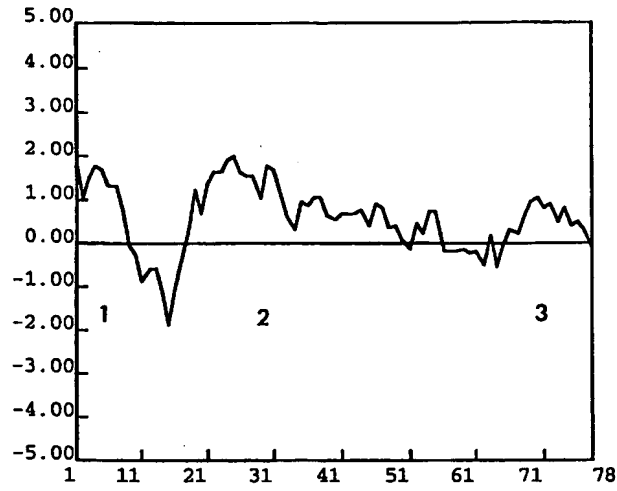
24 / 79

WO 96/053102

PCT/JP95/01599

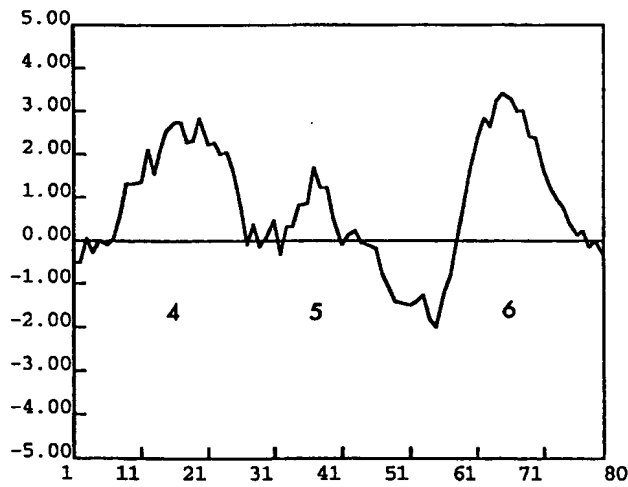
3 1 / 7 9

FIGURE 31



3 2 / 7 9

FIGURE 32



3 5 / 7 9

3 6 / 7 9

FIGURE 35

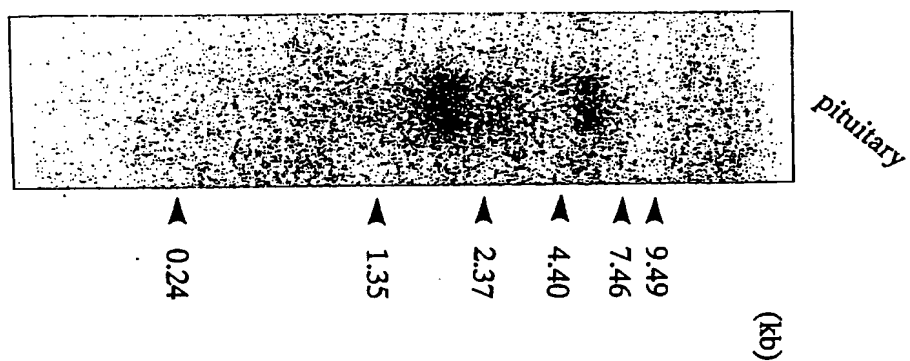
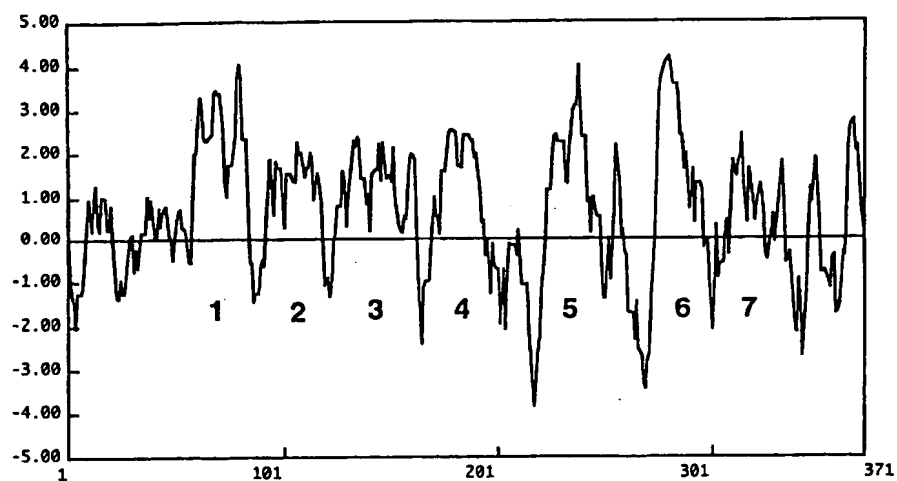


FIGURE 36



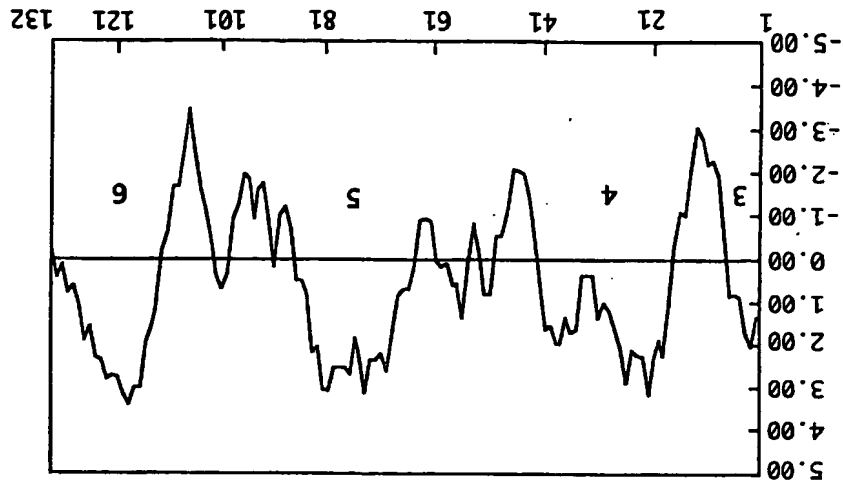
3 7 / 7 9

FIGURE 37

5' GTC GGC CTG GTG GGC ATC CTG GCT TTC TGG CAC CCG GGT GGA GGT GGC GGT
 Val Gly Leu Val Val Gly Asn Ile Leu Ala Ser Trp His Lys Arg Gly Gly Arg Arg
 9 18 27 36 45 54
 63 72 81 90 99 108
 GCT GCT TGG GTA GTG GTC GTC GTC GTC GTC GTC GTC GTC GTC GTC GTC
 Ala Ala Trp Val Val Cys Gly Val Val Trp Leu Ala Val Thr Ala Gln Cys Leu
 117 126 135 144 153 162
 CCC AGC GCA GTC TTT GCT GCT GCA ACA GTC CAG CAC CAC CAC CAC CAC CAC
 Pro Thr Ala Val Phe Ala Ala Thr Gly Ile Gln Arg Asn Arg Thr Val Cys Tyr
 171 180 189 198 207 216
 GAC CTG AGC CCA CCC ATC CTG TCT ACT GTC TAC CAC CAC CAC CAC CAC CAC
 Asp Leu Ser Pro Ile Leu Ser Thr Arg Tyr Leu Pro Tyr Gly Met Ala Leu
 225 234 243 252 261 270
 ACG GTC ATC GTC TTC TGG CTG CCC TTC AUA CCC TTA CAG GCT TGT TAT TAT CAC
 Thr Val Ile Gly Phe Leu Leu Pro Phe Ile Ala Leu Ala Cys Tyr Cys Arg
 279 288 297 306 315 324
 ATG GGC CAC CAC CTG TGT CAC CAG GAT GGC CCA GCA GGT GGT GTC GGC CAA GAG
 Met Ala Arg Arg Leu Cys Arg Gln Asp Gly Pro Ala Gly Pro Val Ala Gln Glu
 333 342 351 360 369 378
 CCG CCG ACG ACG GCT GCT GCT GCT GTC GTC GTC GTC GTC GTC GTC GTC
 Arg Arg Ser Lys Ala Ala Arg Met Ala Val Val Val Ala Val Phe Ala Leu
 387 396
 TGC TGC CTG CTT CTC TAC 3'
 Cys Trp Leu Pro Leu Tyr

3 8 / 7 9

FIGURE 38



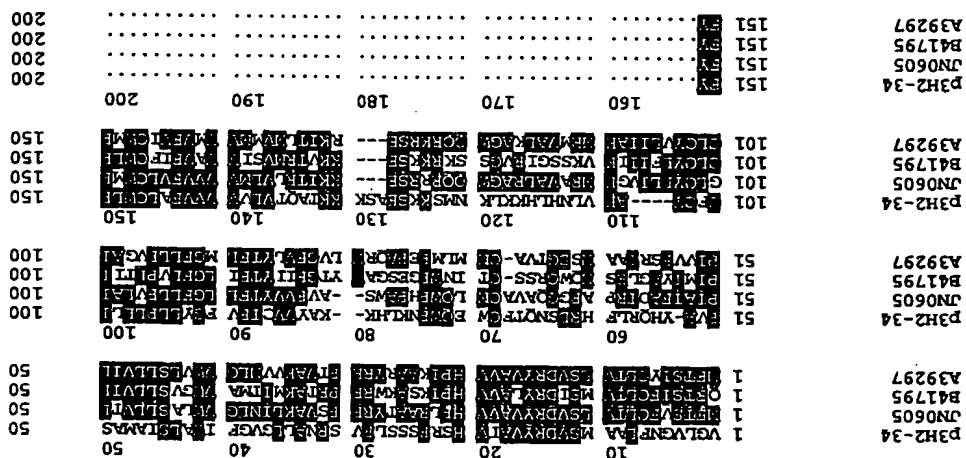


FIGURE 42

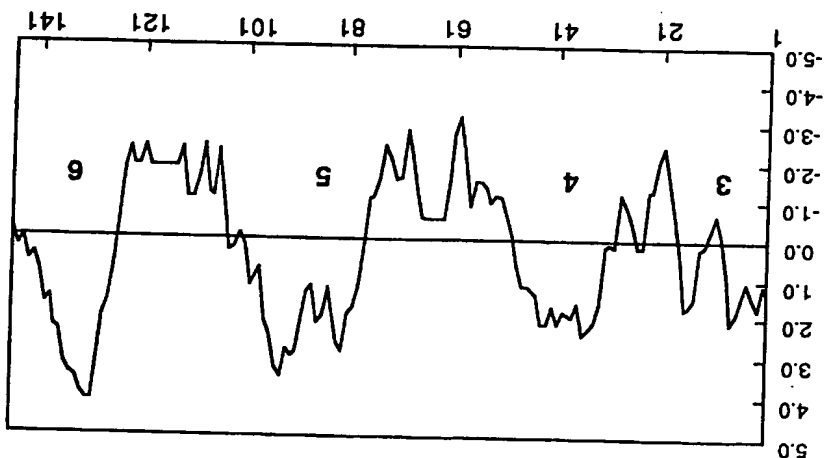


FIGURE 41

FIGURE 43

5'	GTG	GCC	ATG	GTG	GCC	AAC	GTC	CTG	GTG	CTC	TGG	TTC	TTC	GCC	TTC	TCC	ATC	AAG	
	Val	Gly	Met	Val	Gly	Asn	Val	Leu	Val	Leu	Trp	Phe	Phe	Gly	Phe	Ser	Ile	Lys	
	AGG	ACC	CCC	TTC	TCC	GTC	TAC	TTC	CTG	CAC	CTG	GCC	AGC	GCC	GAC	GCC	GCC	TAC	
	Arg	Thr	Pro	Phe	Ser	Val	Tyr	Phe	Leu	His	Leu	Ala	Ser	Ala	Asp	Gly	Ala	Tyr	
	CTC	TTC	AGC	AAG	GCC	GTG	TTC	TCC	CTG	CTG	AAC	GCC	GCC	GCC	TTC	CTG	GCC	ACC	
	Leu	Phe	Ser	Lys	Ala	Val	Phe	Ser	Leu	Leu	Asn	Ala	Gly	Gly	Phe	Leu	Gly	Thr	
	TTC	GCC	CAC	TAT	GTG	CGC	AGC	GTG	GCC	CGG	GTG	CTG	GGG	CTC	TGC	GCC	TTC	GTG	
	Phe	Ala	His	Tyr	Val	Arg	Ser	Val	Ala	Arg	Val	Leu	Gly	Leu	Cys	Ala	Phe	Val	
	GCG	GCC	GTG	AGC	CTC	CTG	CCG	GCC	GTG	AGC	ATG	GAG	CGC	TGC	GCG	TCT	G	3'	
	Ala	Gly	Val	Ser	Leu	Leu	Pro	Ala	Val	Ser	Met	Glu	Arg	Cys	Ala	Ser			

43 / 79

PCT/JP95/01599

FIGURE 44

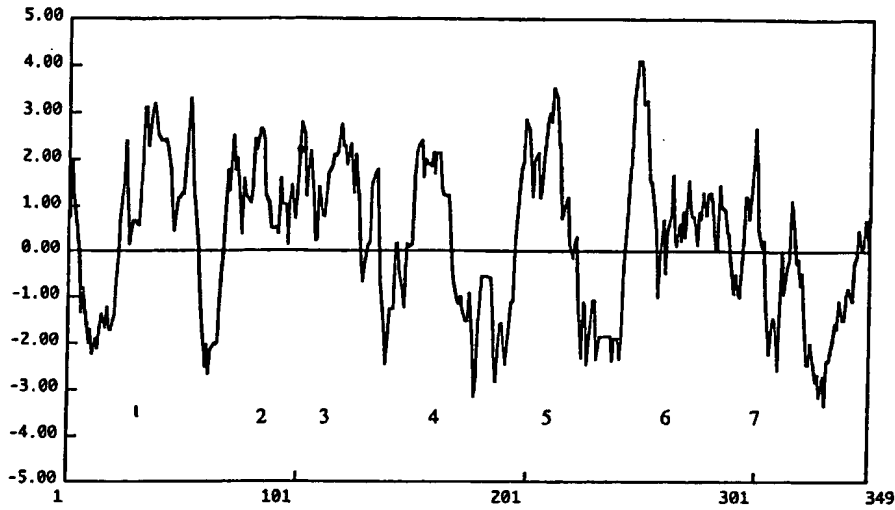


44 / 79

WO 96/05302

PCT/JP95/01599

FIGURE 47



47 / 9

PCT/JP95/01599

FIGURE 48

MOUSEGALRECE	1	10	20	30	40	50	
HUMGALAMI	1	MELAMNLSE	GNCEPPEPPA	PEERPLFGIG	VENETLWVF	GHIFAMSVLC	50
	1	MELAVGNLSE	GNCEPPEPPA	PEERPLFGIG	VENETLWVF	GHIFAMSVLC	50
MOUSEGALRECE	51	60	70	80	90	100	
HUMGALAMI	51	NSLVITVLAR	SKFGKPRSTT	ILFILNLSTF	DLAYLLFCIF	FOATVVALPT	100
	51	NSLVITVLAR	SKFGKPRSTT	ILFILNLSTF	DLAYLLFCIF	FOATVVALPT	100
MOUSEGALRECE	101	110	120	130	140	150	
HUMGALAMI	101	VLGAFICKF	IHYFFTVSMI	/SIFTLAAMS	/DRYVAIVHS	RRSSSLRVSE	150
	101	VLGAFICKF	IHYFFTVSMI	/SIFTLAAMS	/DRYVAIVHS	RRSSSLRVSE	150
MOUSEGALRECE	151	160	170	180	190	200	
HUMGALAMI	151	JALLGVSTN	ALSTAFASPV	AYHCRLEFH	DENQTECHEC	JENKLEKRAY	200
	151	JALLGVSTN	ALSTAFASPV	AYHCRLEFH	DENQTECHEC	JENKLEKRAY	200
MOUSEGALRECE	201	210	220	230	240	250	
HUMGALAMI	201	VCIEYFCYL	PLLLICFCY	AKVLTHLIRK	LKKHKKKSEA	SKKTAQTVL	250
	201	VCIEYFCYL	PLLLICFCY	AKVLTHLIRK	LKKHKKKSEA	SKKTAQTVL	250
MOUSEGALRECE	251	260	270	280	290	300	
HUMGALAMI	251	VVVVFGISL	LPHHWVLLRA	EECAFPPLTPA	SECFRITAHG	LAYSNSSVNP	300
	251	VVVVFGISL	LPHHWVLLRA	EECAFPPLTPA	SECFRITAHG	LAYSNSSVNP	300
MOUSEGALRECE	301	310	320	330	340	350	
HUMGALAMI	301	IYAFISENF	RKAYKOVFKC	VCDESPRSE	KRENKSRDT	PPSINCTHVX	350
	301	IYAFISENF	RKAYKOVFKC	VCDESPRSE	KRENKSRDT	PPSINCTHVX	350
MOUSEGALRECE	351	360	370	380	390	400	
HUMGALAMI	351	X.....	400

48 / 9

WO 96/03502

PCT/JP95/01599

FIGURE 51

pmj10	1	LT	LHPVSS	KHRTSHWASR	VLSVLSAT	AFSVPLVLF	EPYDOR-K-G	50
B42009	1	ICVLHPVAC	HRTVSLAMK	VIVGPHILAI	VLTLPLVFL	TTVTI	N-N	50
JC2014	1	VCVLHPVAC	HRTVSLAKK	VITGPMAL	LTLPLVITRV	TVPGK-T-		50
A46520	1	ICVLHPVAC	HRNVSLAKK	VIVGPHICAL	LTLPLVITRV	TLSH	RAP	50
A46525	1	LT-KH	KVGTGLAM	ACGVALAI	LTITSFVY	EAYKD	FYS-E	50
S28787	1	AIVPATNS	KPKLLAEV	VIVGPHILAI	LTITSFVY	DI	EV-D-E	50
pmj10	51	VTNRNIVAV	STDESKEMO	TTRAMIATC	FISRFILSFL	LPFLVIGFCY		100
B42009	51	DYCTIFPMS	GG-TDEERL	QVATTLIAF	GIIRFVIGFS	LPMSIVAICY		100
JC2014	51	TVACTIFPMS	WTN-DEKEEL	QVAVALLIVE	GIIRFVIGFS	APMSIVAVS		100
A46520	51	KMACTFDWS	WTE-DPAEKL	QVATSFMMZH	GIIRFVIGFS	TPMSIVAVCY		100
A46525	51	HVAGITGG	GS--FKEKA	VA-----	LTLMVGV	LHLLTLNICV		100
S28787	51	VTDRFAP	SDL-----	VVFQ-----	IVVGLL	EGIVILSCY		100
pmj10	101	ERVAR	MKE	GLKSSKFEK	ITTAVI			150
B42009	101	GLIAKIHKK	GLIKSSRPLE	ITTAVI				150
JC2014	101	GLIAKIHKK	GLIKSSRPLE	ITTAVI				150
A46520	101	GLIAKIHKK	GLIKSSRPLE	ITTAVI				150
A46525	101	TFLLLRWS	KATREKTK	ITTAVI				150
S28787	101	CTISLSHS	KGQQRKA	ITVILIL				150

51 / 79

FIGURE 52

5	CTG	ACT	GCT	CTG	GGG	ACT	GAC	GGG	TAT	TTC	MAG	ATT	GTC	MAG	CCC	CTT	TCC	AGC	54
63	TTC	TTC	ATC	CAG	TCT	GTC	AGC	TAC	ACC	AAA	CTC	GTC	TGC	CTG	GTC	TGC	TTC	AGC	54
72	Ser	Phe	Ile	Gln	Ser	Val	Asn	Tyr	Ser	Lys	Leu	Val	Ser	Leu	Val	Val	Trp	Leu	108
117	CTC	ATG	CTC	CTC	GCC	GTC	CCC	ACC	GTC	ATT	CTC	ACC	ACC	AGA	GTT	AGC			162
126	Leu	Met	Leu	Leu	Leu	Ala	Val	Pro	Asn	Val	Ile	Leu	Thr	Asn	Gln	Arg	Val	Lys	162
180	GAC	GTC	ACT	CAG	ATA	AAA	TCC	ATG	GAA	CTT	AAA	ACC	GAA	CTG	CCC	CAG	TGC		216
189	ASP	Val	Thr	Gln	Ile	Lys	Cys	Met	Glu	Leu	Lys	Asn	Glu	Leu	Gly	Arg	Gln	Trp	216
225	CAC	AGC	GCG	TCA	ACC	TAC	ATC	TTT	GTC	GGC	ATT	TTC	TGC	CTT	GTC	TTC	TTG		270
234	Hls	Lys	Ala	Ser	Asn	Tyr	Ile	Phe	Val	Gly	Ile	Phe	Trp	Leu	Val	Phe	Leu	Leu	270
288	CTA	ATC	ATT	TTC	TAC	ACT	GCT	ATC	ACC	AGG	AAA	ATC	TTT	MAG	TTC	CAC	CTG	AAA	324
297	Leu	Ile	Ile	Phe	Tyr	Thr	Ala	Ile	Thr	Arg	Lys	Ile	Phe	Lys	Ser	Hls	Leu	Lys	324
342	TCC	AGA	MAG	MAI	TCC	ATC	TGC	GTC	AAA	MAG	AAA	TCT	AGC	CCC	ACC	ATC	TTC	AGC	378
351	Ser	Arg	Lys	Asn	Ser	Ile	Ser	Val	Lys	Lys	Ser	Ser	Arg	Asn	Ile	Phe	Ser		378
396	ATC	GTC	TTT	ATC	CTC	TGT	TGC	CCC	CCC	TAC	CAC	ATC	3						414
405	Ile	Val																	414

52 / 79

54/79

[illegible]

5 5 / 7 9

FIGURE 55

5' GCC ACC AAC GTG TTC ATC CTG TGT CTG GTG GAC CTG GCT GCT GCG ACC CTC
 9 18 27 36 45 54
 Vol Asp Leu Leu Ala Leu Thr Leu
 63 72 81 90 99 108
 ATG CCT CTG GGC ATG TTC TCC AGC TCC GGC CTC TTT GAC CAC GGC CTC TTT GGG
 Met Pro Leu Ala Met Leu Ser Ser Ser Ala Leu Phe Asp His Ala Leu Phe Gly
 117 126 135 144 153 162
 GAG GTG GGC TGC CCG CTC TAC TTG TTC CTG AGC GTC TGC TTT GTC ACC CTG GGC
 Glu Val Ala Cys Arg Leu Tyr Phe Leu Ser Val Cys Phe Val Ser Leu Ala
 171 180 189 198 207 216
 ATC CTC TCG GTG TCC GGC ATC AAT GTG GAG CCG TAC TAT TAT GTG GTC CAC CCC
 Ile Leu Ser Val Ser Ala Ile Asn Val Glu Arg Tyr Tyr Val Val His Pro
 225 234 243 252 261 270
 ATG CCG TAT GAG GTG CCG ATG AAA CTG GGG CTG GTC GGC TCT GTG CTG GTC GGC
 Met Arg Tyr Glu Val Arg Met Lys Leu Gly Leu Val Ala Ser Val Leu Val Gly
 279 288 297 306 315 324
 GTG TGG GTG AAG GGC CTG GGC ATG GCT TCT GTG CCA GTG TTG GGA AAG GTG TCC
 Val Trp Val Lys Ala Leu Ala Met Ala Ser Val Pro Val Leu Gly Arg Val Ser
 333 342 351 360 369 378
 TGG GAG GAA GGC CCT CCC AGT GTC CCC CCA GGC TGT TCA CTC CAA TGG AGC CAC
 Trp Glu Glu Gly Pro Pro Ser Val Pro Pro Gly Cys Ser Leu Gln Trp Ser His
 387 396 405 414 423 432
 AGT GGC TAC TGC CAG CTT TTC GTG GTG GTC TTC GGC GTC CTC TAC TTC CTG CTG
 Ser Ala Tyr Cys Gln Leu Phe Val Val Phe Ala Val Leu Tyr Phe Leu Leu
 441 450 459 468 477 486
 CCC CTG CTC CTC ATC CTT GTG GTC TAC TGC AGC ATG TTC CCG GTG GCT CGT GTG
 Pro Leu Leu Leu Ile Leu Val Val Tyr Cys Ser Met Phe Arg Val Ala Arg Val
 495 504 513 522 531 540
 GGT GGC ATG CAG CAC GGG CCG CTG CCC ACG TGG ATG GAG ACG CCC CGG CAA CCG
 Ala Ala Met Gln His Gly Pro Leu Pro Thr Trp Met Glu Thr Pro Arg Gln Arg

5 6 / 7 9

FIGURE 56

549 558 567 576 585 594
 TCC GAG TCT CTC AGC AGC CCG TCC ACT ATG GTC ACC AGC TCG GGG GGC CCG CAG
 Ser Glu Ser Leu Ser Ser Arg Ser Thr Met Val Thr Ser Ser Gly Ala Pro Gln
 603 612 621 630 639 648
 ACC ACC CTT CAC CCG ACG TTT GGC GGA GAG GCA GCA GTC CTC CTC GCT
 Thr Thr His Arg Thr Phe Gly Gly Gly Lys Ala Ala Val Val Leu Leu Ala
 657 666 675 684 693 702
 GTG GGA GGA CAG TTC CTC TGT TGG TTG CCC TAC TTC TCC TTC CAC CTC TAT
 Val Gly Gly Gln Phe Leu Leu Cys Trp Leu Pro Tyr Phe Ser Phe His Leu Tyr
 711 720 729 738 747 756
 GTG GGC CTG AGC GCT CAG CCC ATT GCA GCG GGG CAG GTG GAG AAC GTG GTC ACC
 Val Ala Leu Ser Ala Gln Pro Ile Ala Ala Gly Gln Val Glu Asn Val Val Thr
 765 774 783 792 801 810
 TGG ATT GGC TAC TTC TGC ACC TCC AAC CCT CTC CTC TAT TCC TTC CTC CCT 3'
 Trp Ile Gly Tyr Phe Cys Phe Thr Ser

5 8 / 7 9

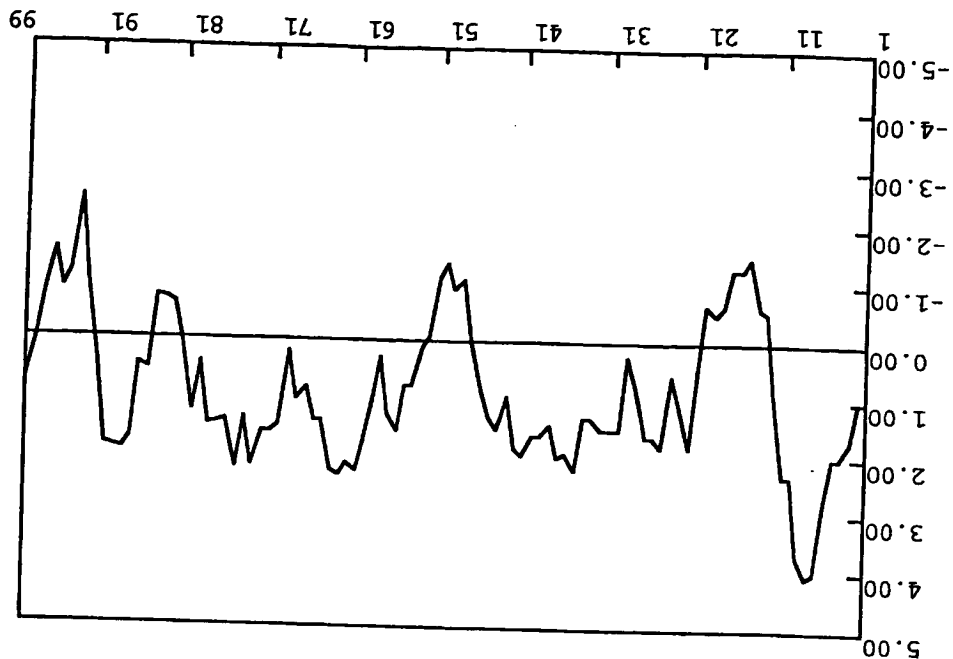


FIGURE 58

5 7 / 7 9

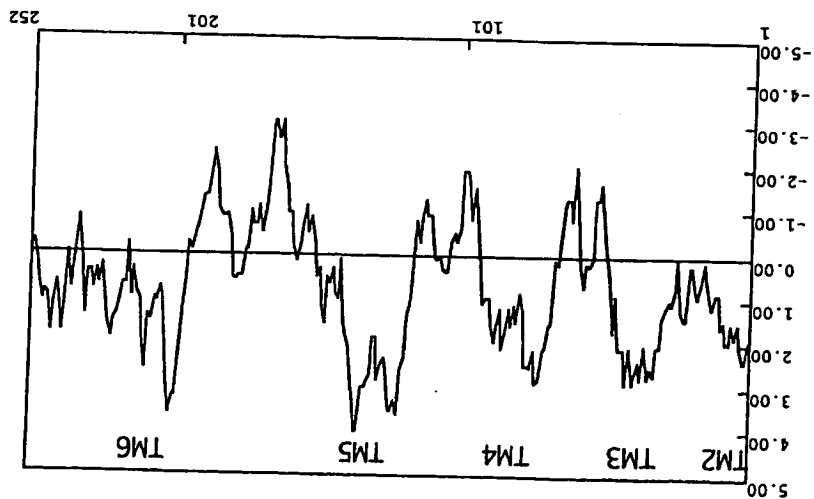
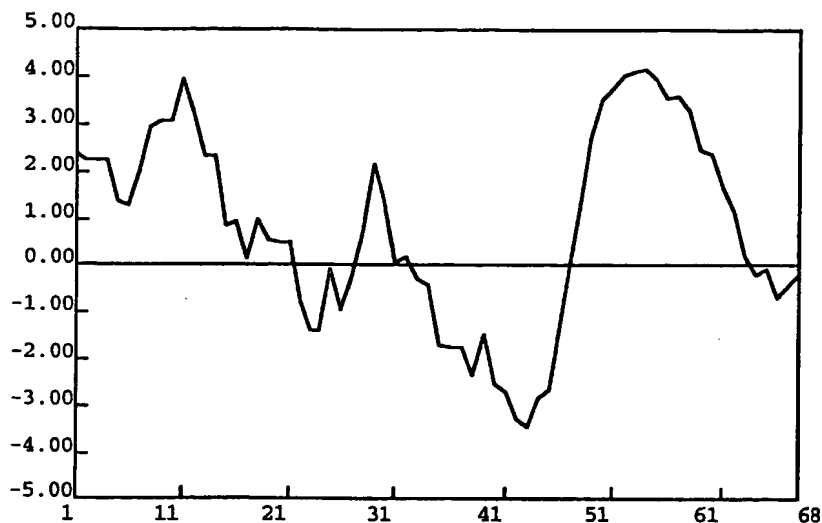


FIGURE 57

FIGURE 59



59 / 79

FIGURE 60

p19P2 S12863	1	10	20	30	40	50	
	1	VGMVENVLLV	LVIAVRRLH	NVINSFLICNL	ALSDVIMCTA	QVELILAZAF	50
	1	LGVSGLALII	IIILKQKEMR	NVTNILLIVNL	SFSDLLVAVM	GLEFFIVNIL	50
p19P2 S12863	51	60	70	80	90	100	
	51	EPRGTVFCGG	LCHVFFLELP	VIVYVSVFTE	TTIAVDRYVV	LVHFLRRRI-	100
	51	MDH-VVFGET	MCKENPEVOC	VSITVSIESL	VLIAVERHQL	IINERGWRPN	100
p19P2 S12863	101	110	120	130	140	150	
	101	NRHAYIGITV	IWVLAVASSL	PFVIYQILTD	EPFQNVSLAA	FKDKYVCFDK	150
							150
p19P2 S12863	151	160	170	180	190	200	
	151	GLLV	TYMLPLVIL	LS-----	VRVSVK	VVPGCVTQSQ	200
	151	FPSDSHRLSY	ITILLVLOYF	GPLCFIFICV	FKIYIR	NNMDKIRDS	200
p19P2 S12863	201	210	220	230	240	250	
	201	ADWDRARRRR	TFCLLVVVV	VFAICVLEFY	250
	201	KYRSSETKRI	NVMLLSIIVA	-FAVCHLELT	250

60 / 79

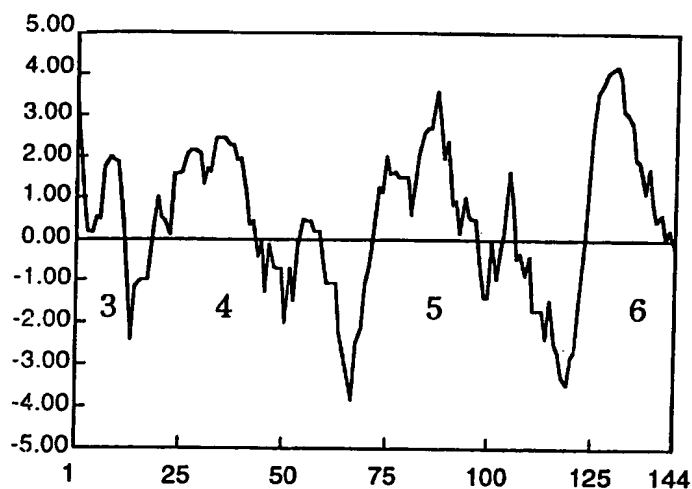
FIGURE 63

p19P2	1	10	20	30	40	50	
pG3-2/pG1-10	1	VGMVGNILLV	LVIAVRRLH	NVTNELLIGNI	ALSDVLMCTA	CVPLTLAYAF	50
p5S38	-79	VGMVGNILLV	LVIAVRRLH	NVTNELLIGNI	ALSDVLMCTA	CVPLTLAYAF	50
							-30
p19P2	51	60	70	80	90	100	
pG3-2/pG1-10	51	EPFGWVFGGG	LCHLVFFLQP	VIVYVSVEFI	ITIAVDRYV	LVHPLRRRI	100
p5S38	-29	EPFGWVFGGG	LCHLVFFLQA	VIVYVSVEFI	ITIAVDRYV	LVHPLRRRI	100
							21
p19P2	101	110	120	130	140	150	
pG3-2/pG1-10	101	LRLSAYAVIA	IVVLSAVLAI	PAVHTYHVE	LKPHDVRLCE	EFWGSQERQF	150
p5S38	22	LRLSAYAVIG	IVVLSAVLAI	PAVHTYHVE	LKPHDVSLCE	EFWGSQERQR	150
							71
p19P2	151	160	170	180	190	200	
pG3-2/pG1-10	151	GLLL	FYLLPLLVI	LSYVRVSVKI	RNRVVFQCVI	QSQADNDRAF	200
p5S38	72	GLYAKGLLL	FYLLPLLVI	LSYVRVSVKI	RNRVVFQCVI	QSQADNDRAF	200
							121
p19P2	201	210	220	230	240	250	
pG3-2/pG1-10	201	RRRTFCLLV	VVVFAICHL	PFF			250
p5S38	122	RRRTFCLLV	VVVFTLCWL	PFF			250
							171

6 3 / 7 9

PCT/JP95/01599

FIGURE 64



6 4 / 7 9

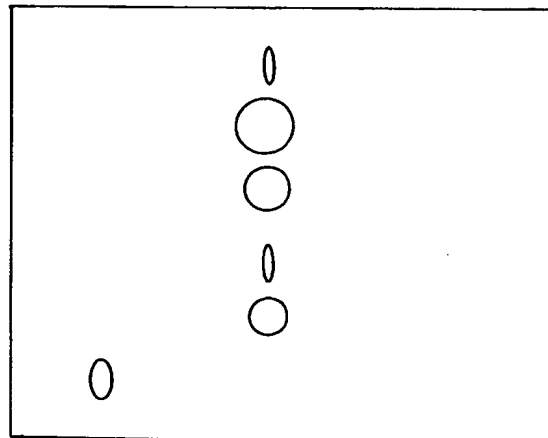
W/O 96/05302

PCT/JP95/01599

6 5 / 7 9

FIGURE 65

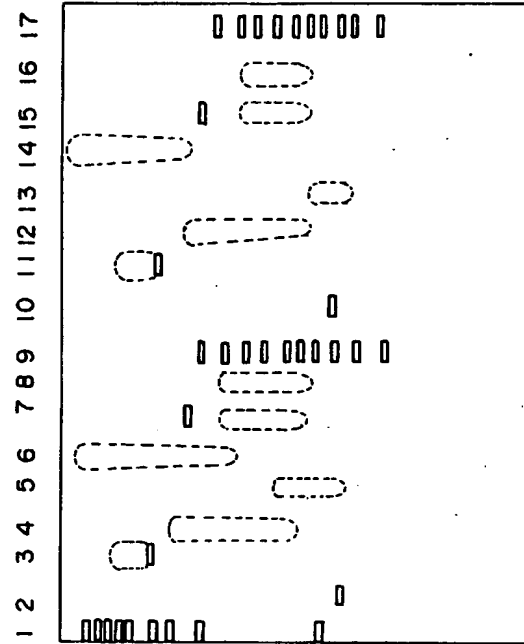
Neuro-2a
BRAIN
THYMUS
SPLEEN
PANCREAS



49.5
47.5
44.4
42.4
41.4
40.2

6 6 / 7 9

FIGURE 66



67 / 79

FIGURE 67

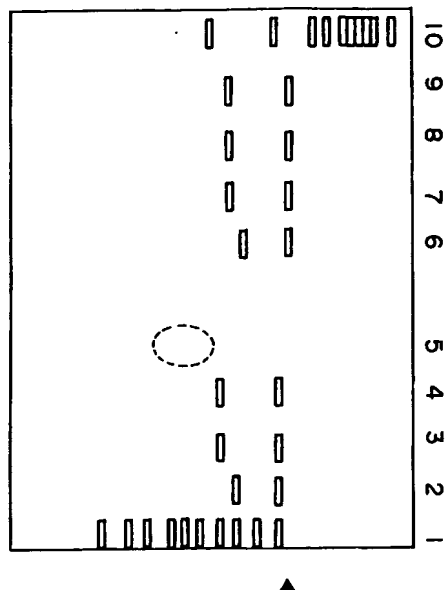
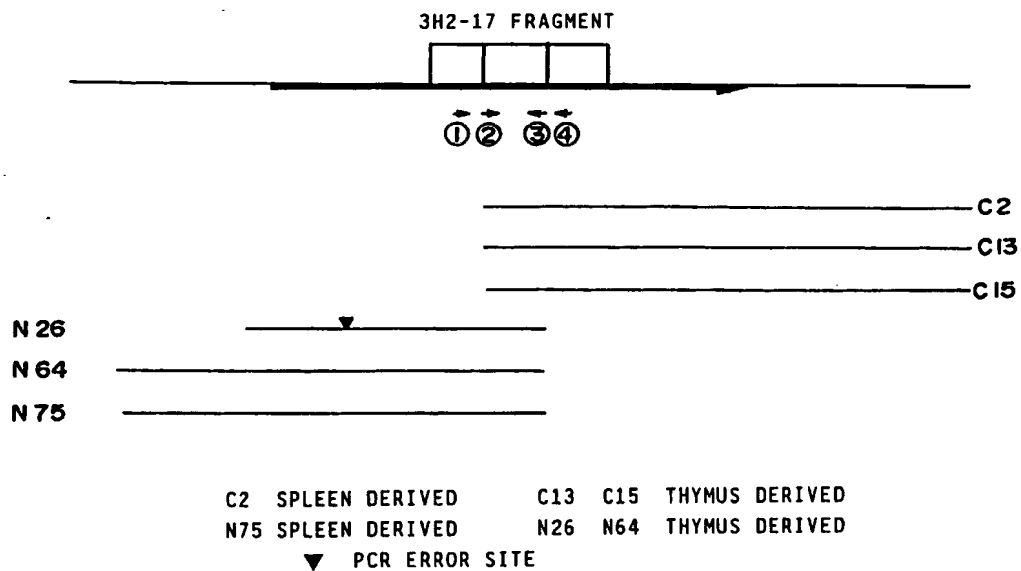


FIGURE 68



68 / 79

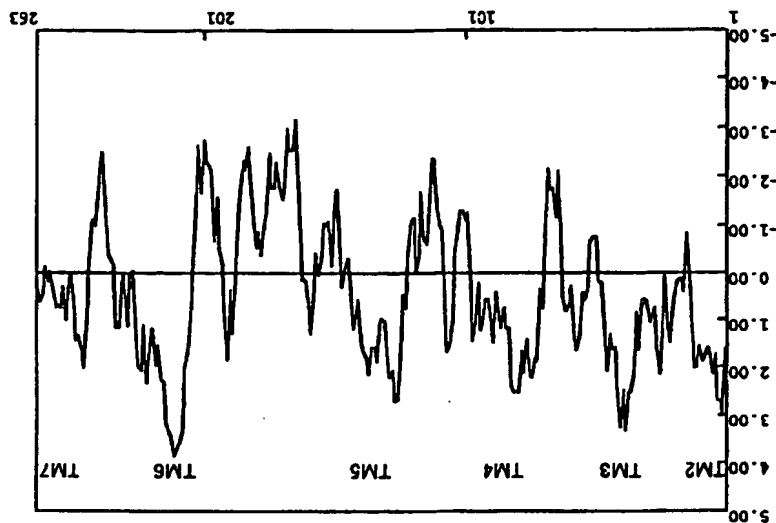
7 4 / 7 9

7 3 / 7 9

FIGURE 73

Gly	Arg	Gln	Ile	Pro	Gly	Thr	Thr	Ser	Ala	Leu	Val	Arg	Asn	Trp	Lys	Arg	Pro
549					558		567		576		585						
TCA	GAC	CAG	CTG	GAC	GAC	CAG	GCC	CAG	CTG	AGC	TCA	GAG	CCC	CAG	CCC	CAG	CGG
Ser	Asp	Gln	Leu	Asp	Gln	Gly	Gln	Gly	Leu	Ser	Ser	Glu	Pro	Gln	Pro	Arg	
603					612		621		630		639						
GCC	CGC	GCC	TTC	CTG	GCC	GAG	GTG	AAA	CAG	ATG	CGA	GCC	CGG	AGG	AAG	ACG	GCC
Ala	Arg	Ala	Phe	Leu	Ala	Glu	Val	Lys	Gln	Met	Arg	Ala	Arg	Arg	Lys	Thr	Ala
657					666		675		684		693						
AAG	ATG	CTG	ATG	GTG	GTG	CTG	CTG	GTC	TTC	GCC	CTC	TGC	TAC	CTG	CCC	ATC	AGT
Lys	Met	Leu	Met	Val	Val	Leu	Leu	Val	Phe	Ala	Leu	Cys	Tyr	Leu	Pro	Ile	Ser
711					720		729		738		747						
GTC	CTC	AAC	GTC	CTC	AAG	AGG	GTC	TTC	GGG	ATG	TTC	CGC	CAA	GCC	AGC	GAC	CGA
Val	Leu	Asn	Val	Leu	Lys	Arg	Val	Phe	Gly	Met	Phe	Arg	Gln	Ala	Ser	Asp	Arg
765					774		783		792		801						
GAG	GCC	ATC	TAC	GCC	TGC	TTC	ACC	TTC	TCC	CAC	TGG	CTG	GTG	TAC	GCC	AAC	AGC
Glu	Ala	Ile	Tyr	Ala	Cys	Phe	Thr	Phe	Ser	His	Trp	Leu	Val	Tyr	Ala	Asn	Ser
819					828		837										
GCC	GCC	AAT	CCC	CTC	CTC	TAC	TCC	TTC	CTC	CTC	CTC	CTC	CTC	CTC	CTC	CTC	CTC
Ala	Ala																

FIGURE 74



7 5 / 7 9

FIGURE 75

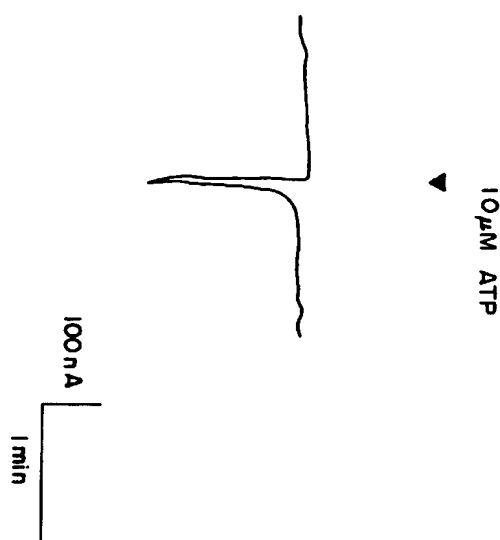


FIGURE 76

	10	20	30	40	50	
h3H2-17(5-3)	1					50
p3H2-17(5')	1	GTGGGCTGG	TGGGCAACAT	CCTGGCTTCC	TGGCACAAGC	GTGGAGGTCG
	60	70	80	90	100	50
h3H2-17(5-3)	51					100
p3H2-17(5')	51	CCGTGCTGCT	TGGGTAGTGT	GTGGAGTGGT	GTGGCTGGCT	GTGGAGGTCG
	110	120	130	140	150	100
h3H2-17(5-3)	101					150
p3H2-17(5')	101	AGTGGCTGGC	PAGAGGCATG	TCGCTGGTGC	CAGGCATGCA	GGTATAGGCG
	160	170	180	190	200	150
h3H2-17(5-3)	151					200
p3H2-17(5')	151	AGTGGCTGGC	TCGCTGGTGC	CAGGCATGCA	GGTATAGGCG	GGTATAGGCG
	210	220	230	240	250	200
h3H2-17(5-3)	201					250
p3H2-17(5')	201	AGTGGCTGGC	TCGCTGGTGC	CAGGCATGCA	GGTATAGGCG	GGTATAGGCG
	260	270	280	290	300	250
h3H2-17(5-3)	251					300
p3H2-17(5')	251	AGTGGCTGGC	TCGCTGGTGC	CAGGCATGCA	GGTATAGGCG	GGTATAGGCG
	310	320	330	340	350	300
h3H2-17(5-3)	301					350
p3H2-17(5')	301	CCAGCAGGTC	CTGTGGCCCA	AGAGCGGCGC	AGCAAGGCGG	CTCGTATGGC
	360	370	380	390	400	350
h3H2-17(5-3)	351					400
p3H2-17(5')	351	TGTGGTGGTG	GCAGCTGTCT	TGCCCCCTTG	CTGGCTGGCT	CTCTAC....

7 6 / 7 9

7 7 / 7 9

FIGURE 77

1 TGACTCTCTGACATAGAGAACCCACCTGGGACGATGGATGGACATGGACACGGC 60
 1 NetGluTrpAspSerGlyThrGly 8
 61 CAGGCTCTGGGCTTCACACACACCTCTGTCTACGGGAGCACTTCAAGCACTGCTG 120
 8 GluAlaLeuGlyLeuProThrCysValTyrArgIuAsnPhelYsgInLeu 28
 121 CTGCACTCTGTATCTGGGGCTCTGGGGCTGGCTTGGCTTGGCTTGGCAATCTGTGAT 180
 28 LeuProValTyrSerAlaValAlaAlaLeuThrArgThrAlaValTyrThrLeuAsn 48
 181 ACCGATCTGCACCTCTGGGGCTTCAGCCCTGACCCGACGGCTGTACACCCCAACCTT 240
 48 ThrGlnIleCysThrSerArgAlaLeuThrArgThrAlaValTyrThrLeuAsn 68
 241 CTCTGGCTGACCTCTATATGCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCT 300
 68 AlaLeuAlaLeuLeuLeuLeuLeuLeuLeuLeuLeuLeuLeuLeuLeuLeuLeu 88
 301 GGTATCTCTGCT 360
 88 GlyAlaIleIleIleIleIleIleIleIleIleIleIleIleIleIleIleIle 108
 361 ACCTGCAACGACGACATCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCT 420
 108 AsnLeuIleIleIleIleIleIleIleIleIleIleIleIleIleIleIleIle 128
 421 TCCACCT 480
 128 CysHisProLeuAlaProIleIleIleIleIleIleIleIleIleIleIleIleIle 148
 481 GTACCT 540
 148 ValThrValTrpLeuAlaValThrGlnCysLeuProIleIleIleIleIleIle 168
 541 GGCATCT 600
 168 GlyIleGlnArgAsnArgThrValCysTyrAspLeuSerProIleAlaLeuThrHis 188
 601 TATATCT 660
 188 TyrMetProTyrGlyMetAlaLeuThrValIleGlyPheLeuLeuProPheAlaLeu 208
 661 CTGGCT 720
 208 LeuAlaCysTyrCysLeuLeuAlaCysArgLeuCysArgGlnAspGlyProIleGlnPro 228
 721 GTGGCT 780
 228 ValAlaGlnIleArgGlyIleAlaLeuArgGlnAlaValValAlaAlaAlaPhe 248
 781 GGCATCT 840
 248 AlaIleSerPheLeuProPheHisIleThrThrThrThrThrThrThrThrThr 268
 841 CCGGGCT 900
 268 ProGlyValProCysThrValLeuGlnAlaPheAlaLeuIleTyrIleGlyThrArgPro 288
 901 TTTCCT 960
 288 PheAlaSerAlaAsnSerValLeuAspProIleLeuPheThrThrThrThrThrThr 308
 961 CCGGGCT 1020
 308 ArgArgProIleIleGlnLeuGlnIleIleIleIleIleIleIleIleIleIleIleIle 328
 1021 TGA
 328 ***

7 8 / 7 9

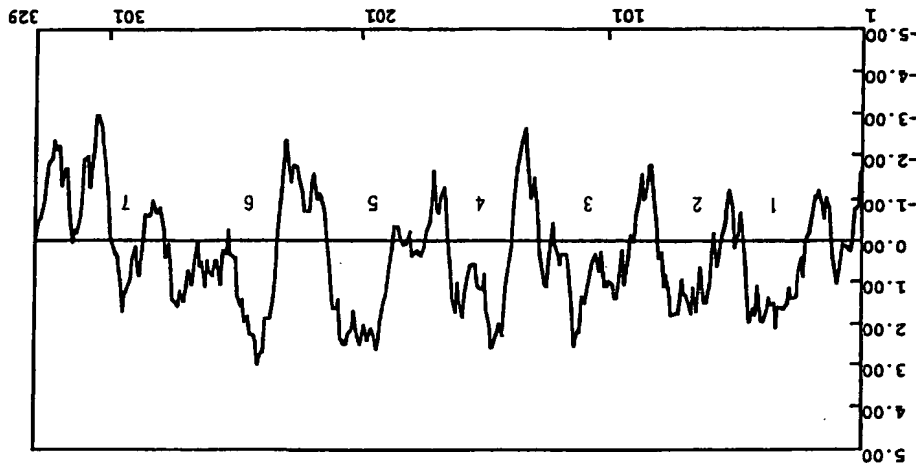


FIGURE 78

INTERNATIONAL SEARCH REPORT

Inventor: Application No. PCT/JP 95/01599

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/12 CO/K14/705 C12Q1/68

B. FIELDS SEARCHED
Minimum documentation searched (classification system followed by classification symbols)
IPC 6 CO/K C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Character of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	MOL. CELL. ENDOCRINOLOGY, vol. 5, January 1991, pages 1331-1338, M.T. HARRIGAN ET AL., 'Identification of a gene induced by glucocorticoids in murine T-cells: a potential G protein-coupled receptor' see the whole document	4-6, 10, 11, 13
Y	WO. A. 92 01810 (LENNER MICHAEL R; LERNER ETHAN A (US)) 6 February 1992 see abstract; claims 1-17	14-18
A	EP. A. 0 578 962 (AMERICAN CYANAMID CO) 19 January 1994 see example 2	1-3
	--- -/-	

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Patent family members are listed in annex.

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E* earlier document but published on or after the international filing date
L* documents which may throw doubt on priority claim(s) or which are cited to establish the publication date of another claim or other special reason (as specified)
O* document referring to an oral disclosure, use, exhibition or other event
P* document published prior to the international filing date but later than the priority date claimed

Date of the actual completion of the international search

Date of mailing of the international search report

18 December 1995

05.01.96

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Fax (+31) 79 460 3010

Authorized officer
Gurdjian, D

FIGURE 79

human prino, mouseFULL3H2	1	10	20	30	40	50	50
	1	10	20	30	40	50	50
human prino, mouseFULL3H2	51	60	70	80	90	100	100
	51	60	70	80	90	100	100
human prino, mouseFULL3H2	101	110	120	130	140	150	150
	101	110	120	130	140	150	150
human prino, mouseFULL3H2	151	160	170	180	190	200	200
	151	160	170	180	190	200	200
human prino, mouseFULL3H2	201	210	220	230	240	250	250
	201	210	220	230	240	250	250
human prino, mouseFULL3H2	251	260	270	280	290	300	300
	251	260	270	280	290	300	300
human prino, mouseFULL3H2	301	310	320	330	340	350	350
	301	310	320	330	340	350	350

7 9 / 7 9

INTERNATIONAL SEARCH REPORT

<div> <div>Intern</div> <div>al Application No</div> <div>PCT/JP 95/01599</div> </div>	
<div> <div>C.(Continuation)</div> <div>DOCUMENTS CONSIDERED TO BE RELEVANT</div> </div>	
Category *	<div> <div>Citation of document, with indication, where appropriate, of the relevant passages</div> <div>Relevant to claim No.</div> </div>
A	<div> <div> <div>SCIENCE,</div> <div>vol. 244, 1989</div> <div>pages 569-571,</div> <div>LIBERT F. ET AL. 'selective amplification</div> <div>and cloning of four new members of the G</div> <div>protein-coupled receptor family'</div> <div>cited in the application</div> <div>see the whole document</div> <div>-----</div> </div> <div>1-3</div> </div>

INTERNATIONAL SEARCH REPORT

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<div> <div>Information on patent family members</div> </div>	
Patent document cited in search report	<div> <div>Publication date</div> <div>Patent family member(s)</div> <div>Publication date</div> </div>
WO-A-9201810	<div> <div>06-02-92</div> <div>US-A- 5462856</div> <div>31-10-95</div> </div>
EP-A-0578962	<div> <div>19-01-94</div> <div>EP-A- 0539518</div> <div>05-05-93</div> </div>
	<div> <div></div> <div>JP-T- 6502757</div> <div>31-03-94</div> </div>
	<div> <div></div> <div>AU-B- 4203493</div> <div>20-01-94</div> </div>
	<div> <div></div> <div>CA-A- 2100616</div> <div>18-01-94</div> </div>
	<div> <div></div> <div>JP-A- 6153954</div> <div>03-06-94</div> </div>